

Staphylococcus aureus sortase A mediated incorporation of peptides: effect of peptide modification on incorporation

Silvie Hansenová Maňásková^{1,2}, Kamran Nazmi¹, Alex van Belkum², Wim van t Hof¹, Nathaniel I. Martin³, Floris J. Bikker¹, Willem J.B. van Wamel², Enno C.I. Veerman¹

¹ Department of Oral Biochemistry, Academic Centre for Dentistry Amsterdam, University of Amsterdam and VU University Amsterdam, Amsterdam, The Netherlands

² Department of Medical Microbiology and Infectious Diseases, Erasmus MC, Rotterdam, The Netherlands

³ Department of Medicinal Chemistry and Chemical Biology, Utrecht Institute for Pharmaceutical Sciences, Utrecht University, Utrecht, The Netherlands



Introduction

Endogenous *Staphylococcus aureus* Sortase A covalently anchors bacterial surface proteins equipped with a specific recognition motif (LPXTG) into the peptidoglycan layer of the cell wall:

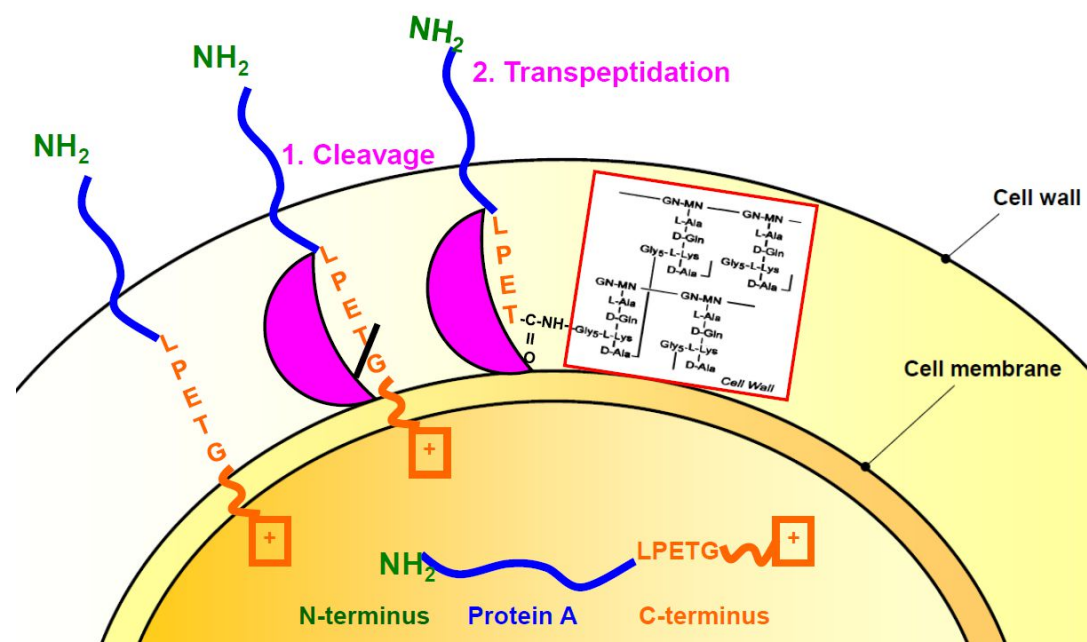


Figure 1: LPETG containing *S. aureus* surface protein incorporation into the staphylococcal peptidoglycan.

Previously, we showed:

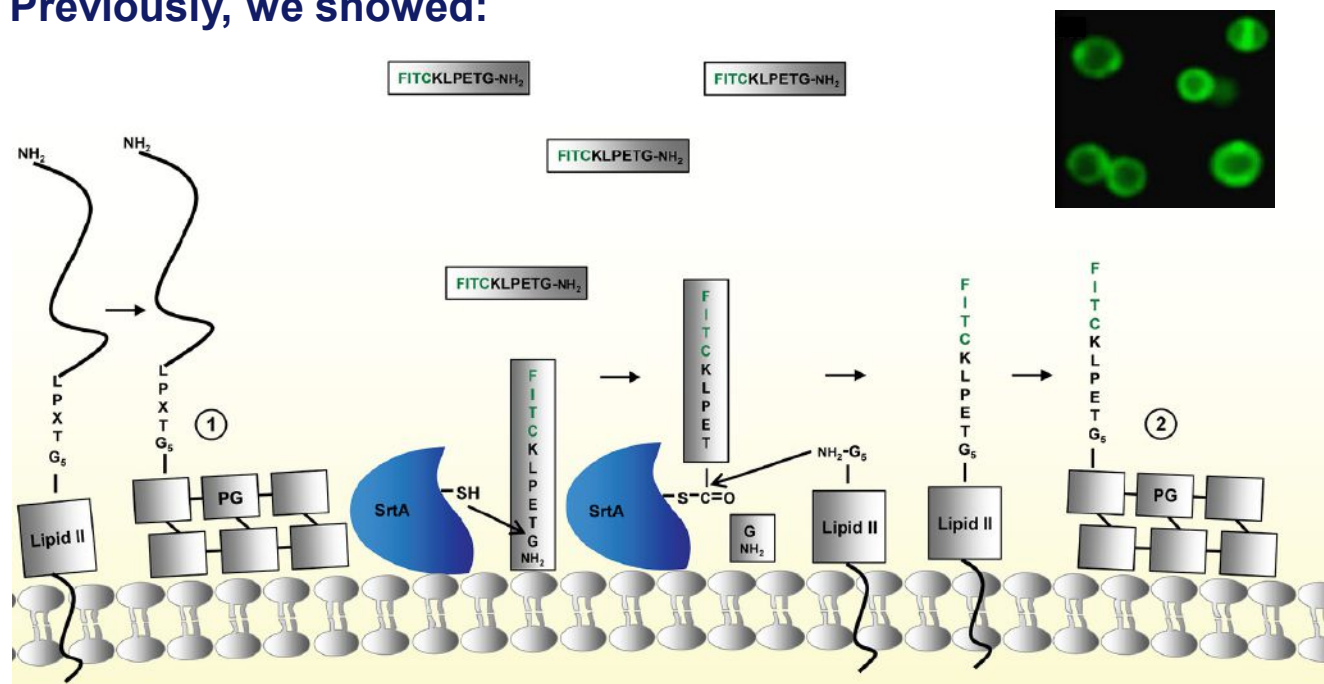


Figure 2: A proposed model of SrtA synthetic substrate incorporation. We propose that the exogenously added SrtA synthetic substrates, such as a FITC-K-LPETG-amide are recognized and processed in the same manner as the native CWA proteins (as described in Fig. 1). CWA proteins are covalently incorporated into the bacterial cell-wall (1) in addition to native SrtA substrates (2).

Aim of this study:

Since high concentrations of the synthetic FITC-K-LPETG-amide substrate were required (1mM), we aimed in this study to improve the incorporation efficiency by modification of the substrate.

Materials & Methods

S. aureus strains (WT and $\Delta SrtA$) were exposed to (fluorescently) labelled synthetic sortase substrates (Table 1) during growth in a Luria broth (LB) medium and the fluorescence was determined with Facs scan.

Table 1: Substrates used in this study:

Substrate number	N-term	Substrate sequence	C-term
1	K(FITC)	LPETG	amide
2	K(FITC)	EGTLP	amide
3	K(FITC)	AKKSELPETGGEESTNKGMLFGGLFSILGLALLRRNKKNHK	amide
4	K(FITC)	AKKSELPETGGEESTNKRRNKKNHKAGMLFGGLFSILGLALL	amide
5	K(FITC)	AKKSELPETGGEESTNKRRNKLKNHK	amide
6	K(FITC)	LPETGGEESTNKRKKW	amide
7	K(FITC)	LPMTG	amide
8	K(FITC)	K(Vancomycin)LPMTG	amide
9	K(FITC)	K(Vancomycin)MGTLT	amide
10	K(FITC)	LPMTGG-K(vancomycin)	amide
11	K(FITC)	Vancomycin-K-AKKSELPETGGEESTNKRRNKLKNHK	amide
12	K(FITC)	K(Vancomycin)LPMTG	amide
13	K(FITC)	K(Vancomycin)MGTLT	amide

Results

The elongation of the FITC-K-LPETG-amide substrate with the positively charged sequence derived from membrane spanning domain of physiological SrtA substrates resulted in a 20-fold higher incorporation.

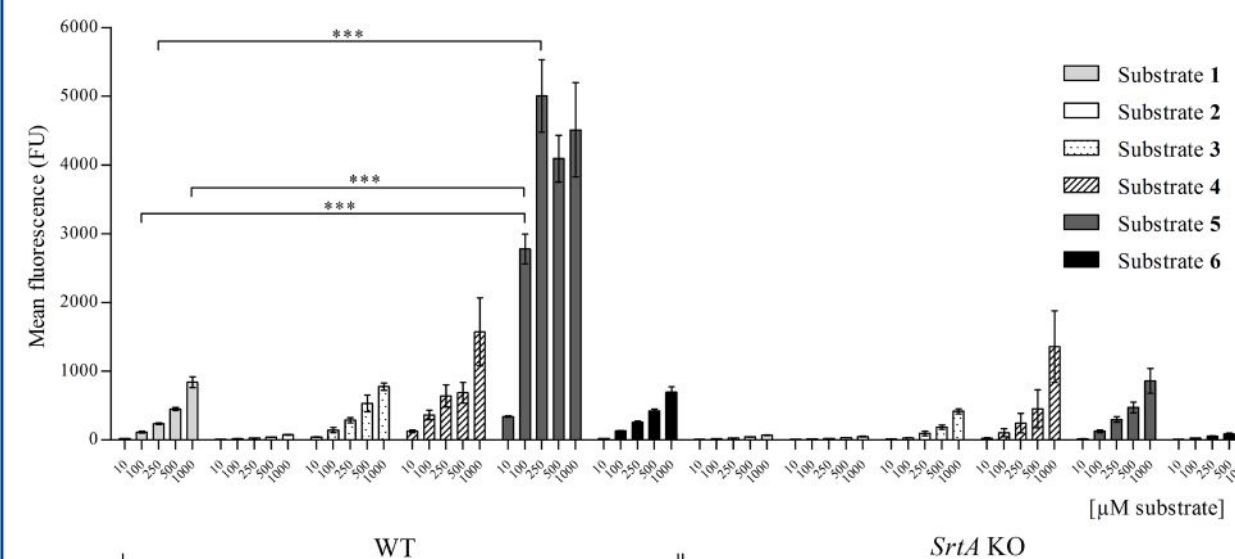


Figure 3: Titrations of the SrtA substrates.

FITC-K-LPMTG-amide is incorporated three and half times more efficiently into the WT *S. aureus* bacteria than the FITC-K-LPETG-amide.

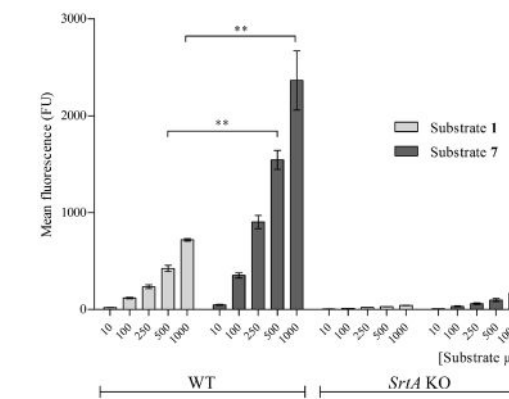


Figure 4: Titration of the FITC-K-LPMTG-amide substrate.

Covalent coupling of vancomycin to FITC-K-LPMTG-amide resulted in the same incorporation at a 500-times lower substrate concentration. However, this incorporation did not result in an increased vancomycin efficiency.

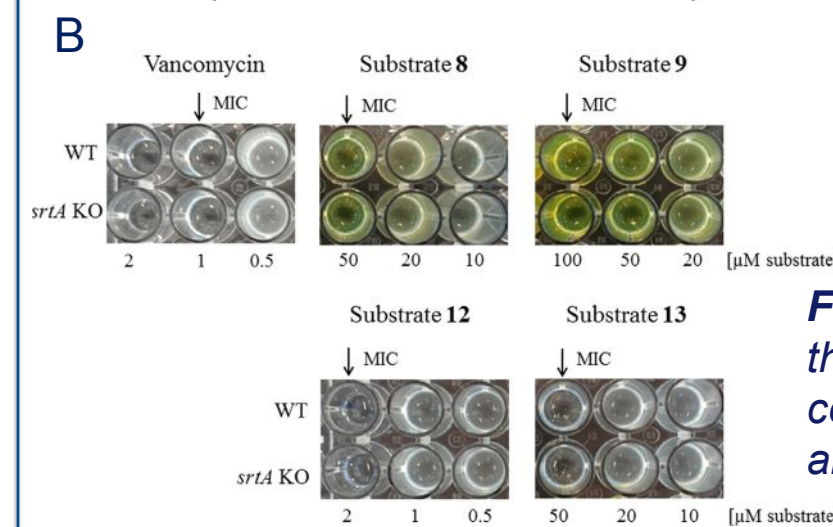
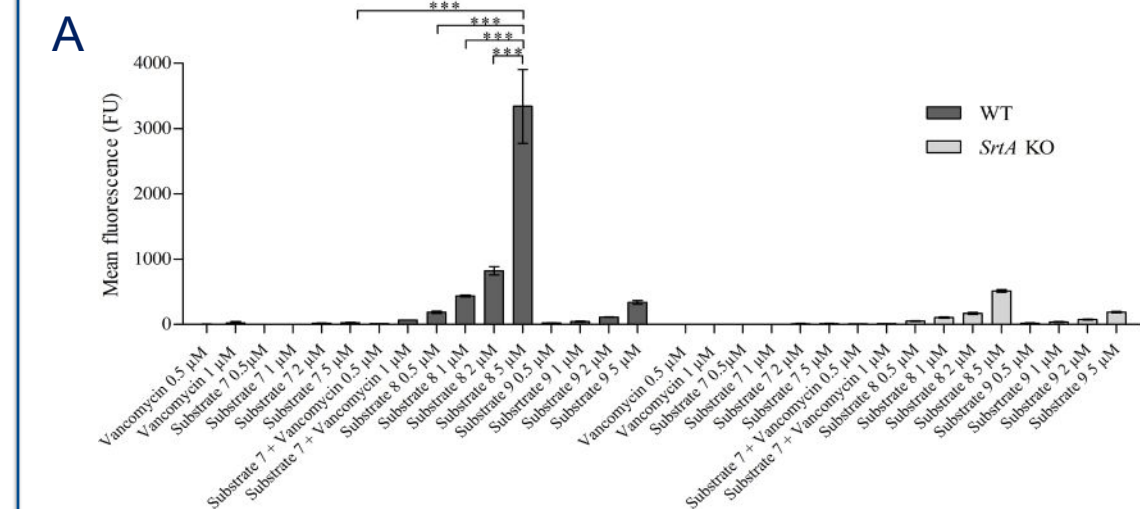


Figure 5: (A) Titration of the FITC-K-LPMTG-amide coupled to vancomycin and (B) MIC determination.

Conclusions

These newly developed synthetic substrates can potentially find broad applications in for example the *in situ* imaging of bacteria; the targeted delivery and covalent incorporation of antimicrobial compounds into the bacterial cell wall.