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Can we start or stop antifungal therapy based on biomarkers detection?

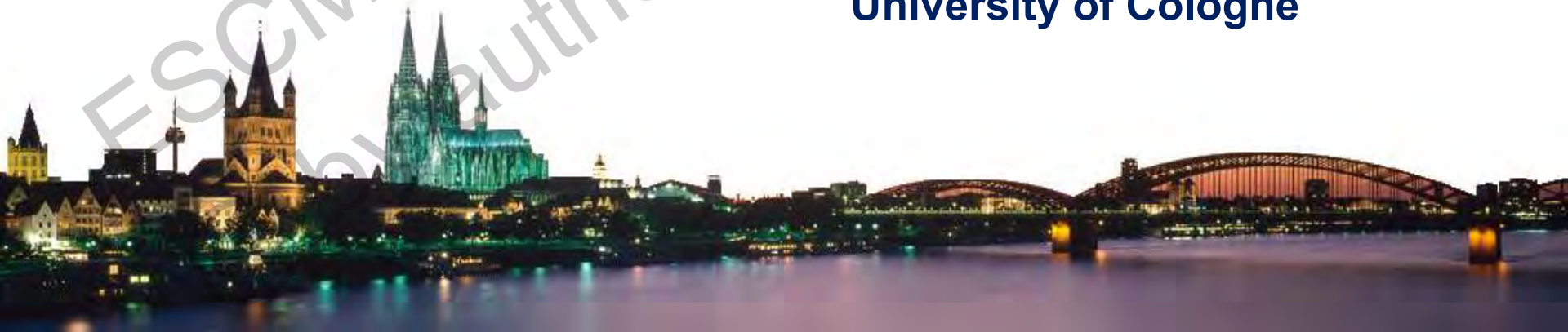
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Chair, Translational Research Institute

Chair, Centre for Clinical Trials

Deputy Head, Infectious Diseases



University of Cologne





- **European Commission**
 - **FP7, IMI-JU 6 (COMBACTE), 8 (APC), 9 (CARE)**
- **European Organisation for Research and Treatment of Cancer (EORTC)**
- **European Society for Clinical Microbiology and Infectious Diseases (ESCMID)**
- **European Confederation of Medical Mycology (ECMM)**
- **German Federal Ministry of Research and Education**
 - **BMBF 01KN1106, 01KN0706, 01GH1001E, 01EZ0931, 01EK1422**
- **German Center for Infection Research (DZIF)**
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- **German José Carreras Leukaemia Foundation (DJCLS)**
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 - **3M, Actelion, Astellas, AstraZeneca, Basilea, Bayer, Celgene, Cidara, Da Volterra, Daiichi Sankyo, F2G, Genentech, Genzyme, Gilead, GSK, Medpace, Merck Serono, MSD, Miltenyi, NanoMR, Novartis, Parexel, Pfizer, Quintiles, Rempex, Roche, Sanofi Pasteur, Shionogi, Summit, Vifor, Viropharma**



Strategy	Definition
Prophylaxis	Administration of the antifungal agent is initiated at a period of high risk of infection to prevent fungal infections
Empirical Treatment	Initiation or modification of an existing antifungal treatment in persistently febrile patients with neutropenia (4–7 days in duration) that is without a known source of infection and is unresponsive to appropriate antibacterial agents. 
Pre-Emptive Therapy	Similar to empirical antifungal therapy, preemptive therapy aims to treat a suspected early IFI but uses radiologic studies, laboratory markers, or both (rather than clinical criteria alone) to stratify the likelihood of an IFI; specific criteria would trigger preemptive initiation of antifungal therapy. 
Treatment of established IFI	Corresponds to patients who meet European Organization for Research and Treatment of Cancer/Mycoses Study Group criteria for proven and probable IFI



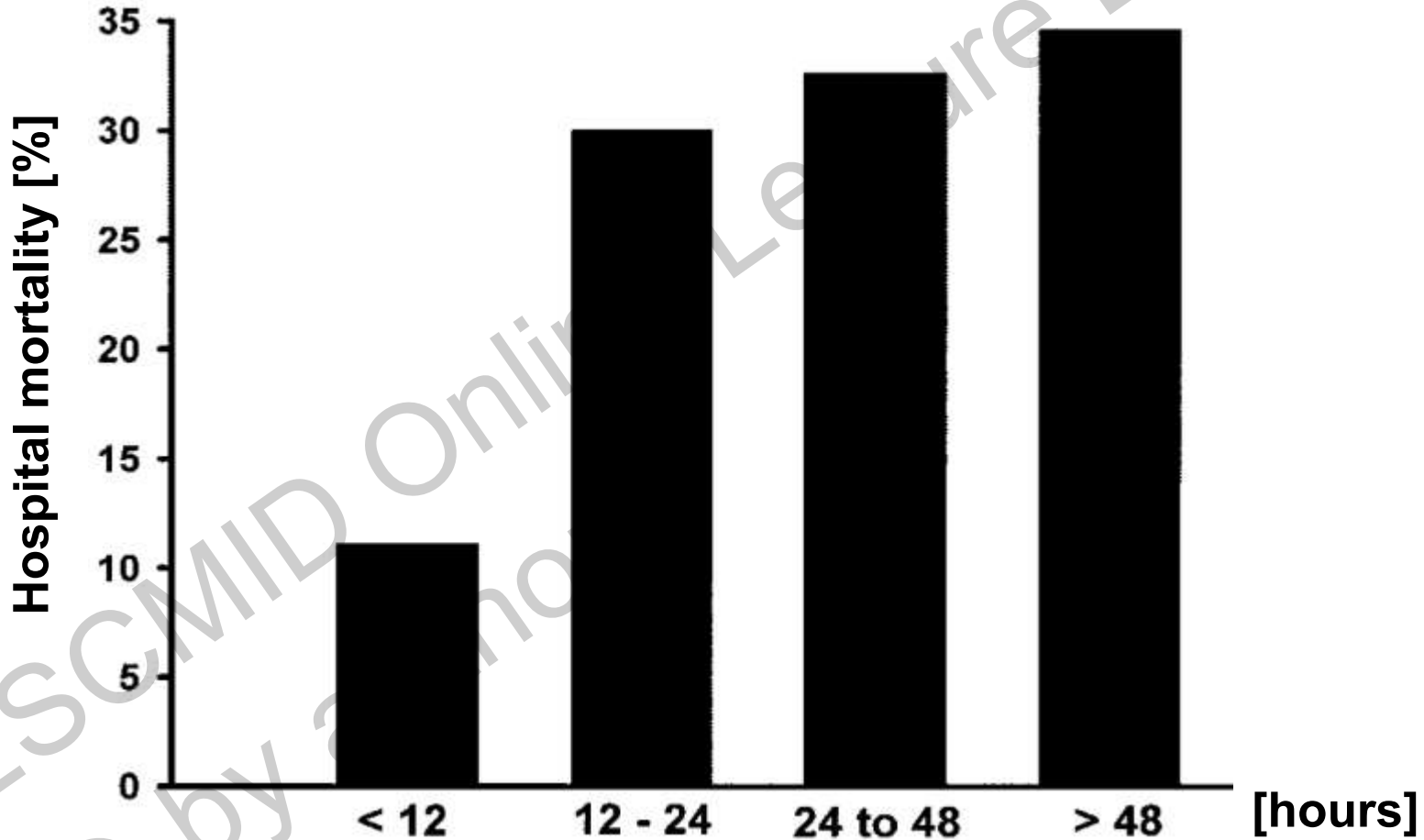
	Start	Stop	Start	Stop
Candidiasis	N/A	N/A	BDG	BDG
Aspergillosis	GM	GM	BDG	BDG



- Patient ventilated, Pip/Tazo Day 6, persistent fever, otherwise stable, no pathogen isolated
- ICU rounds twice daily, continuous discussion pro/con empiric antifungal treatment
- The right decision: Depends ...
- Did the patient undergo abdominal surgery ?
- Is the patient colonised ?



Treatment Delays Increase Mortality in IC



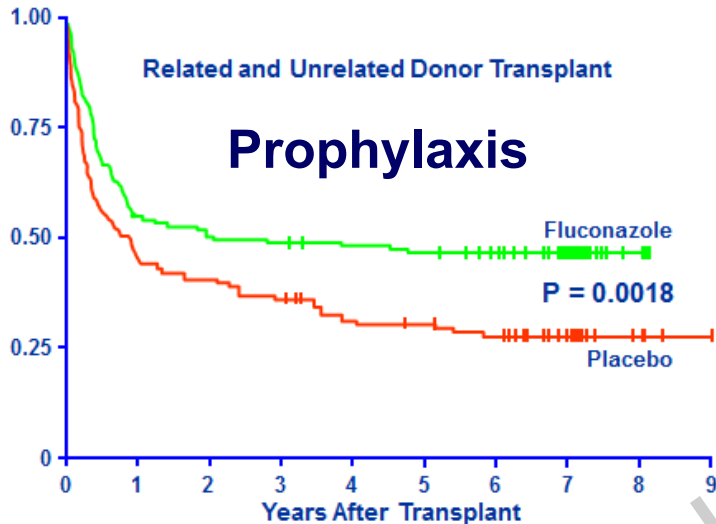


**Early Exposure to Antifungals is a
Common Pattern of all Trials
Improving Survival Rates**

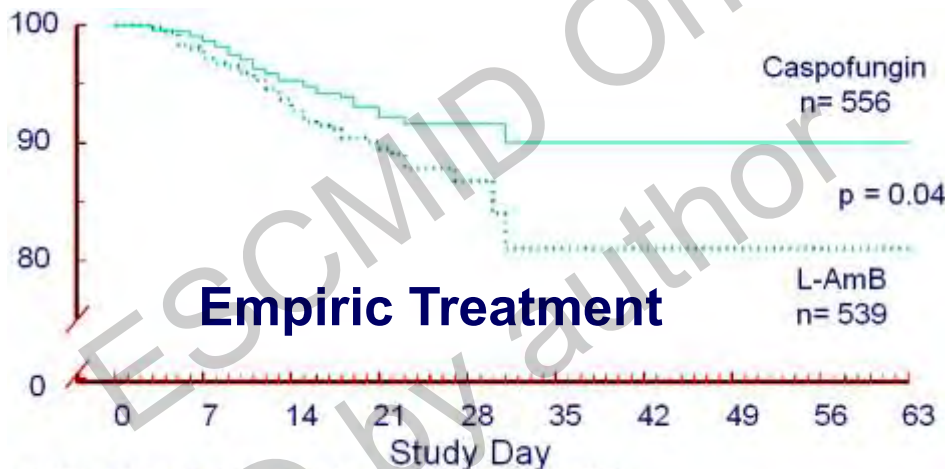
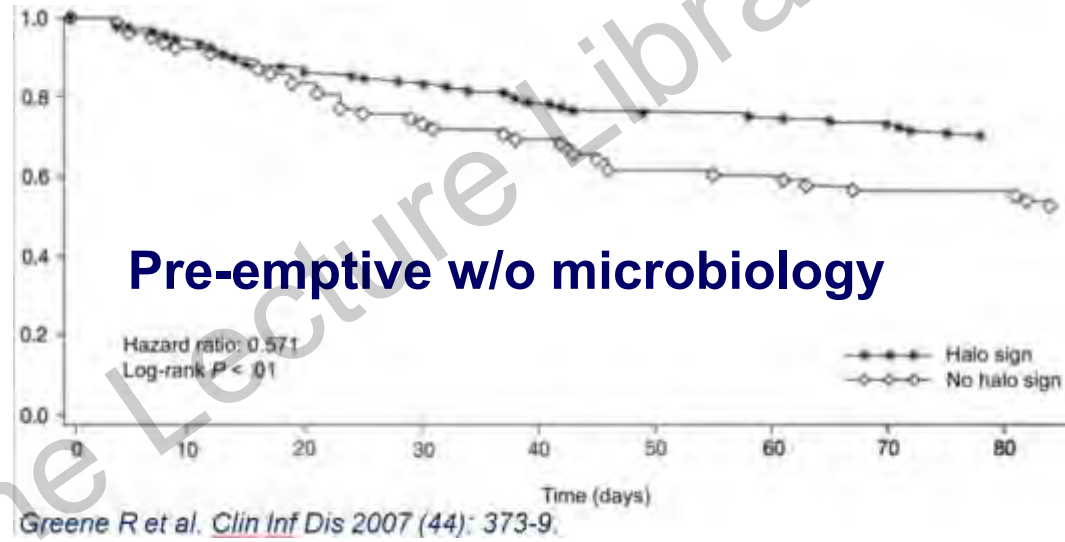
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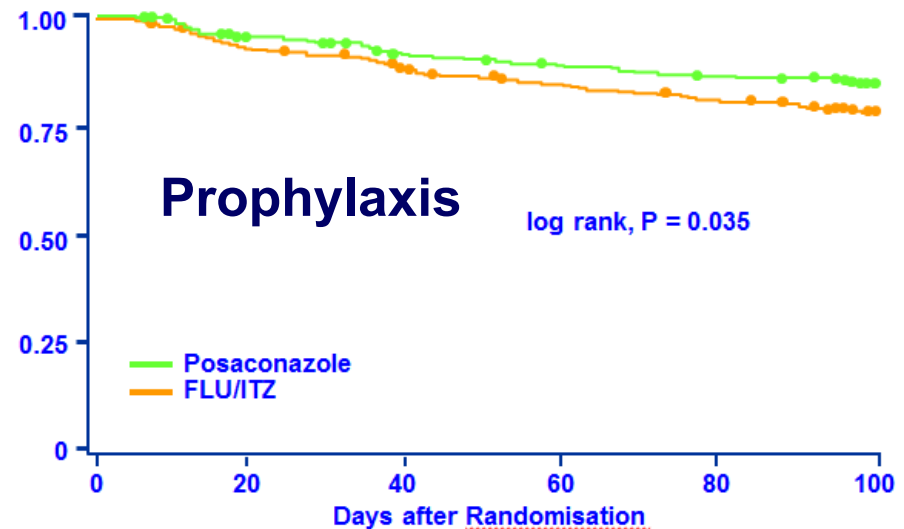
Trials That Yielded a Difference in Survival



Slavin M et al. *J Infect Dis* 1995 / Marr K et al. *Blood* 2000.



Walsh TJ et al. *N Engl J Med* 2004; 351:1391-402.



Comely OA et al. *N Engl J Med* 2007.



**Reliable Diagnostic Tests would
Allow Early Treatment to be Targeted**

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Diagnostic tools are too few and are unreliable

- „One fungus – one name“ we welcome
- „One fungus – one test“ is no ! solution
 - **Aspergillus – GM:** 10 years to a cut-off
 - **Aspergillus – PCR:** 15 years to standardization
 - **Mannan/Anti-Mannan:** Any good at all?
 - **β-D-Glucan:** Benefits not yet fully explored

All rely on
the same
principle!

**Give up the paradigm of proving the presence of
the pathogen?**



Galactomannan

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- AML
- Neutropenia
- Fever >72h
- Cough
- Dyspnea
- Pleuritis

Galactomannan



A typical AmBiLoad Study Patient from Cologne.

Blood galactomannan

Population	Intention	Intervention	SoR	QoE	Comment
Prolonged neutropenic patients and allogeneic stem cell transplantation recipients not on mold-active prophylaxis	To diagnose invasive aspergillosis	Galactomannan in blood	A	I	Highest test accuracy requiring 2 consecutive samples with an OD index ≥ 0.5 ; prospective monitoring should be combined with HRCT and clinical evaluation
		Draw samples every 3-4 days	C	III	
Patients with a hematological malignancy	To diagnose invasive aspergillosis	Galactomannan in blood			Significant lower sensitivity in non-neutropenic patients
			•Neutropenic •Non-Neutropenic	A B	

Diagnostic Tools - B-D-glucan Assay

Population	Intention	Intervention	SoR	QoE	Comment
Mixed population: Adult ICU, Hematological disorders, SOT	To diagnose IFD (not specific for aspergillosis)	Diagnostic assay	C	II	4 different assays; Fungitell FDA approved and available in US and Europe; others only available in Japan; overall sensitivity of 77% and specificity of 85% Specificity limits its value in this setting Two or more consecutive samples: sensitivity = 65%, specificity = 93%; studies included once to thrice weekly
		Screening assays	C	II	Varies with assay and cut-off: Wako assay sensitivity = 40-97%, specificity = 51-99%



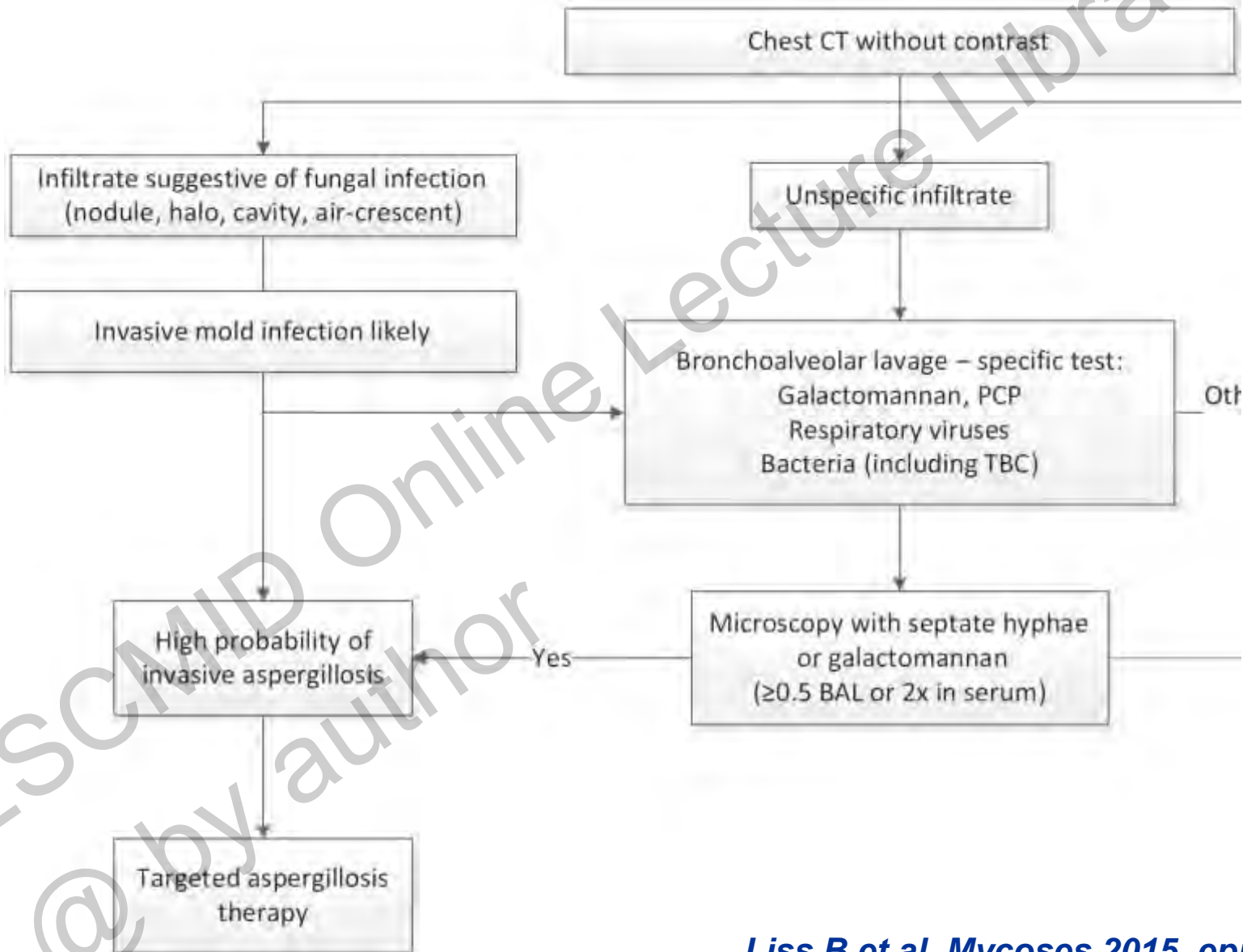
Our 2015 approach to invasive pulmonary aspergillosis

**B. Liss,^{1,2} J. J. Vehreschild,^{1,2,3} C. Bangard,⁴ D. Maintz,⁴ K. Frank,⁵ S. Grönke,⁵ G. Michels,⁵
A. Hamprecht,⁶ H. Wisplinghoff,⁶ B. Markiefka,⁷ K. Hekmat,⁸ M. J. G. T. Vehreschild^{1,2,3} and
O. A. Cornely^{1,2,3,9,10}**

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Starting with Positive GM





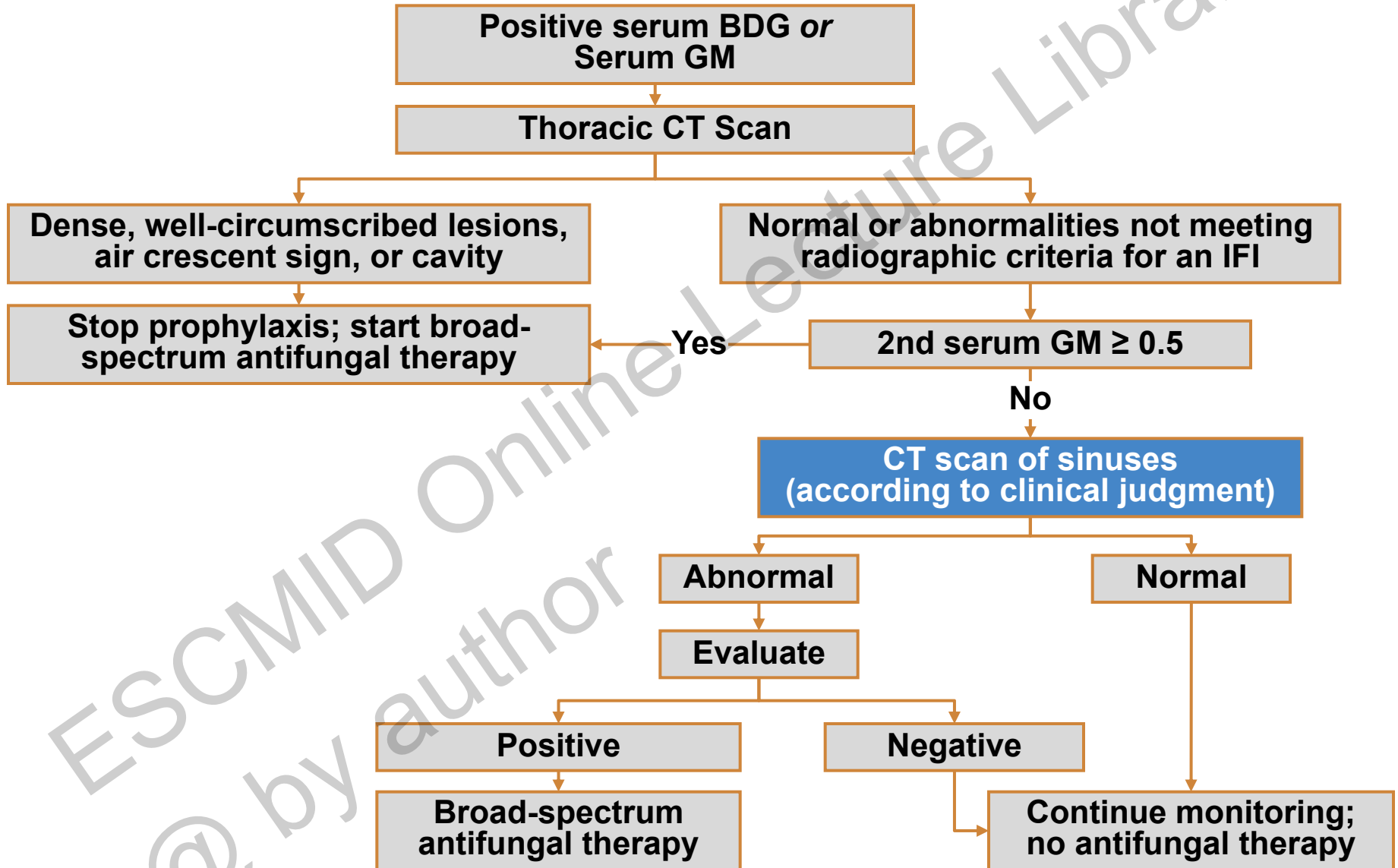
Patients routinely monitored for signs and symptoms of IFI throughout the study period

- Twice weekly galactomannan (GM) and β -D-glucan (BDG)
- Diagnostic workup if 1 positive GM/BDG antigen assay
- Algorithms followed for investigation and management of suspected IFI

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Algorithm for Suspected IFI





Pathogen Causing IFD ^{a, b}	Isavuconazole (N = 143)	Voriconazole (N = 129)
Proven/Probable IFD	29 (11.2%) / 114 (44.2%)	36 (14.0%) / 93 (36.0%)
Galactomannan only ^c	71 (49.7%)	68 (52.7%)
<i>Aspergillus</i> spp. only	49 (34.3%)	39 (30.2%)
<i>Aspergillus</i> spp. plus other filamentous fungi	3 (2.1%)	1 (0.8%)
Non- <i>Aspergillus</i> spp. only	5 (3.5%)	6 (4.7%)
Filamentous fungi NOS	14 (9.8%)	15 (11.6%)

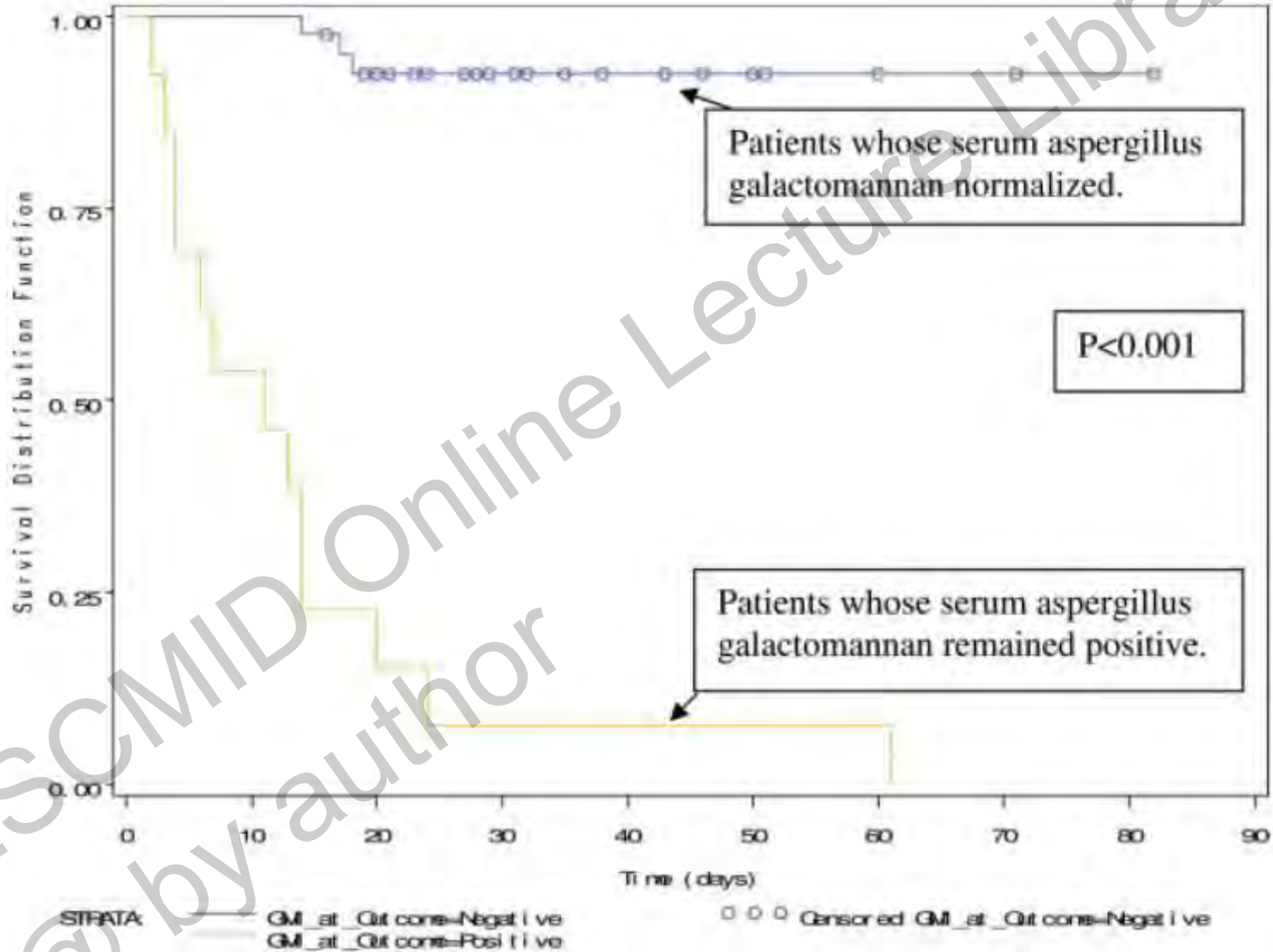
^aAs assessed by the DRC

^bNote, >90% of the mITT population had pulmonary involvement

^cSerum: 1 value ≥ 0.7 or 2 serial values $\geq 0.5 - < 0.7$; Bronchoalveolar lavage: 1 value ≥ 1.0



Stopping with Negative GM



Consensus statement:

Discontinuation of targeted therapy (SoR: C)

- The range of the duration of treatment is huge and the evidence base to support any particular recommendation is weak
- Need to separate between targeted and salvage or secondary prophylaxis (and long-term toxicity)
- Need to consider iv oral switch in stable and PK-reliable patients
 - Duration depending upon reconstitution of the immune system, continuing GvHD, etc. (i.e. secondary prophylaxis)
 - Need CR (radiographic imaging, scaring allowed) which includes no clinical or microbiological evidence of disease prior to discontinuation
- Close monitoring (e.g radiographical imaging) once discontinued.



1,3- β -D-Glucan

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Table 5 Possible confounding factors of BG reactivity

Variable at the time of BG sampling ^a	Variable present		Variable absent		<i>p</i> -Value ^f
	Frequency (no. of sampling episodes)	Mean BG level, pg/ml (95 % CI)	Frequency (no. of sampling episodes)	Mean BG level, pg/ml (95 % CI)	
Fungal mucosal colonization ^b	63	86.1 (63.8–108)	400	76.2 (69.0–83.4)	ns
Bacteremia ^c	15	70.6 (31.3–110)	662	72.3 (67.3–77.4)	ns
Admitted to ICU	9	196 (86.1–306)	668	70.6 (65.8–75.4)	0.0007
Dialysis ^d	9	78.3 (20.4–136)	668	72.2 (67.1–77.2)	ns
Intravenous immunoglobulins ^e	9	177 (61.1–293)	668	70.9 (66.0–75.7)	ns
Plasma, coagulation factors, or albumin ^f	16	159 (85.7–232)	661	70.1 (65.4–74.9)	0.0145
Total parenteral nutrition ^g	119	79.8 (66.2–93.4)	558	70.7 (65.3–76.0)	ns
Intravenous antibiotics of any kind ^h	184	75.7 (64.9–86.4)	493	71.0 (65.4–76.6)	ns
Piperacillin–tazobactam ^h	22	65.5 (44.9–86.2)	655	72.5 (67.4–77.6)	ns
Meropenem ^h	107	70.1 (57.1–83.2)	570	72.7 (67.3–78.1)	ns
Ceftazidime ^h	20	106 (52.7–160)	657	71.2 (66.3–76.1)	ns
Vancomycin ^h	45	68.4 (52.9–83.9)	632	72.5 (67.3–77.8)	ns
Pegylated asparaginase ⁱ	85	118 (93.0–142)	592	65.7 (61.5–70.0)	<0.0001



Table 4 Positive and negative predictive values at different hypothetical IFI prevalence rates and with different strategies for BG testing, based on the previously suggested optimal cut-off level of 158 pg/ml

Hypothetical prevalence of IFI	Strategy for BG testing to define IFI (pg/ml)					
	Mean BG >158 ^a		One BG >158 ^b		Two consecutive BG >158 ^c	
	PPV	NPV	PPV	NPV	PPV	NPV
5 %	0.54	1.0	0.16	0.99	0.49	0.99
8 %	0.66	0.99	0.24	0.99	0.62	0.99
12% ^d	0.75	0.99	0.33	0.99	0.71	0.98
15 %	0.80	0.99	0.39	0.98	0.77	0.97
20 %	0.85	0.98	0.48	0.98	0.82	0.96

Pre-emptive Therapy: β -D-Glucan

Popu- lation	Intention	Inter- vention	SoR	QoE	Reference	Comments
ICU	Early treatment of invasive candidiasis / candidaemia	To treat when β -D-glucan test is positive	C	II _u	Desmet JCM 2009 Digby Clin Diagn Lab Immunol 2003 Koo CID 2009 Mohr JCM 2011 Presterl Int JID 2009 Takesue WJSurg 2004 Pickering JCM 2005	<ul style="list-style-type: none"> • Low specificity • Low sensitivity • High NPV • False positives with <ul style="list-style-type: none"> • Haemodialysis • Other fungal or • Bacterial infection • Wound gauze • Maybe useful in PCP



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Stopping with Negative BDG

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Descriptive analysis of biomarkers at EOT assessment (FAS)*

Biomarker	Placebo		Micafungin 100 mg	
	n		n	
Mean change (SD) from baseline at EOT				
Beta-D-glucan [†] , pg/mL	54	53.0 (355.7)	44	-34.9 (206.6)
<i>Candida</i> antibody, AU/mL	103	13.1 (21.4)	87	12.7 (20.9)
Mannan antigen, pg/mL	103	24.2 (281.3)	87	18.0 (236.8)
EOT assessment				
PCR detection of <i>Candida</i> in patients (%)	105	7.6%	89	5.6%

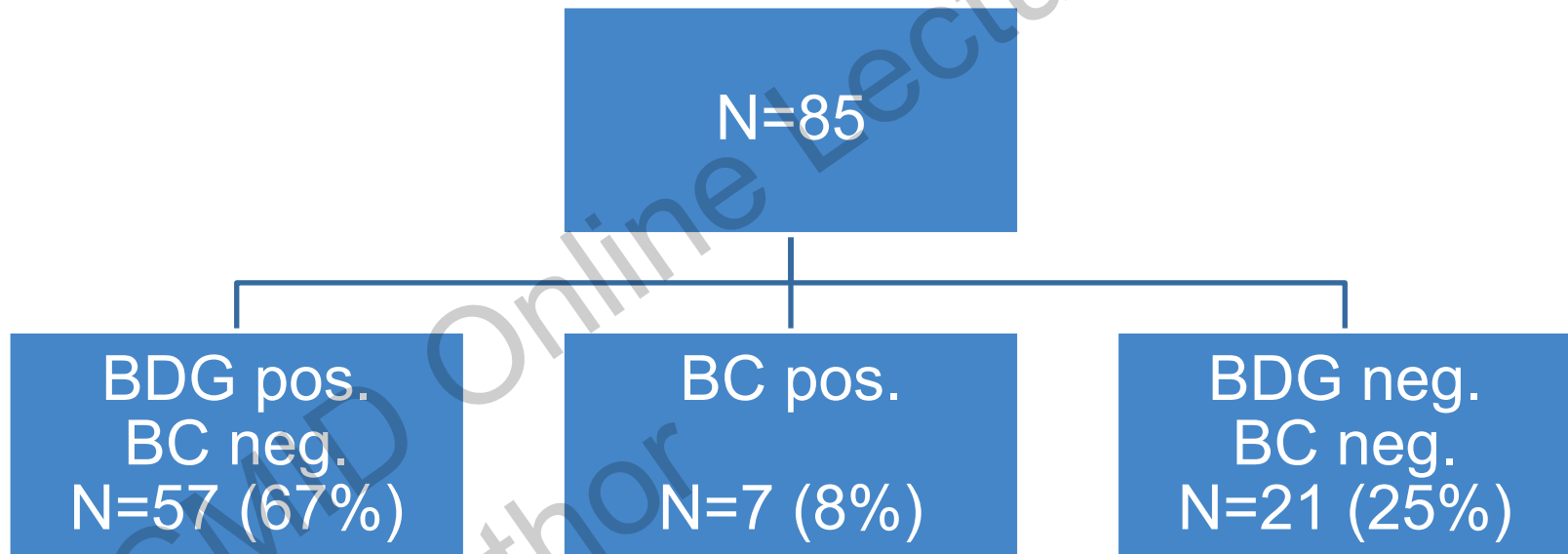
*Biomarker data were not available for all patients in the FAS; [†]Positive results ≥ 62.5 pg/mL) provided, negative results recorded and imputed as < 62.5 pg/mL



- 85 of 2148 ICU patients had all of the below:
 1. CVC
 2. Antibiotic treatment
 3. 2 of: dialysis, surgery, pancreatitis, steroids/immunosuppression, parenteral nutrition
 4. 1 of: fever, hypothermia, hypotension, leukocytosis, acidosis, or CRP \uparrow
- Received echinocandin treatment and
 - Diagnostic screening
 - Day 1 and 2: Blood culture
 - Day 1, 2, and 3: β -D-Glucan



Stopping with Negative BDG





LFD
PCR
T2
T-cells

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Biomarker/LFD

Population	Intention	Intervention	SoR	QoE	Comment
Hematological malignancy and solid organ transplant	Diagnose IA	Evaluation of LFD using BAL samples (retrospective study)	B	III	Sensitivity and specificity of BAL LFD tests for probable IPA were 100% and 81% (PPV 71%, NPV 100%); 5 pts with possible IPA had positive LFD ; no proven patients
Hematopoietic stem cell transplantation (HSCT)	Diagnose IA LFD using serum samples	Prospective screening in 101 patients undergoing allo-HSCT Comparison to Asp-GM to serum	B	III	IA: 1 proven, 9 probable, 20 pos. 1 serum vs 2 serum samples positive: sensitivity 40%/20%; specificity 86.8%/97.8%; diagnostic odds ratio 3.03/11.13
Immunocompromised pts (hematological malignancies 64%)	Diagnose IA	Evaluation of LFD using BAL samples (retrospective study)	B	II	Sensitivities for LFD, GM, BDG, PCR were between 70 and 88%; combined GM (cut-off >1.0 ODI) with LFD increased the sensitivity to 94%, while combined GM (>1.0) with PCR resulted in 100% sensitivity (specificity for probable/proven IPA 95-98%).

BAL *Aspergillus* PCR

Population	Intention	Intervention	SoR	QoE	Reference	Comment
Patients undergoing allogeneic stem cell transplantation recipients not on mold-active prophylaxis	To predict IA	BAL PCR	B	II	Einsele, Lancet, 1998	In house assay
Patients with hematological malignancies and prolonged neutropenia ICU pts (mixed pts populations) Lung Tx pts	To diagnose IA	BAL PCR	B	II	Tang, Am Rev Respir Dis, 1993 Bretagne, JCM, 1995 Jones, J Clin Pathol, 1998 Skladny, JCM, 1999 Buchheidt, CID, 2001 Hayette, JCM, 2001 Buchheidt, Brit J Haem, 2002 Raad, Chest, 2002 Spiess, JCM, 2003 Meletiadis, Med Mycol, 2003 Sanguinetti, JCM, 2003 Rantakokko, JCM, 2003 Lass-Flörl, JCM, 2004 Musher, JCM, 2004 Khot, BMC Inf Dis, 2008 Frealle, EJClinMicrobInfDis, 2009 Bergeron, JCM, 2011 Luong, Transpl 2011 Buess, BMC Inf Dis, 2012 Reinwald, Eur J Hematol, 2012 Reinwald, JAC, 2012 Hönigl et al, JCM 2014	Methodically different in-house assays; better performance in pts without AFT; PCR+GM: increases specificity



Clinical Practice Patterns in Hospitalized Patients at Risk for Invasive Candidiasis: Role of Antifungal Stewardship Programs in an Era of Rapid Diagnostics

Setting

- Prospective cohort study
- Patients with candidemia or receiving systemic antifungals
- University-affiliated tertiary care hospital

Endpoints

- Time to initiation of therapy
- *Candida* species and time to identification
- Indications for antifungal use



Clinical Practice Patterns in Hospitalized Patients at Risk for Invasive Candidiasis: Role of Antifungal Stewardship Programs in an Era of Rapid Diagnostics

Table 1. Sensitivity and Specificity of Rapid Diagnostic Tests.

Rapid Diagnostic	<i>Candida</i> Species Detected	<i>Candida</i> Detected in Current Study (%)	Date Test is Performed	Sensitivity	Specificity	Positive Predictive Value
T2Candida assay	<i>C. albicans</i> , <i>C. parapsilosis</i> , <i>C. krusei</i> , <i>C. glabrata</i> , <i>C. tropicalis</i>	98%	Day of blood culture	98%	98%	91% ^a
MALDI-TOF MS	All ^b	100%	Day of yeast identification	94%	100%	94% ^c
PNA-FISH	<i>C. albicans</i> / <i>C. parapsilosis</i> (probe 1), <i>C. glabrata</i> / <i>C. krusei</i> (probe 2), <i>C. tropicalis</i> (probe 3)	98%	Day of yeast identification	98%	98%	99% ^c

Abbreviations: MALDI-TOF, matrix-assisted laser desorption/ionization time of flight; PNA-FISH, fluorescence in situ hybridization using peptide nucleic acid probes.

^aAssume an 18% rate of occurrence of candidemia multiplied by percentage of *Candida* detected.

^bMALDI-TOF was assumed to identify all clinically relevant *Candida* species.

^cAssume a 100% rate of occurrence of candidemia multiplied by percentage of *Candida* detected.



Clinical Practice Patterns in Hospitalized Patients at Risk for Invasive Candidiasis: Role of Antifungal Stewardship Programs in an Era of Rapid Diagnostics

Results

- N=162 patients with candidemia
- Average time to yeast identification: 2.2 ± 1.3 days
- Average time to start of antifungal therapy: 3.5 ± 2.1 days
 - 0.6 ± 0.2 days for T2Candida
 - 2.6 ± 1.3 days for PNA-FISH (peptide nucleic acid probes)
 - 2.5 ± 1.4 days for MALDI-TOF
- T2Candida in simulation resulted in fewer doses of echinocandins

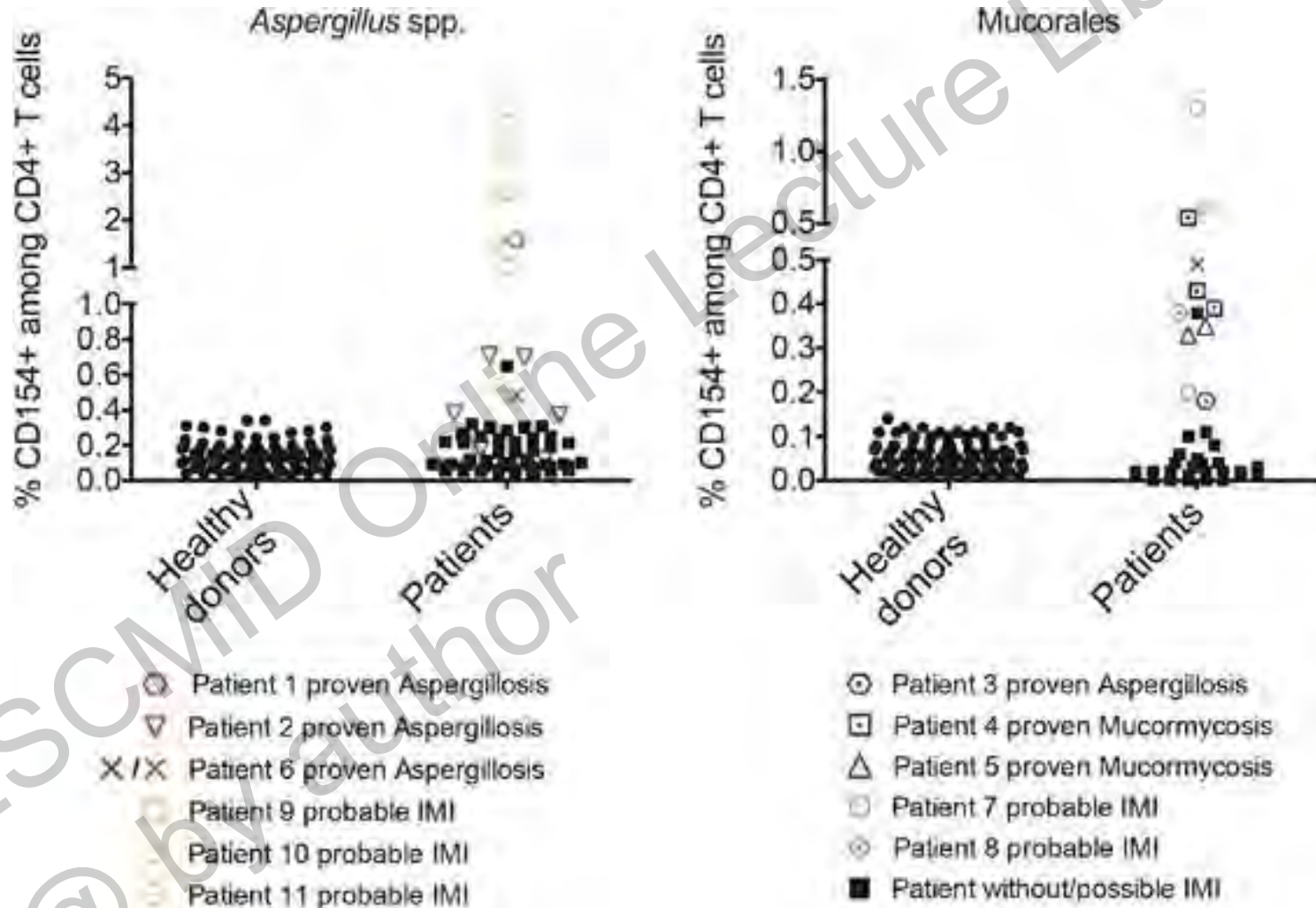


Turning to host response instead of fungal molecules

- T cells as specific diagnostic sensors for invasive fungal infections
- Monitor mold-reactive CD154+ peripheral blood T cells
- Pilot study completed

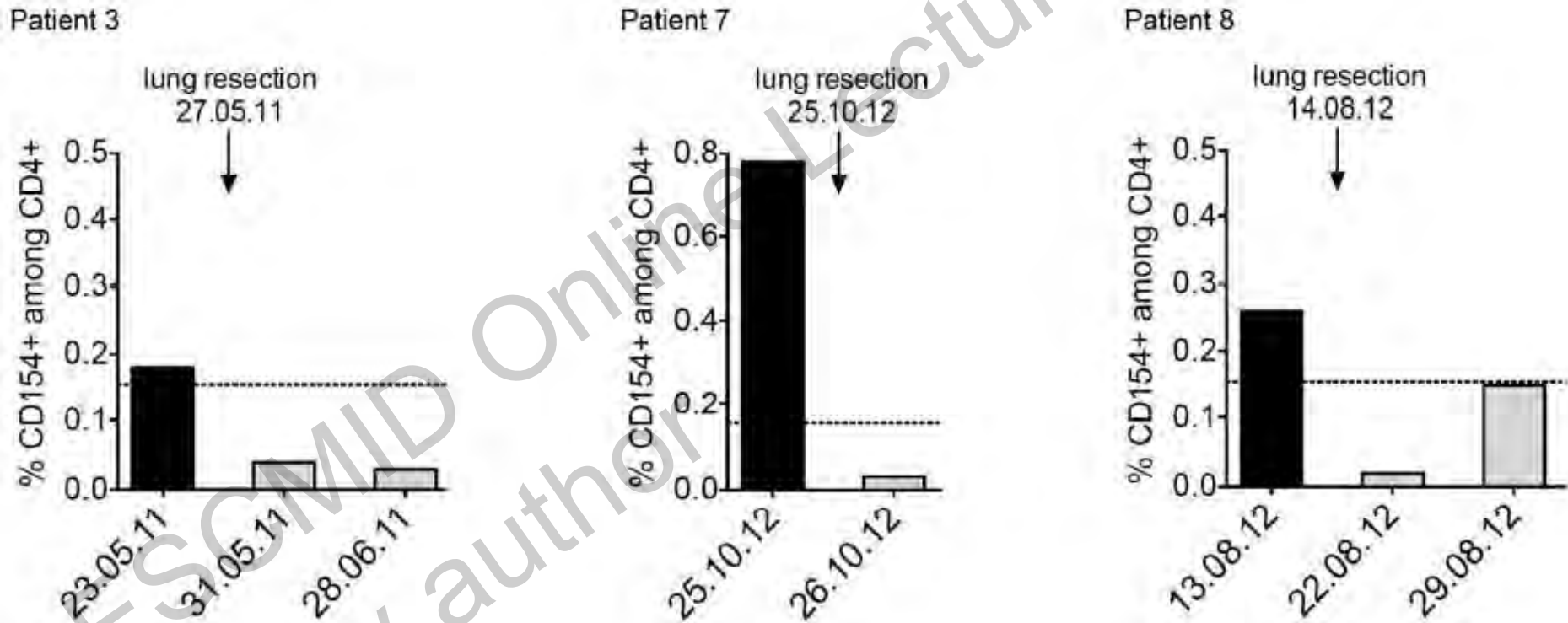


Frequencies of fungus-reactive T cells





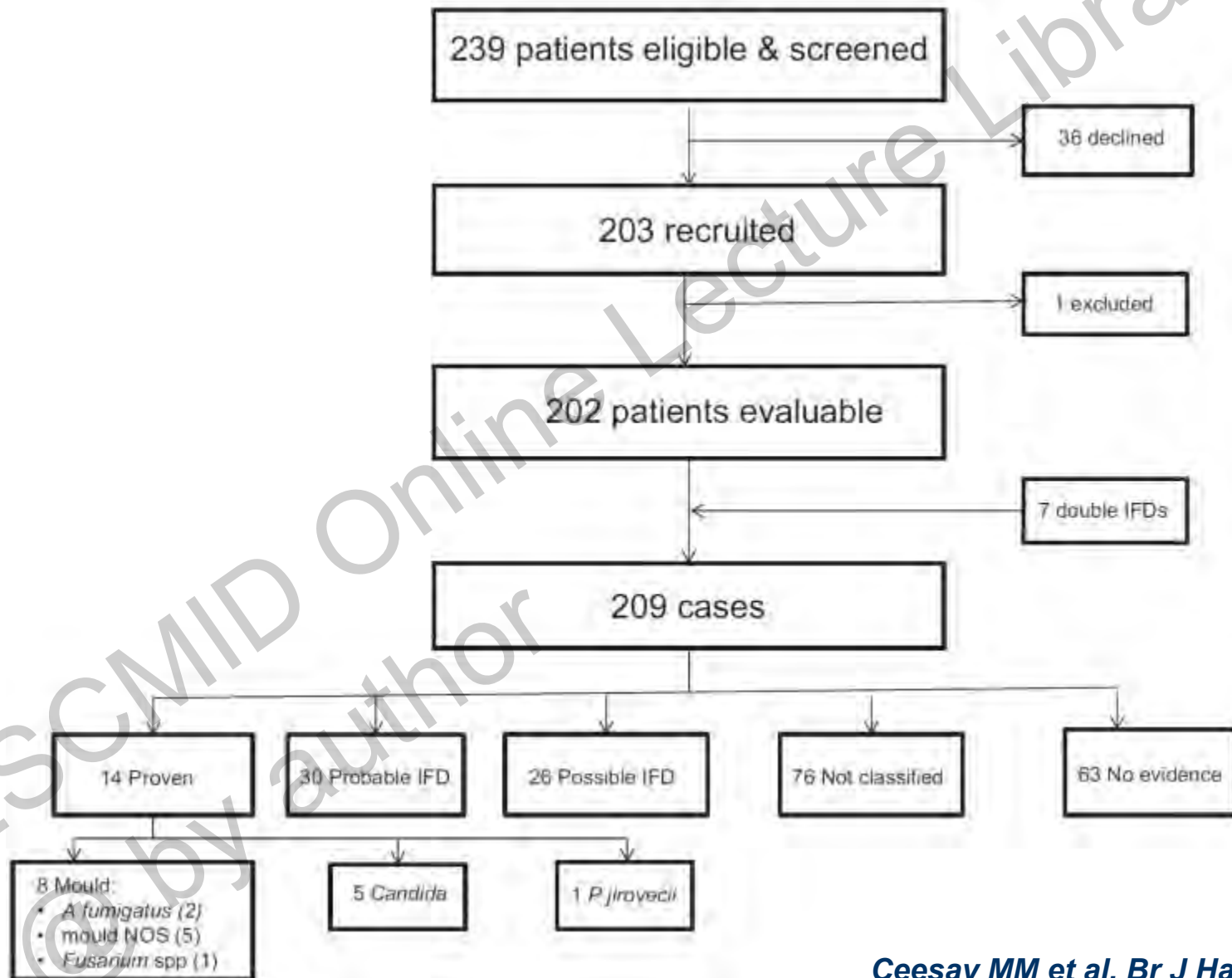
Mold-reactive T cell frequencies and fungal burden in 3 patients with invasive mold infection





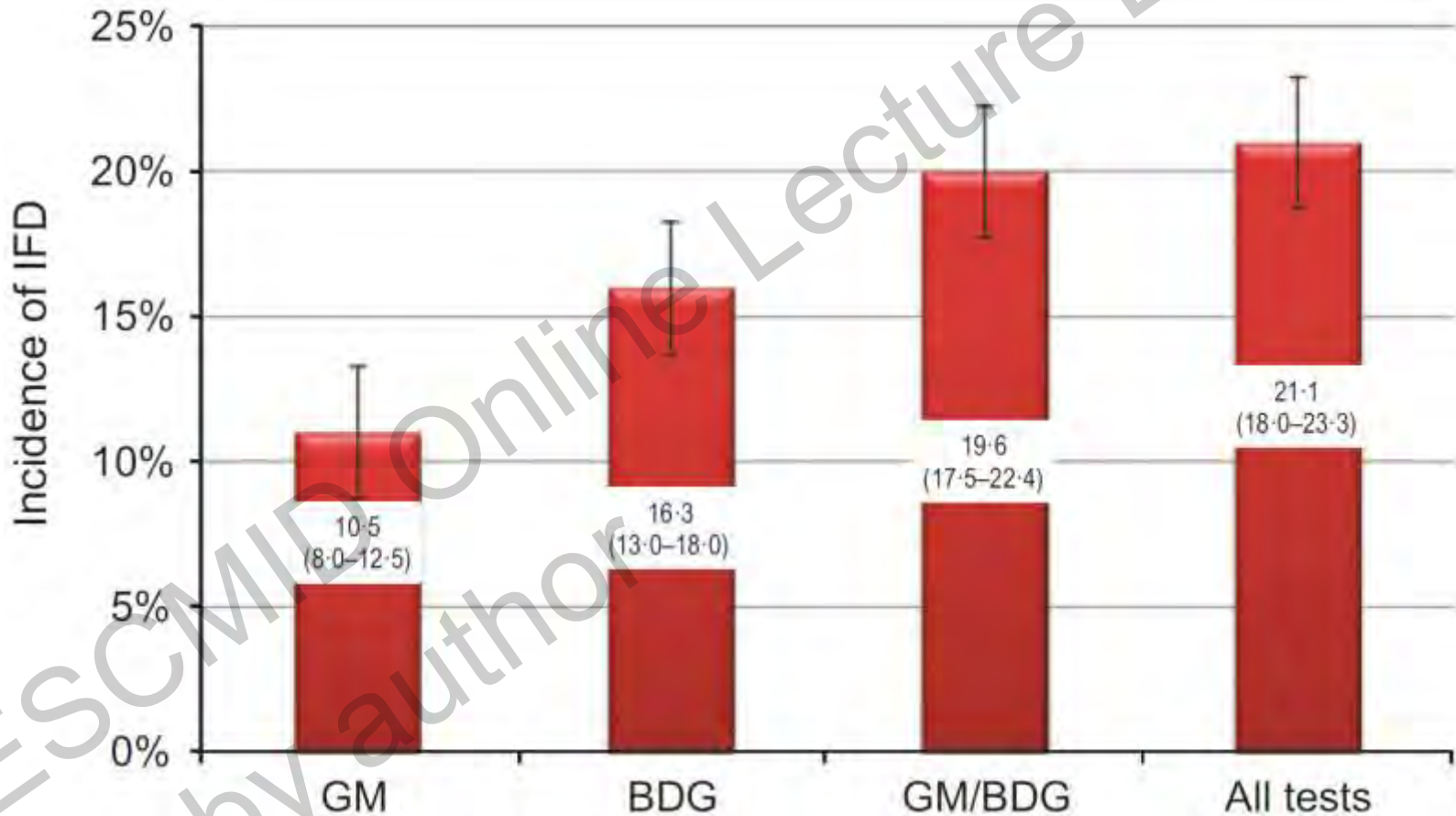
A comprehensive diagnostic approach using galactomannan, targeted β -D-glucan, baseline computerized tomography and biopsy yields a significant burden of invasive fungal disease in at risk haematology patients

- N=203
- Intensive therapy
- Expected neutropenia ≥ 10 d
- Prospective, F/U median (range) of 556 (12–730) d
- Baseline CT, GM biw, targeted BDG in possible IFD or when GM positive





Incidence (95% CI) of proven/probable IFD by tools used



= GM, BDG, CT, biopsy, blood cultures, BAL, autopsy



- Positive GM should trigger immediate diagnostic work-up
- Negative GM is a pre-requisite, but not sufficient for stopping treatment
- Positive BDG should not trigger treatment
- Negative BDG should be used for stopping empiric Candida-directed treatment

Both tests should ideally be used in the context of clinical judgement, other IVD assays, and imaging studies

Institutional algorithms should be informed by ESCMID-ECMM guidance.

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