

**What is the role of the asymptomatic carrier in environmental contamination with *C. difficile* spores and nosocomial transmission, and can we do anything to prevent it?**

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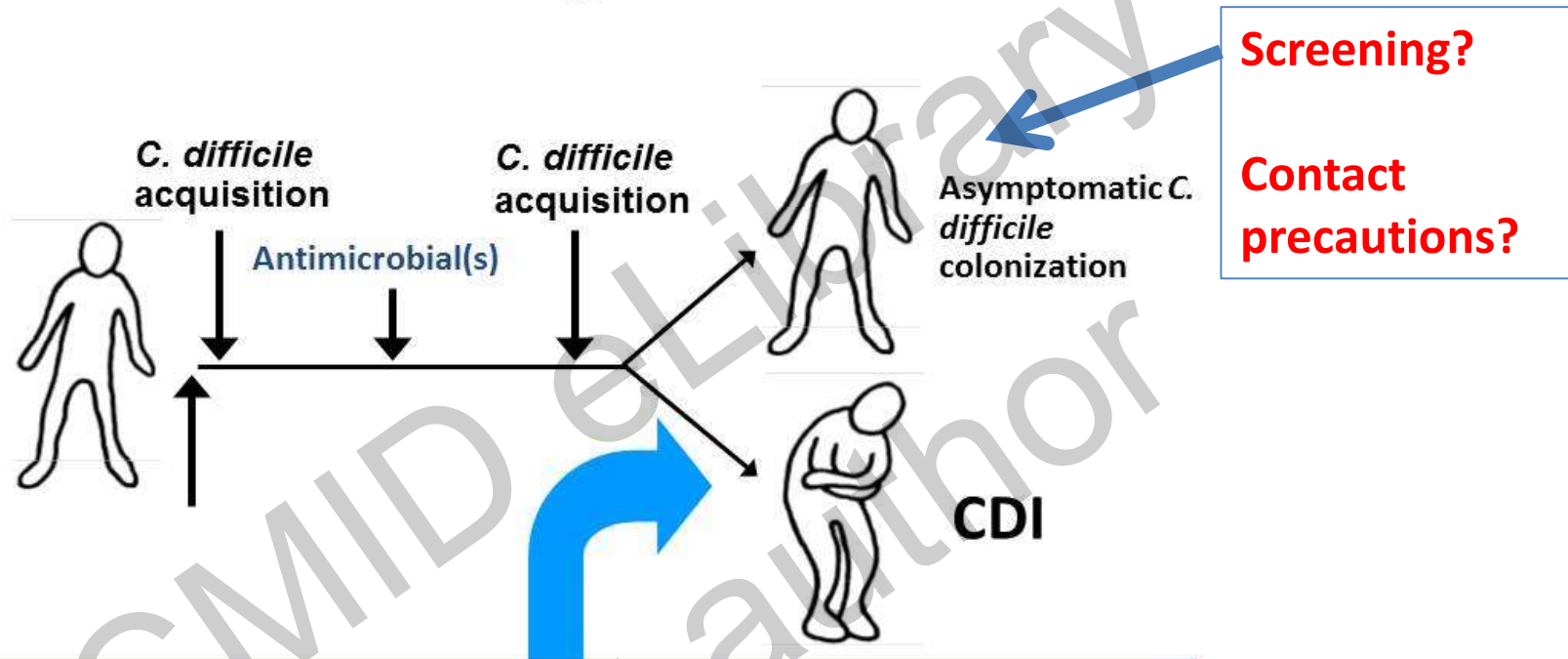
# Overview of session

1. Introduction to asymptomatic carriage
2. Studies of asymptomatic carriers
3. Extant guidance 2018
4. Conclusions

# 1. Introduction to asymptomatic carriage

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# Figure 1. Current Pathogenesis Model for CDI



Acquisition of a toxigenic strain of *C. difficile* and failure to mount an anamnestic toxin A IgG antibody response results in CDI.

# Asymptomatic carriage of *C. difficile*

- Some people carry *C. difficile* in their gut without having any symptoms
- Asymptomatic carriage upon hospitalisation was **8.1%** in pooled meta-analysis (2005-2014); **3-26%** among acute inpatients, **5-7%** among elderly in long-term care facilities.
- For adults with no recent healthcare exposure colonisation is **2%**
- There is uncertainty with regards to the management of asymptomatic carriers (incl. screening and control of spread of spores from carriers)

## 2. Studies of asymptomatic carriers

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# Risk factors for asymptomatic carriage

**Table II**

Characteristics associated with rectal carriage of *Clostridium difficile* in hospitalized patients with no symptoms of *C. difficile* infection

Factor	Unadjusted		Adjusted <sup>a</sup>	
	OR	95% CI	OR	95% CI
Used antibiotics in the previous 3 months	6.16	1.36–27.9	4.49	0.95–21.3
<i>C. difficile</i> infection (CDI) in previous 3 months	5.95	1.49–23.7	3.14	0.70–14.0
Length of stay >14 days	3.68	1.33–10.1	1.99	0.64–6.12

OR, odds ratio; CI, confidence interval.

<sup>a</sup> For other factors in Table I.

Guerrero et al, 2013

- Risk factors for asymptomatic colonisation have been associated with previous antibiotic exposure, healthcare exposure and previous CDI.
- Colonisation rates on admission to hospital :
  - Acute hospital: 4%-21% (TCD)
  - Acute geriatric hospital: 2% (TCD)

# Ribotype distributions in carriers vs CDI cases

Overall 15% of patients were colonised with TCD on admission

**Table 4. Five Most Common Toxigenic *Clostridium difficile* Strains From Asymptomatic Carriers, Protocol A, and Protocol B**

Asymptomatic Carriers (n = 40)		Protocol A (n = 74)		Protocol B (n = 49)	
Strain <sup>a</sup>	No.	Strain <sup>a</sup>	No.	Strain <sup>a</sup>	No.
014/020 <sup>*.***</sup>	14 (35%)	027	23 (31%)	027	8 (16%)
012 <sup>*.***</sup>	10 (25%)	106/174	9 (12%)	WU42	8 (16%)
053 <sup>*</sup>	4 (10%)	014/020	7 (9%)	014/020	6 (12%)
077	3 (8%)	002	7 (9%)	001	4 (8%)
027 <sup>*.***</sup>	1 (3%)	005	4 (5%)	106/174	3 (6%)

<sup>a</sup> Strain name is the polymerase chain reaction ribotype. If the strain did not match to a ribotype, the Washington University (WU) strain number is provided. If unable to discriminate between different ribotypes, both ribotypes the strain matched to are provided.

\*  $P \leq .005$ , asymptomatic carriers compared with protocol A.

\*\*  $P < .001$ , asymptomatic carriers compared with protocol B.

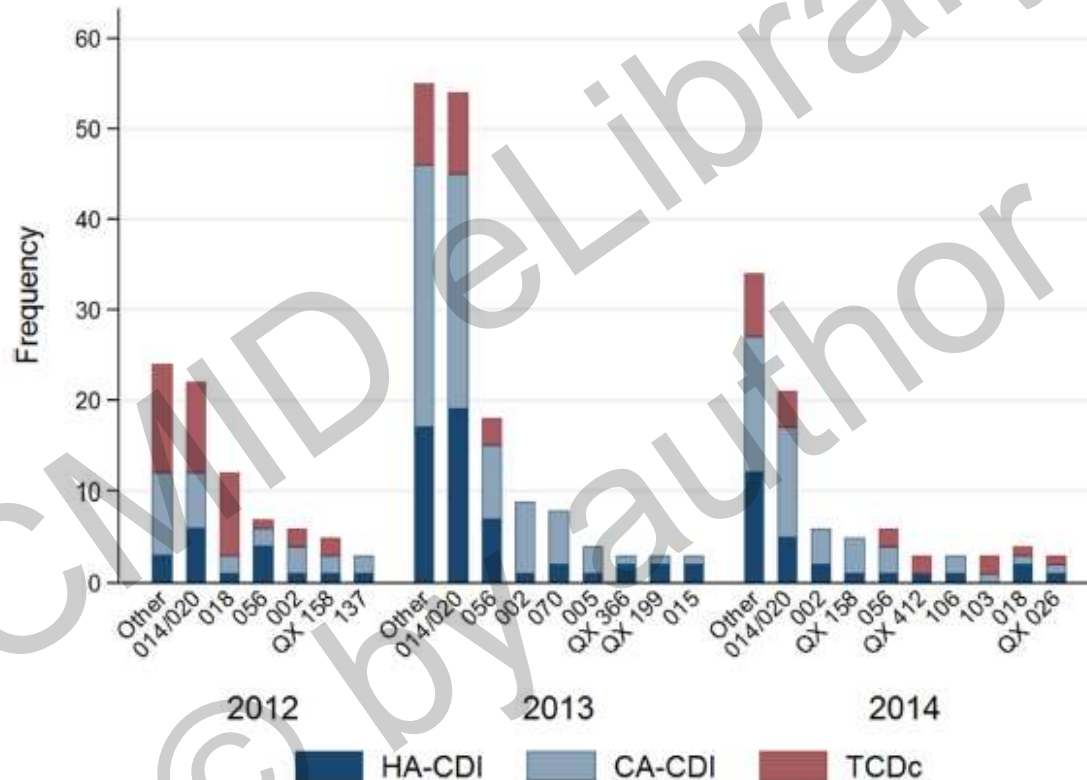
\*\*\*  $P \leq .03$ , asymptomatic carriers compared with protocol B.

Protocol A/B : identified patients with symptomatic CDI

Alasmari et al. 2014



# Ribotype distribution in hospital and community CDI cases, and carriers



**FIG 1** Distribution of ribotypes by year among symptomatic HA-CDI and CA-CDI patients and asymptomatic toxigenic *C. difficile*-colonized (TCDc) patients. Ribotypes found at a frequency of less than 3 isolates in a year were grouped into the other category.

# Development of CDI following asymptomatic colonisation (or contamination)

**Table 3** Clinical impact of *Clostridium difficile* colonization in adults

Country	Study year	Follow-up duration	Clinical impact	Refs
			<b>Development of <i>C. difficile</i> infection</b>	
USA	1983–1993	9 wk–20 mo	1.0% (2/192) CdC vs. 3.6% (22/618) no CdC	29
USA	1991	5 mo	47% (9/19) tCdC	40
Israel	1998	5 mo	51.3% (19/37) CdC at admission vs. 12.0% (28/234) no CdC ( $p < 0.0001$ )	30
Taiwan	2011	6 mo	17.9% (5/28) tCdC vs. 1.4% (2/140) no tCdC ( $p = 0.002$ )	9
Taiwan	2011–2012	18 mo	14.1% (11/79) tCdC vs. 0.9% (3/328) no CdC, 0% (0/34) ntCdC ( $p < 0.001$ )	8
			<b>Contamination</b>	
USA	2008	90 d	17% (3/18) CdC: skin and/or environmental contamination	42
USA	2006	3 mo	Carriers vs. noncarriers: skin (61% vs. 19%; $p = 0.001$ ) or environmental contamination (59% vs. 24%; $p = 0.004$ )	23

CdC = *C. difficile* colonization; ntCdC = nontoxigenic CdC; tCdC = toxigenic CdC.

# Protective effect of *C. difficile* colonisation

- Incubation period 2-3 days (median), but one study showed that 7% had been colonised > 1 week before developing CDI (Curry et al, 2013)
- Earlier studies showed that being colonised over longer time periods has a **protective effect against CDI** (Kyne et al, 2000, Shim et al 1998)
- It is possible that risk of **progression** from colonisation to infection **decreases over time due to boosting of serum antibodies against toxin A and B** (Kyne et al, 2001)
- Colonisation with non-toxigenic strains may be protective against symptomatic CDI due to competition for nutrients or access to the mucosal surface (Shim et al. 1998, Sambol et al. 2001)

# Effect of antibiotics on progression from colonised to infected states

- The **individual risk of symptomatic CDI** was found to be higher in patients admitted to a room **where a previous patient without CDI was administered antibiotics**, suggesting induced shedding of *C. difficile* from asymptomatic carriers (Brown et al. 2015)
- Ward-level antibiotic abx prescribing** (among CDI and non-CDI patients) was a risk factor for CDI that was independent of the risk from antibiotics and other factors in individual patients (Freedberg et al. 2016).

# Increased shedding of spores after recovery from symptomatic CDI

- **Shedding of *C. difficile* spores into the environment** is high after resolution of diarrhoea/end of treatment for CDI (Bobulsky et al. CID 2008)
- The frequency of **skin contamination** and **environmental shedding** remained high at the time of resolution of diarrhoea (60% and 37%, respectively), decreased at the end of treatment, and increased again 1–4 weeks after treatment (58% and 50%, respectively) (Sethi et al. 2010)

# Colonised children as a source of *C. difficile*

- By 2–3 years of age, approximately 1%–3% of children are asymptomatic carriers of *C. difficile*
- Young children are unlikely to have *C. difficile* infection
- Nontoxigenic strains are more common than toxigenic strains among colonized infants and colonisation is transient.
- Asymptomatically colonized infants and children may serve as a source of transmission of the organism to adults, leading to *C. difficile* infection among adult contacts (Wilcox et al 2008, Rosseau et al 2011, Stoesser et al 2011)

# The environment as a source of *C. difficile*

- Based on WGS only 2-7% of new cases acquire *C. difficile* from contaminated environments (Curry et al, 2013, Eyre et al 2013).
- Outbreak in which high-risk fomites (care equipment e.g. blood pressure cuffs, thermometers and stethoscope) shared between patients contributed to transmission (Brooks et al 1992).
- Mathematical modelling estimated that environmental contamination with spores only contributes to 10% of new CDI cases (Starr et al. 2009).
- Effectiveness of sporicidal agents have mostly been demonstrated in outbreak settings when implemented concurrently with other interventions (McMullen et al 2007, Orenstein et al 2011).

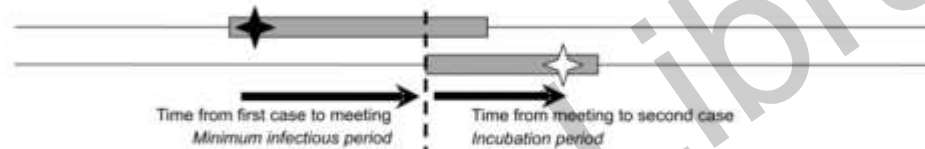
# Scenarios for ward-based contacts with symptomatic CDI cases leading to transmission

(B)

◆ Positive Sample (Donor)    ☆ Positive Sample (Recipient)    ◆ Positive Sample (Donor/Recipient)    █ Ward Stay    █ Ward Contamination Time

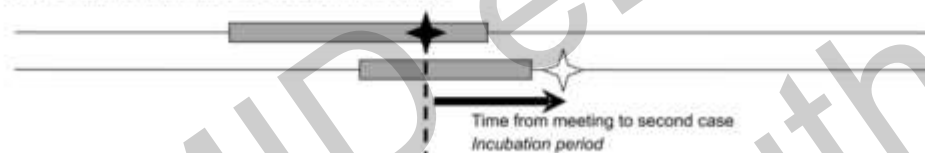
Scenario 1 'Directional'

Ward contact following donor positive sample



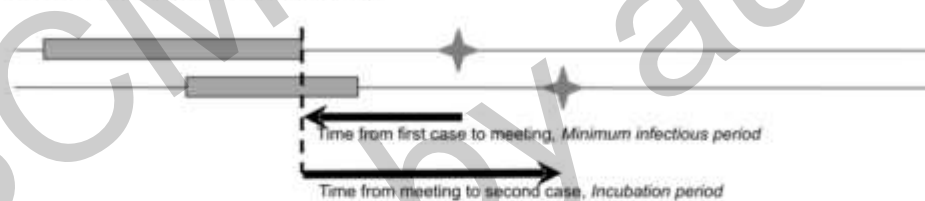
DIRECTIONAL

Ward contact at the time of donor positive sample



Scenario 2, 'Non-Directional'

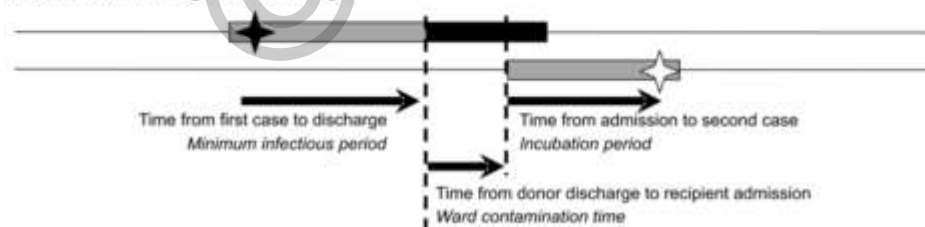
Ward contact prior to either positive sample



NON-DIRECTIONAL

Scenario 3, 'Ward Contamination'

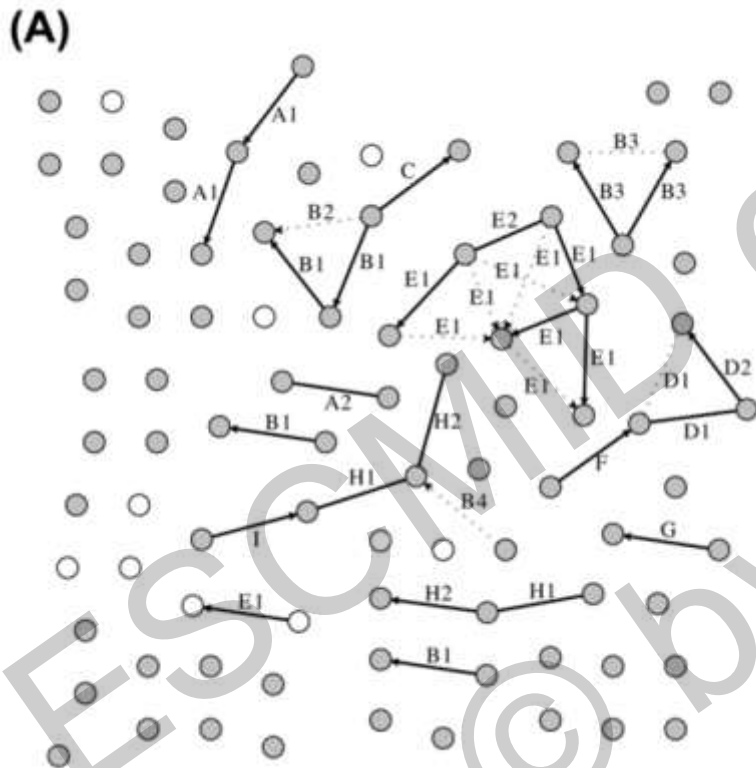
Shared ward following donor discharge



WARD CONTAMINATION



# How do people acquire CDI in hospital in endemic settings?



MLST typing showed:

Overall, only 25% of new cases could be linked to previous symptomatic CDI inpatients -

Specialist surgery – 6%

Acute elderly medicine – 28%

Haematology/oncology – 29%

Renal/transplant – 37%

Walker et al. 2012

**Conclusions:** In an endemic setting with well-implemented infection control measures, ward-based contact with symptomatic enzyme-immunoassay-positive patients cannot account for most new CDI cases.

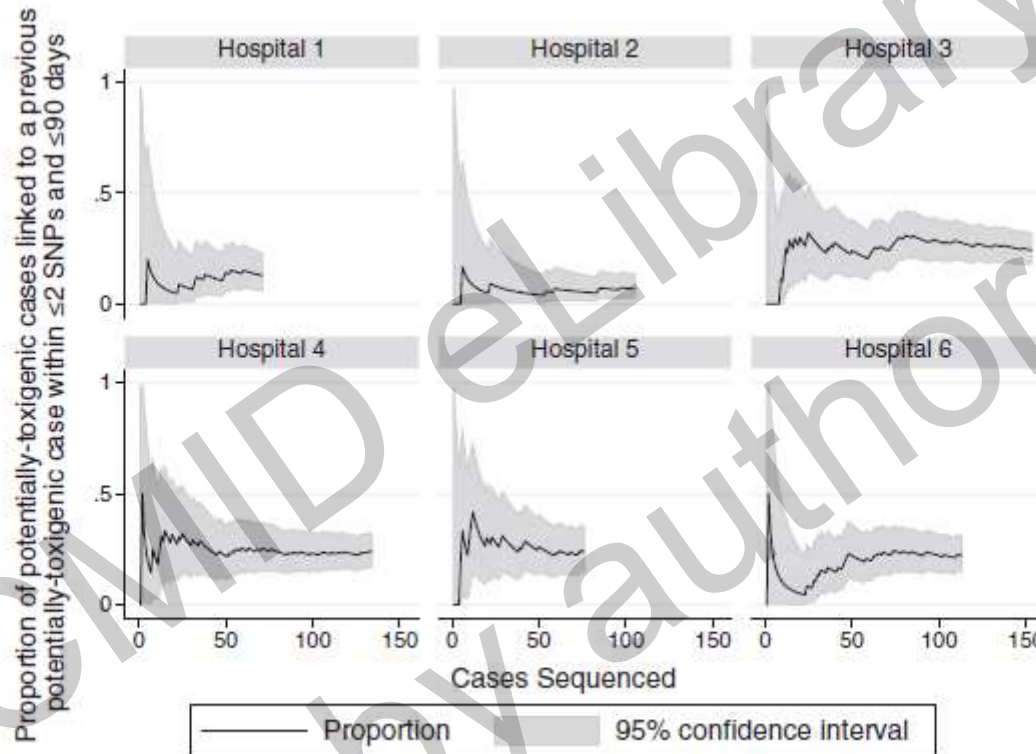
# HA-CDI acquisition in endemic setting (US)

## MLVA typing showed:

- 30% of new cases were associated with CDI patients (4% using same rooms)
- 29% of new cases were associated with asymptomatic carriers (4% using same rooms).
- Concluded identification and isolation of carriers may be necessary to further reduce transmission of *C. difficile*



# Multi-centre study using WGS to identify HA-CDI acquisition (UK)



Eyre et al 2017

**Figure 2.** Proportion of potentially toxicigenic cases linked to a previous potentially toxicigenic case by hospital and number of sequences obtained. Abbreviation: SNP, single-nucleotide polymorphism.

- 20% of CDI patients were linked to previous CDI patients (< 2 SNP difference) (range 7-26%).
- Variation among ribotypes (57% of RT 027 cases were linked).

# Mathematical modelling of screening and contact precautions for carriers

- Mathematical modelling studies have suggested that reductions in CDI incidence by 10%–25% could be achieved by identifying and isolating carriers upon hospital admission (Grigoras et al. 2016, Lanzas et al, 2014)
- Exposed individuals who have not yet been colonised will be missed in screening tests (Yakob et al, 2013)
- ‘Screening + isolation’ approach was tested in a hospital in Quebec with high incidence of CDI (Longtin et al 2016)
  - Significant decrease in incidence
  - Sustained for at least 1 year after the study.

# Basic research gaps - colonisation

- What enables *C. difficile* to colonize patients?
- What are the triggers for sporulation and germination of *C. difficile* in the GI tract?
- Is there a nutritional niche that allows colonization?
- What is the role of mucosal/systemic immunity in preventing symptomatic CDI?
- What causes *C. difficile* colonization to end?
- What are the factors in infants and young children that influence susceptibility to infection vs. colonization?

### 3. Extant guidance 2018

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# CDI Guidance Scotland 2017

## Asymptomatic carriers



- Testing of stools from asymptomatic carriers is not recommended (IB)
- Routine screening of asymptomatic persons for CDI is not recommended (no evidence)
- Transmission based precautions (or other interventions) are not recommended for carriers (insufficient evidence)

# ESCMID guidance 2018

## Recommendations relating to asymptomatic carriers

Categories of recommendations	Recommendations relating to asymptomatic carriers
Diagnostic testing	No*
Surveillance	No
Screening	No**
Hand hygiene	No
PPE	No
Contact precautions	No
Environmental cleaning and disinfection	No
Infrastructure changes	No
Antibiotic stewardship	No
Education	Yes

\*The ESCMID Diagnostic Guidance (Crobach et al. 2016) addresses issues relating to ambiguous test results

\*\* No recommendations given and reasons indicated.



# ESCMID Guidance 2018

## Screening

### Does screening help to identify colonised patients?

#### ***Recommendation for outbreak and endemic settings***

4. We do not recommend screening for *C. difficile* to identify colonised/carrier patients as a way of altering the risk of developing CDI in either colonized subjects or other patients and thus reducing CDI-rates (conditional recommendation, low level of evidence in the endemic setting).

### Should Healthcare workers be screened?

#### ***Recommendation for outbreak and endemic settings***

5. We do not recommend HCW screening for *C. difficile* gut colonization as a routine control measure for CDI (strong recommendation, very low level of evidence in the endemic setting).

# ESCMID Guidance 2018

## Environmental control

**Does environmental disinfection of rooms of CDI patients decrease transmission of *C. difficile*?**

***Recommendation for the outbreak setting***

14. Introduce daily environmental sporicidal disinfection and terminal disinfection of rooms of patients with CDI to decrease the transmission of CDI (strong recommendation, very low quality of evidence).

**Are no-touch disinfection systems effective in reducing contamination?**

**Recommendation for outbreak and endemic settings**

17. The panel concludes that both in the outbreak and the endemic setting, no touch disinfection systems may be effective in reducing transmission/ incidence of CDI (very low quality of evidence).

# ESCMID Guidance 2018

## Education – Environmental control

### Does specific education enhance thoroughness of cleaning aimed at prevent CDI?

Education of environmental service personnel proved to be of particular importance for prevention of CDI as reducing environmental *C. difficile* contamination was associated with lower *C. difficile* prevalence in hospitals.

Environmental service personnel require repeated training and regular quality control measurements (e.g. by labelling of surface areas before cleaning with a fluorescence marker) to ensure sustained high quality cleaning.

# CDI guidance IDSA/SHEA 2018

## Should asymptomatic carriers be tested and isolated?

- No recommendation on screening for asymptomatic carriage
- No recommendation on contact precautions

## Is there a role of asymptomatic testing - Should repeat testing be allowed for asymptomatic patients?

- Do not retest during the same episode
- Do not test asymptomatic patients (unless it is part of an epi study)

## 4. Discussion and conclusions

# Discussion and conclusions (1)

- There is conflicting evidence regarding the contribution of asymptomatic carriage to the epidemiology and spread of CDI; which may also vary with burden of disease, ribotype and healthcare setting.
- Multi-centre studies in different hospital settings are needed to support the “screening-isolation” approach more widely.
- Currently national guidelines generally agree on recommendations re. **asymptomatic carriers**:
  - **No testing**
  - **No screening**
  - **No contact precautions**
- Education, environmental measures and standard infection control precautions and antimicrobial stewardship should be in place **to prevent and control spread of *C. difficile* spores and development of CDI.**

# Discussion and conclusions (2)

- Even if sufficient evidence will be produced to support the implementation of a 'screening-isolation' approach; this will need to be assessed against public health principles for approval and funding (UK principles abbreviated here):
  1. The epidemiology and natural history of the condition, including **latency period**, risk factors and disease markers, should be known
  2. There should be a safe, accurate and validated **screening test available**
  3. There should be agreed evidence-based strategies **for effective intervention or treatment** for identified individuals.
  4. **Implementation should be underpinned by a complete screening programme** (test, diagnostic procedures, treatment/ intervention) that is **clinically, socially and ethically acceptable to health professionals and the public**

# Questions

Q1:

Do you test asymptomatic patients for presence of *C. difficile*?

- A) yes
- B) no

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# Questions

Q2:

Do you screen for asymptomatic carriers of *C. difficile*?

- A) yes
- B) no

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# Questions

Q2:

Do you isolate carriers positive for toxigenic *C. difficile*?

- A) yes
- B) no

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Thank you for your attention

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