

HCV diagnosis and monitoring in the era of direct-acting antiviral agents

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University of Nottingham

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Material to be covered

- Diagnosis
 - Including sampling
- Pre-Treatment Assessment
- On-treatment Monitoring
 - Stopping rules
 - Response Guided Therapy

Changing therapies

- Evolution of different regimens
 - PEG RV
 - PEG RV PI 1st gen
 - PEG RV DAAs
 - DAAs +/- RV
- Has led to evolution of laboratory requirements

Diagnosis

- Venous blood sample
- Dried blood spot



Freepost: RRL-SLJH-H8KB
Health Protection Agency
Clinical Sciences Building
Oxford Road
MANCHESTER
M13 9WL



Form with a grid and various sections. Visible text includes:

- Cellulose Pasteurized
- Biological Control for Infections (if not done above)
- Alcohol Swab
- 70% Isopropyl Alcohol
- FOR EXTRAORAL USE ONLY

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Blood Spot Sample Collection

1) Disposable gloves should be worn for the whole procedure to protect you and to protect the sample from degradation.

2) Ensure the patients hands are clean. If the hands are cold ask the patient to rub them together. It is important the puncture site is warm.

3) Identify and clean the puncture site:

- Use one of the outer 3 fingers
- Avoid finger pad and nail bed
- Clean site with alcohol wipe then dry



4) Break the Seal by Twisting.



5) Hold the lancet with two fingers and place at the puncture site.



6) Gently apply pressure until the lancet is activated. A click sound will be heard when this has happened.

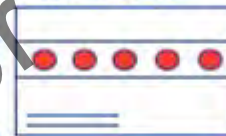


7) Safely discard the lancet in a sharps container.



8) Spot one blood drop in each of the 5 circles on the card. To ensure enough blood flow:

- Hold the puncture site down
- Apply intermittent pressure
- Avoid strong repetitive pressure (milking). It can damage the area.



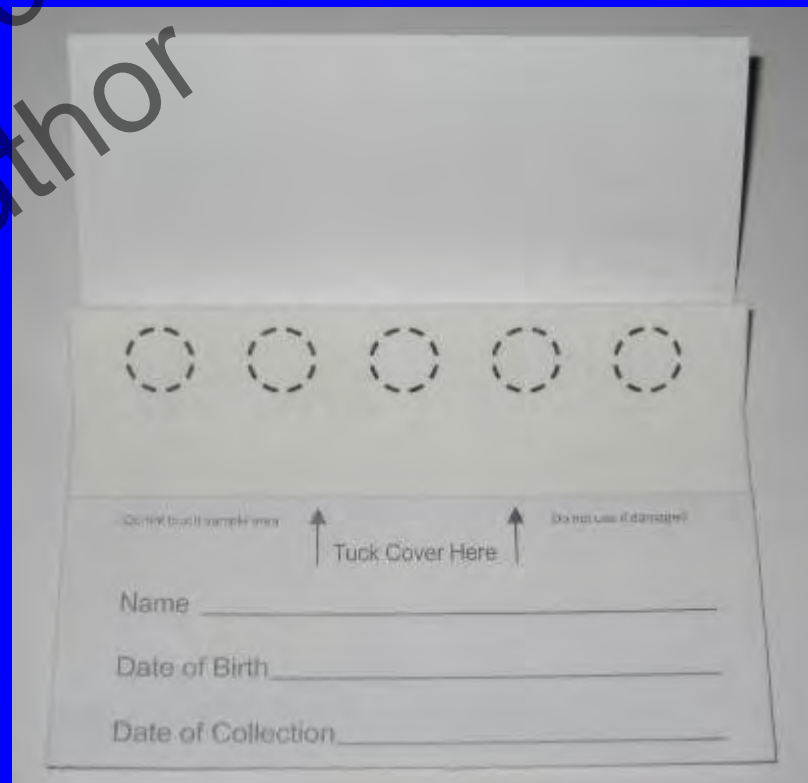
9) Leave the card to dry for a minimum of 2 hours.

10) Place the card along with one bag of desiccant in the plastic bag attached to the request form.

11) Check the request form and the dried blood spot card have had all the required details entered and place the card in the envelope provided and return to the laboratory.



REMOVE COVERING STRIP
PLACE SPECIMEN IN BAG
FOLD TOP OVER TO SEAL



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What happens in the lab

- Dried blood spot – containing paper immersed in buffer
- Elution overnight
- Eluate handled as per VBS (diluted)
- Anti-HCV testing
- HCV RNA testing

DBS Pros and Cons

- Allows sampling in settings where VBS may not be possible
- Patient preference
- Easy transport of samples to lab
- Can test for other analytes eg HBsAg, anti-HIV
- Extra cost in processing
- EIA needs to be recalibrated
- Less sensitive – HCV RNA > 1000 iu/ml

Anti-HCV POSITIVE

- Evidence of infection at some time
- Gives no indication as to when infection occurred
- Gives no indication as to whether infection was cleared or is still present

Virus Detection

- Genome – e.g. Reverse Transcriptase
PCR
 - More expensive, more sensitive
- Core Antigen
 - Less expensive, less sensitive

Diagnosis

- Anti-HCV positive
- HCV RNA (or antigen) positive
- = current infection therefore eligible for therapy

Anti-HCV: Negative

- No evidence of infection with HCV
- BUT - be aware of possible false negatives
 - if infection very recent (window period)
 - if patient immunosuppressed at time of infection
- ? Role for HCV RNA as primary test
- ? Other modes of diagnosis of acute infection

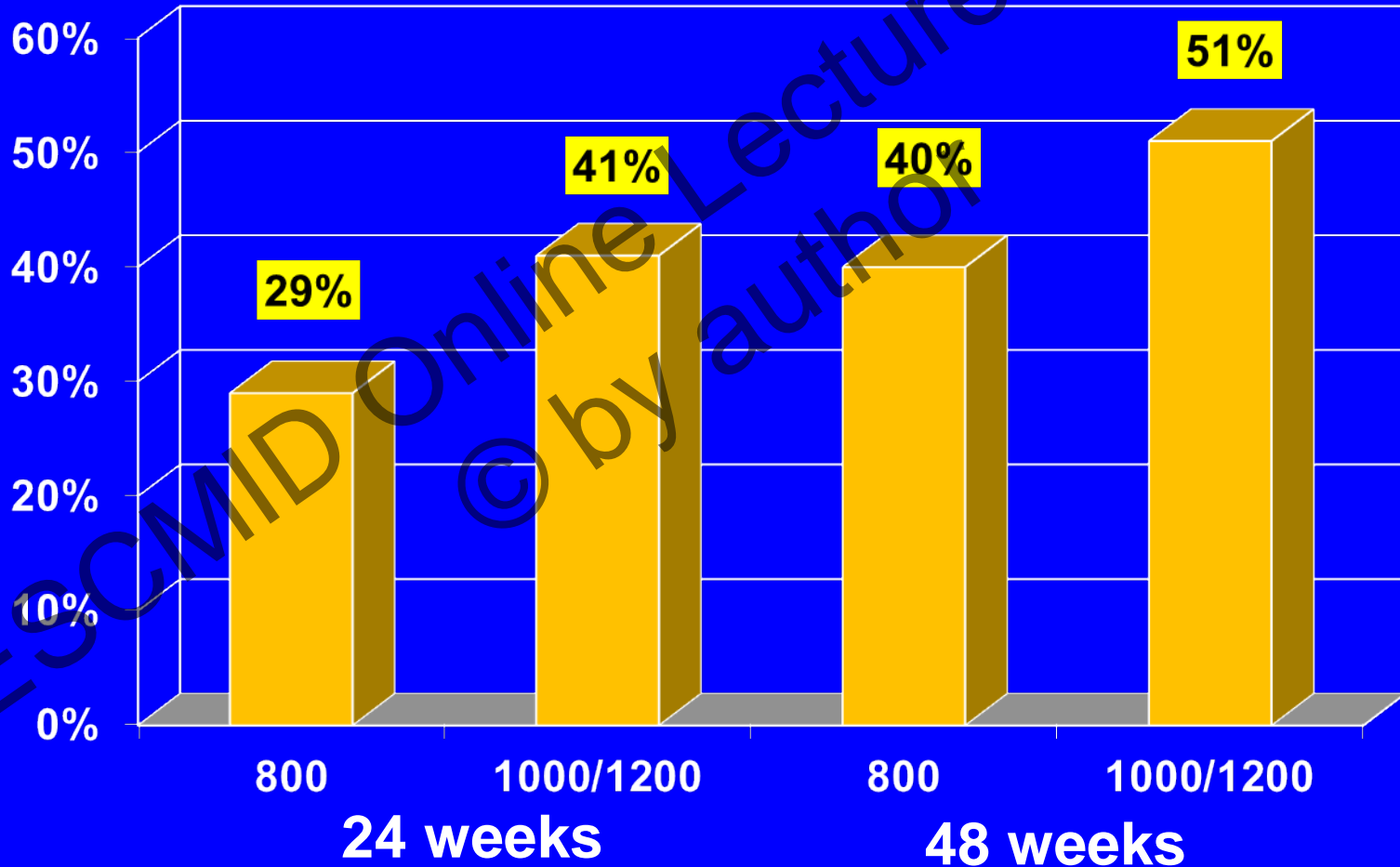
Pre-Treatment assessment

- Genotype

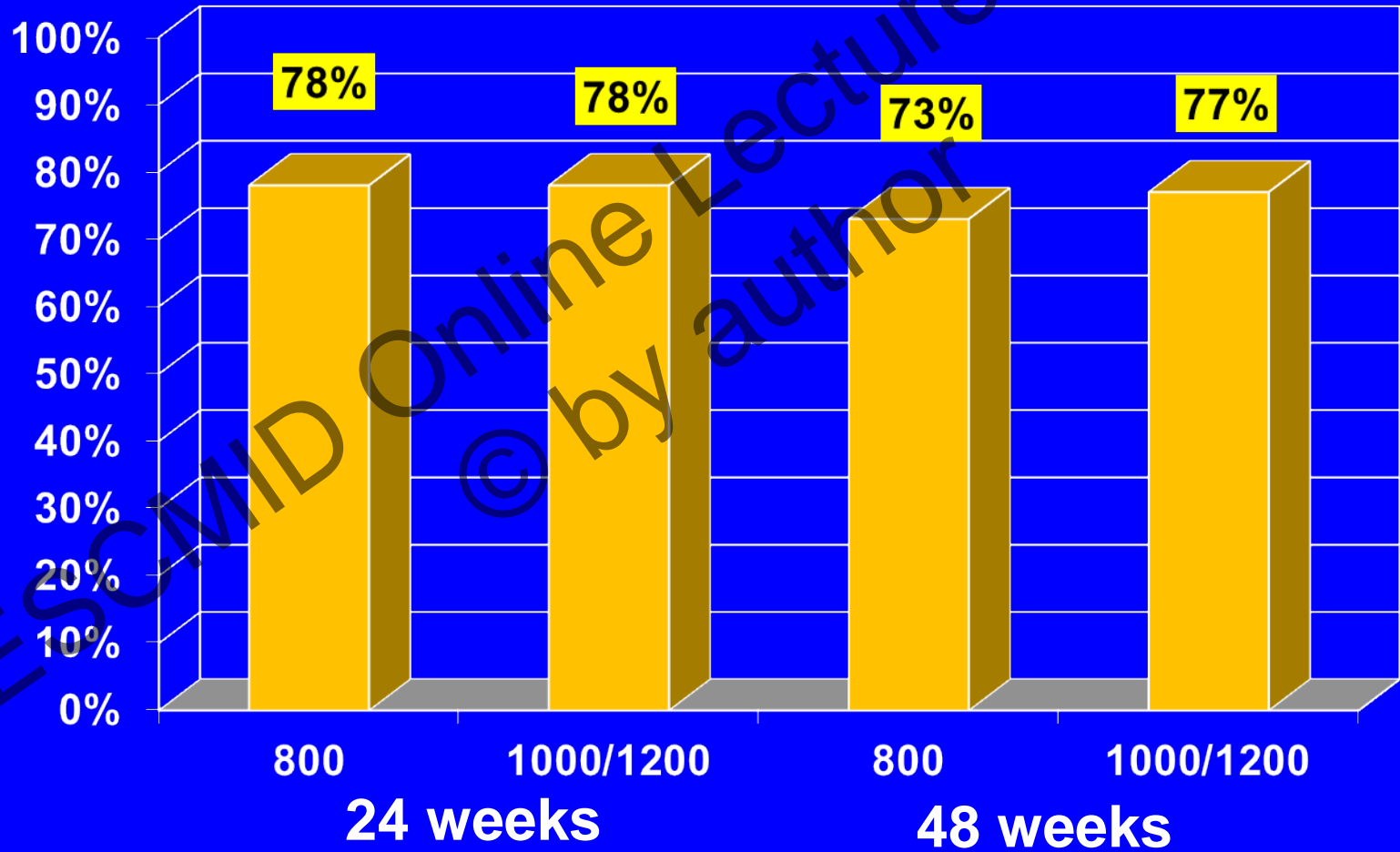


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PEG-IFN alpha2a & Ribavirin Genotype 1



PEG-IFN alpha2a & Ribavirin Genotype non-1 (mainly 2 & 3)



Activity of protease inhibitors by HCV genotype

Genotype 1

Boceprevir^[1,2]

Telaprevir^[3,4]

BI 201335^[5]

Danoprevir^[6]

MK-5172^[7]

TMC435^[8]

Vaniprevir^[9]

1. Poordad F, et al. AASLD 2010. Abstract LB-4; 2. Pawlotsky JM, et al. Gastroenterology 2011 [epub ahead of print]. Abstract 820; 3. Jacobson IM, et al. AASLD 2010. Abstract 211; 4. Foster G, et al. EASL 2010. Abstract 57; 5. Sulkowski M, et al. EASL 2010. Abstract 1190; 6. Terrault N, et al. AASLD 2010. Abstract 32; 7. Brainard DM, et al. AASLD 2010. Abstract 807; 8. Fried M, et al. AASLD 2010. Abstract LB-5; 9. Manns MP, et al. AASLD 2010. Abstract 82.
Zeuzem S. CCO slide set

Activity of protease inhibitors by HCV genotype

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Boceprevir^[1,2]
Telaprevir^[3,4]
BI 201335^[5]
Danoprevir^[6]
MK-5172^[7]
TMC435^[8]
Vaniprevir^[9]

Genotype 2

Boceprevir^[1,2]
Telaprevir^[3,4]
BI 201335^{[5]?}
MK-5172^[7]
TMC435^[8]
Vaniprevir^{[9]?}

1. Poordad F, et al. AASLD 2010. Abstract LB-4; 2. Pawlotsky JM, et al. Gastroenterology 2011 [epub ahead of print]. Abstract 820; 3. Jacobson IM, et al. AASLD 2010. Abstract 211; 4. Foster G, et al. EASL 2010. Abstract 57; 5. Sulkowski M, et al. EASL 2010. Abstract 1190; 6. Terrault N, et al. AASLD 2010. Abstract 32; 7. Brainard DM, et al. AASLD 2010. Abstract 807; 8. Fried M, et al. AASLD 2010. Abstract LB-5; 9. Manns MP, et al. AASLD 2010. Abstract 82.
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Genotype 2

Boceprevir^[1,2]
Telaprevir^[3,4]
BI 201335^{[5]?}
MK-5172^[7]
TMC435^[8]
Vaniprevir^{[9]?}

Genotype 4

Danoprevir^[6]
MK-5172^[7]
TMC435^[8]

1. Poordad F, et al. AASLD 2010. Abstract LB-4; 2. Pawlotsky JM, et al. Gastroenterology 2011 [epub ahead of print]. Abstract 820; 3. Jacobson IM, et al. AASLD 2010. Abstract 211; 4. Foster G, et al. EASL 2010. Abstract 57; 5. Sulkowski M, et al. EASL 2010. Abstract 1190; 6. Terrault N, et al. AASLD 2010. Abstract 32; 7. Brainard DM, et al. AASLD 2010. Abstract 807; 8. Fried M, et al. AASLD 2010. Abstract LB-5; 9. Manns MP, et al. AASLD 2010. Abstract 82.
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Activity of protease inhibitors by HCV genotype

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MK-5172⁷
TMC435⁸
Vaniprevir⁹

Genotype 2

Boceprevir^{1,2}
Telaprevir^{3,4}
BI 201335⁵?
MK-5172⁷
TMC435⁸
Vaniprevir⁹?

Genotype 4

Danoprevir⁶
MK-5172⁷
TMC435⁸

Genotypes 5, 6

MK-5172⁷
TMC435⁸

Genotype 3

MK-5172⁷

1. Poordad F, et al. AASLD 2010. Abstract LB-4; 2. Pawlotsky JM, et al. Gastroenterology 2011 [epub ahead of print]. Abstract 820; 3. Jacobson IM, et al. AASLD 2010. Abstract 211; 4. Foster G, et al. EASL 2010. Abstract 57; 5. Sulkowski M, et al. EASL 2010. Abstract 1190; 6. Terrault N, et al. AASLD 2010. Abstract 32; 7. Brainard DM, et al. AASLD 2010. Abstract 807; 8. Fried M, et al. AASLD 2010. Abstract LB-5; 9. Manns MP, et al. AASLD 2010. Abstract 82.
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Pre-Treatment assessment

- Genotype
 - PEG + RV
 - PEG + RV + PIs
- Sub-genotyping

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Clinical Implications of Genetic Barrier to Resistance – Acquisition of NS3 Inhibitor Resistant Variant V36M+R155K

Subtype 1a

V36M+R155K variant observed clinically^{1,2}



2 steps

Subtype 1b

V36M+R155K variant not observed clinically



4 steps

PIs: Resistance

- May be inherent i.e. not all genotypes are sensitive
- Genetic barriers vary according to subtype
 - How good are our typing assays?
 - J Clin Virol 2014; 60: 301-304
 - 1052 samples genotyped
 - 54 samples reported only as “genotype 1”
 - Sequencing resolved into 1a or 1b

Pre-Treatment assessment

- Genotype
 - PEG + RV
 - PEG + RV + PIs
- Sub-genotyping
 - PEG + RV + PIs
- Strain typing

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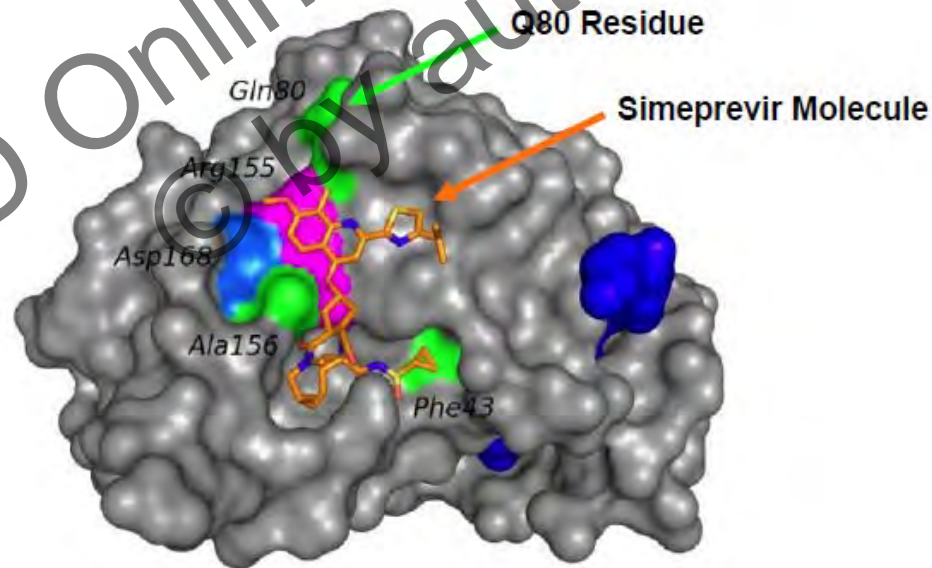
Q80K – individualising treatment

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Naturally occurring polymorphism in the viral amino acid sequence

Q 80 K

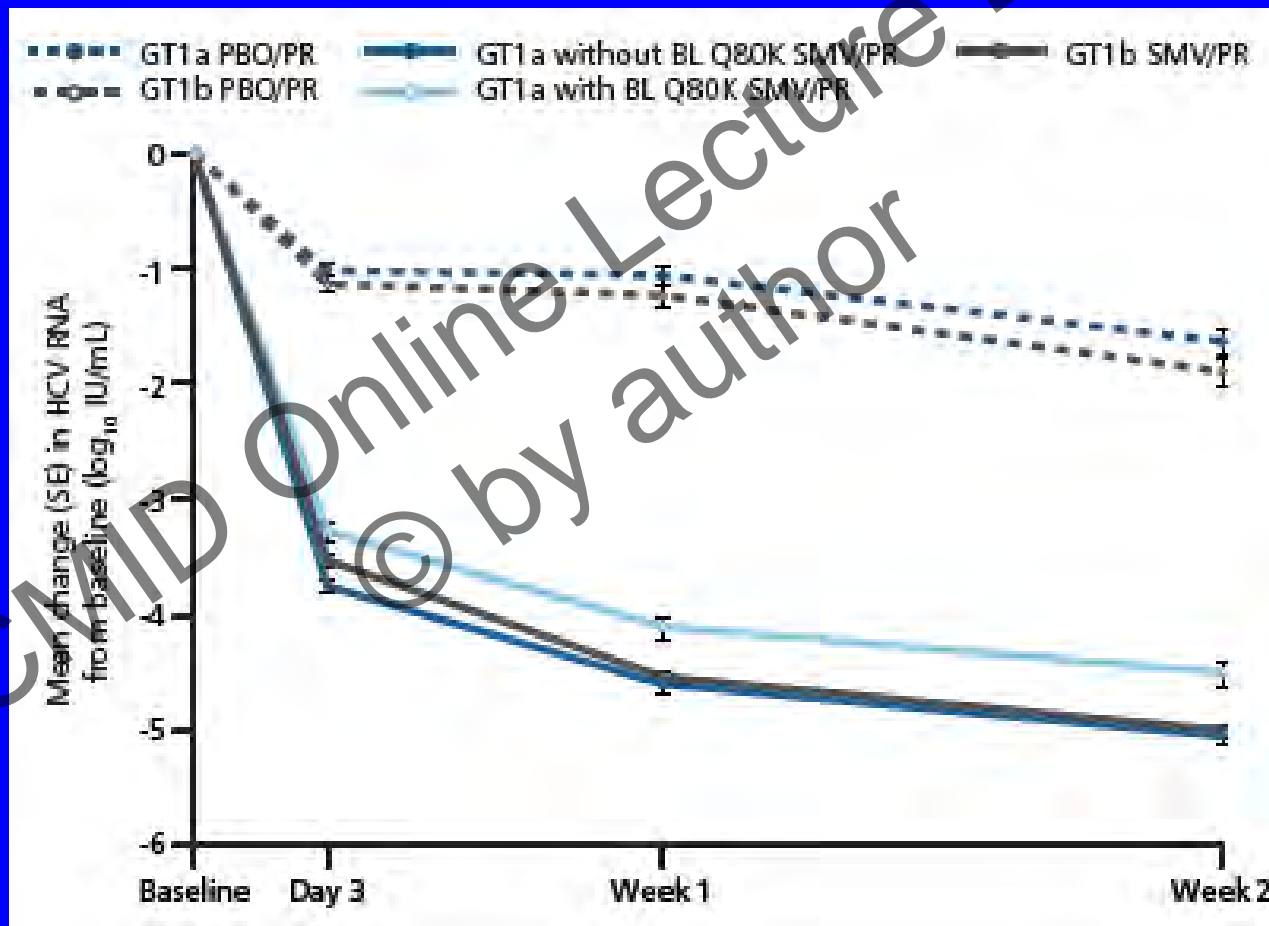
Wild-type amino acid *Amino acid position* *Mutated amino acid*



The Q80K polymorphism is less prevalent in Europe compared to the US

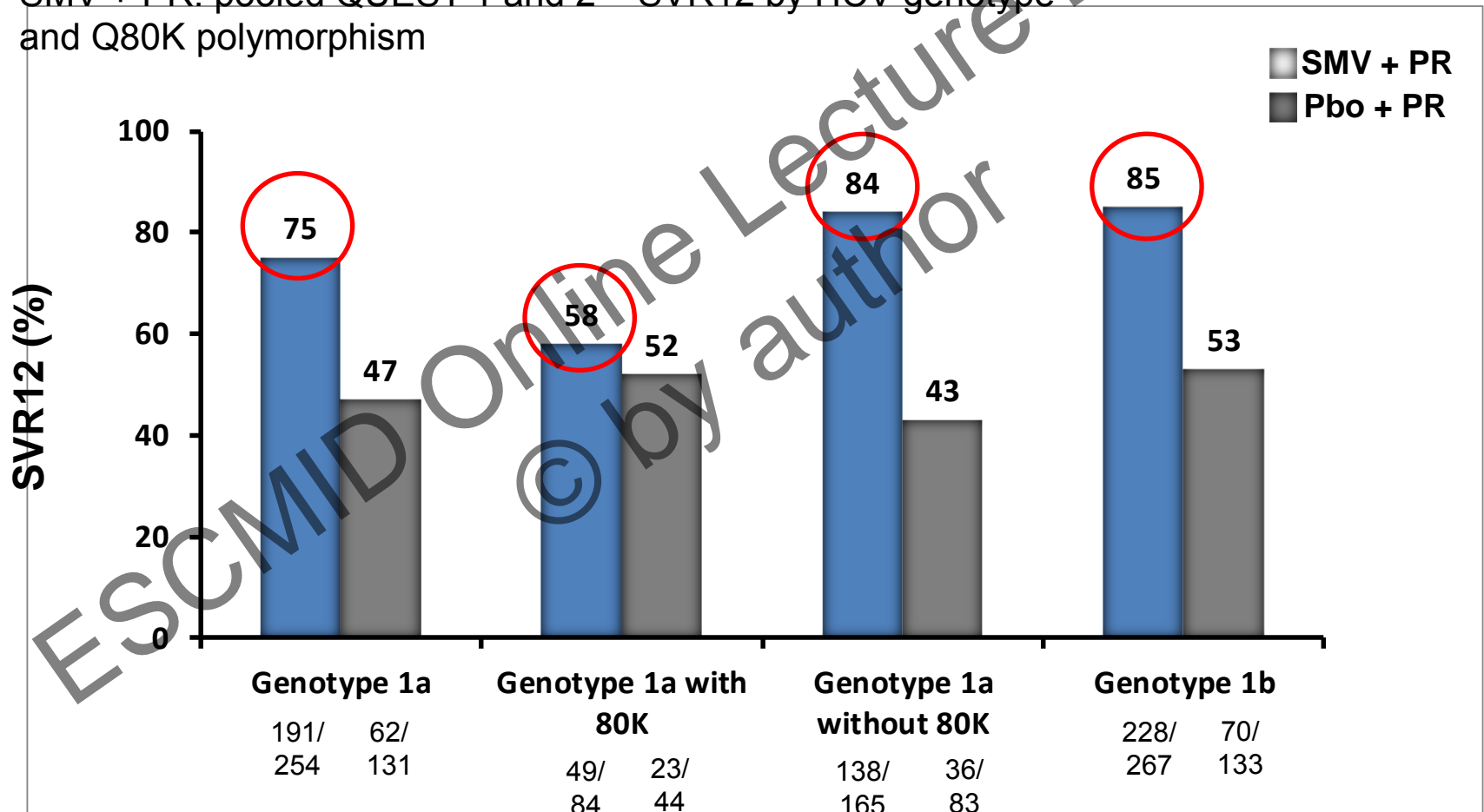
n/N (%)	All HCV genotype 1	HCV genotype 1a	HCV genotype 1b
Overall	274/2007 (13.7)	269/911 (29.5)	5/1096 (0.5)
Europe	76/1254 (6.1)	73/377 (19.4)	3/877 (0.3)
North America	185/538 (34.4)	185/385 (48.1)	0/153 (0)
South America	2/60 (3.3)	2/22 (9.1)	0/38 (0)

Early Viral Decay and Q80K



Q80K is a predictor of response: Treatment Naive

SMV + PR: pooled QUEST 1 and 2 – SVR12 by HCV genotype and Q80K polymorphism



How to test for Q80K?

Process for Q80K Polymorphism Test

The Q80K polymorphism test is strongly recommended as it is likely to be a predictor of response for treatment options for patients starting treatment. The test should be considered as a routine part of the workup for patients with Hepatitis C Genotype 1.¹

Janssen understands that the cost and logistics of providing this test may be challenging for some hospitals and are committed to minimising the potential impact of providing this test.

If you decide to test for Q80K Polymorphism in G1 patients, please complete the Janssen T&C form, provided by the Janssen Account Manager. You can use the following process to access the test:



- The provision of the Q80K test by Janssen does not require you to use any of the company's products or services.
- Janssen will cover the costs of the Q80K test for use in NHS treated patients only.
- The tests being requested are for patients with Hepatitis C Genotype 1 and samples are to be sent to one of the laboratories from the approved list.
- Janssen will cover the costs of the testing for patients with Genotype 1 Hepatitis C but not the cost of sample transportation. The responsibility for transporting the sample to the laboratory remains with the hospital. Details of how to send the sample are on the Sample Request Form.

Further information and sample request forms can be obtained via email on janssenukhepc@its.jnj.com or by contacting your local Janssen Account Manager.

Pre-Treatment assessment

- Genotype
 - PEG + RV
 - PEG + RV + PIs
- Sub-genotyping
 - PEG + RV + PIs
- Strain typing
 - PEG + RV + Simeprevir

Pre-Treatment assessment

- Genotype/Subtype/Q80K
- Viral load assessment
 - Pre-Rx viral load correlates with likelihood of achieving SVR

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On treatment monitoring

- Stopping rules
 - Identify patients in whom continuing therapy is very unlikely to result in SVR ∴ STOP

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Stopping rules PEG/RV:

Davis et al (Hepatology 2003;38:645)

- Used patients from Manns et al Lancet 2001
- Tested a variety of definitions of EVR
 - 4/12 weeks
 - 1/2/3 log decrease, PCR neg
- Which definition captured highest no. SVRs (sensitivity), whilst excluding highest no. NRs (NPV)?

SVRs (n=273)

NRs (n=238)

EVR	No EVR	EVR	No EVR
1-log drop at 12 weeks			
273 (100%)	0	144	94 (38%)
2-log drop at 12 weeks			
273 (100%)	0	107	131 (55%)
1-log drop at 4 weeks			
267 (97.8%)	6	113	125 (52.5%)
2-log drop at 4 weeks			
250 (92%)	23	70	168 (71%)

SVRs (n=273)

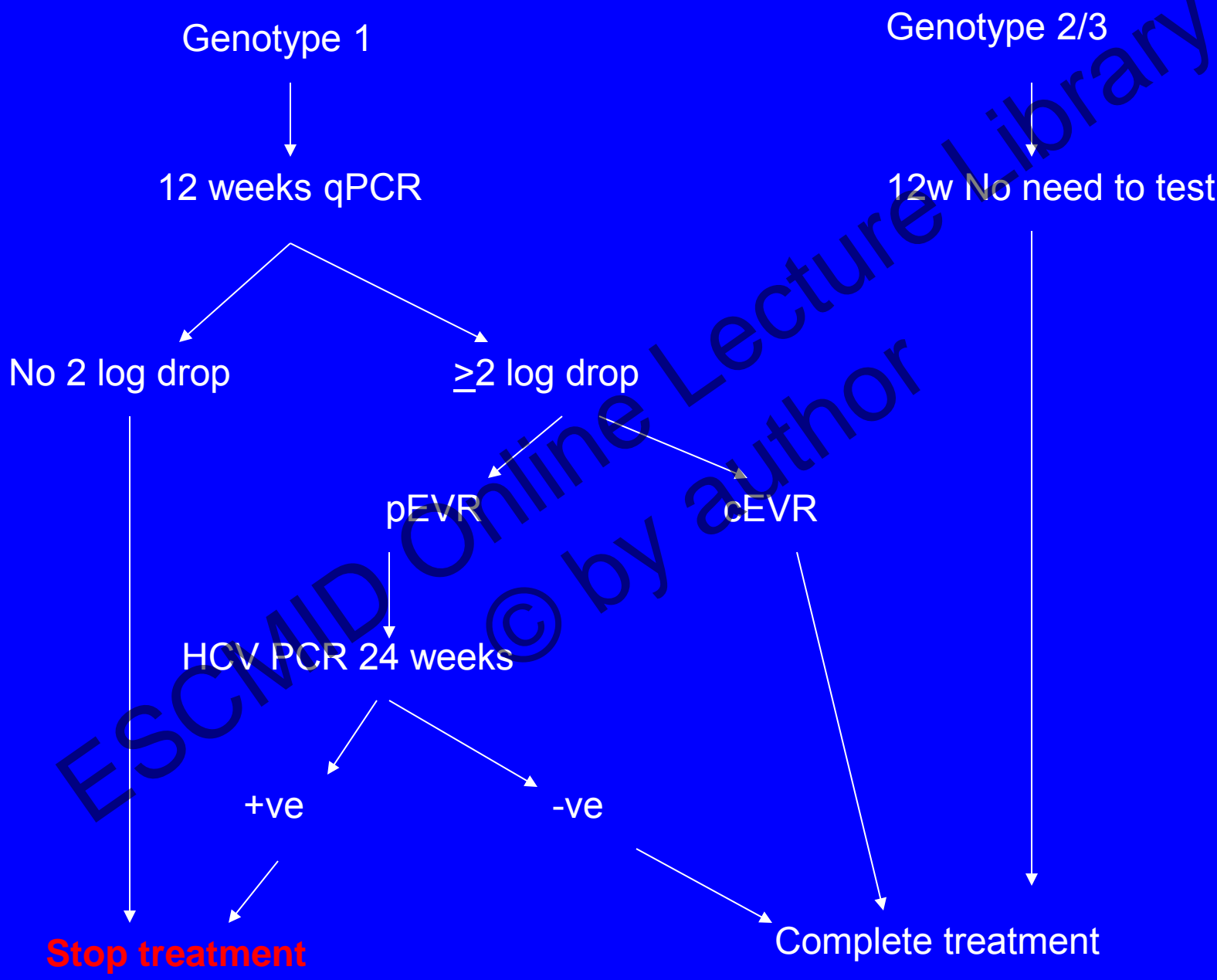
NRs (n=238)

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267 (97.8%)	6	113	125 (52.5%)
2-log drop at 4 weeks			
250 (92%)	23	70	168 (71%)

On Rx monitoring: Definitions

- Early Virological Response (EVR)
 - Measured at 12 weeks on therapy
 - Complete (cEVR) = Negative
 - Partial (pEVR) = > 2 log drop
 - Non-response = ≤ 2 log drop

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On treatment monitoring

- Stopping rules
- Response Guided Therapy (RGT) rules
 - Stratify treatment regimen according to likelihood of achieving SVR i.e. *shorten* therapy
 - Based on 2 parameters
 - Kinetics of viral loss
 - Baseline viral load

On Rx monitoring: Definitions

- Early Virological Response (EVR)
- Rapid Virological Response (RVR)
 - Assessed at 4 weeks on Rx
 - HCV RNA negative = RVR

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What is HCV RNA negativity?

- Different assays have different sensitivities
 - Lower limit of quantification (LLQ)
 - <50? <30? <25? <15? <12? iu/ml
 - Signal may be detectable below LLQ
 - Lower limit of detection (LLD)
 - 9? 12? 15? etc etc
- Original description of RVR was with assays with LLQ around 25-30 iu/ml

Residual virus below LLQ

- Difficult to interpret lab result
 - ? Genuine low level viraemia
 - ? Background noise
- Impacts on likelihood of achieving SVR

Predictors of treatment failure

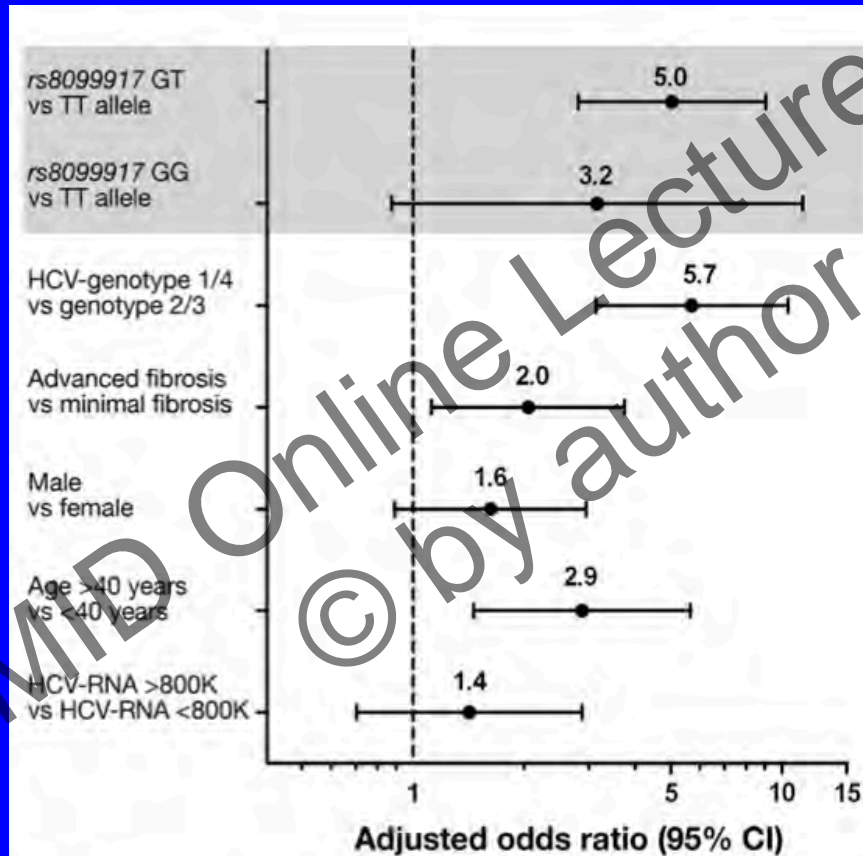


Figure 3. Predictors of failure to respond to pegylated IFN- α and ribavirin therapy. Carriers of the *rs8099917* G-risk genotypes had a higher risk of failing to respond to HCV treatment. ORs were calculated by allele and were adjusted for HCV genotypes, fibrosis stage, sex, age, baseline HCV viral load, and the first 2 ancestry principal components.

RVR vs no RVR

OR is around 7.0

RGT: PEG + RV

- Genotype 1 patients who achieve RVR and who had a low pre-Rx viral load – can shorten therapy from 12 to 6 months *without loss of SVR*
- Genotype 2/3 patients who achieve RVR and who had a low pre-Rx viral load – can shorten therapy from 24 to 16 weeks *without loss of SVR*
- ? How low is low?

On Rx monitoring: Definitions

- Early Virological Response (EVR)
- Rapid Virological Response (RVR)
- Extended Rapid Virological Response (eRVR)
 - Concept arose with 1st gen PIs
 - To take account of possible breakthrough of PI-resistant virus
 - RNA neg at 4 weeks *AND* 12 weeks of triple therapy = eRVR

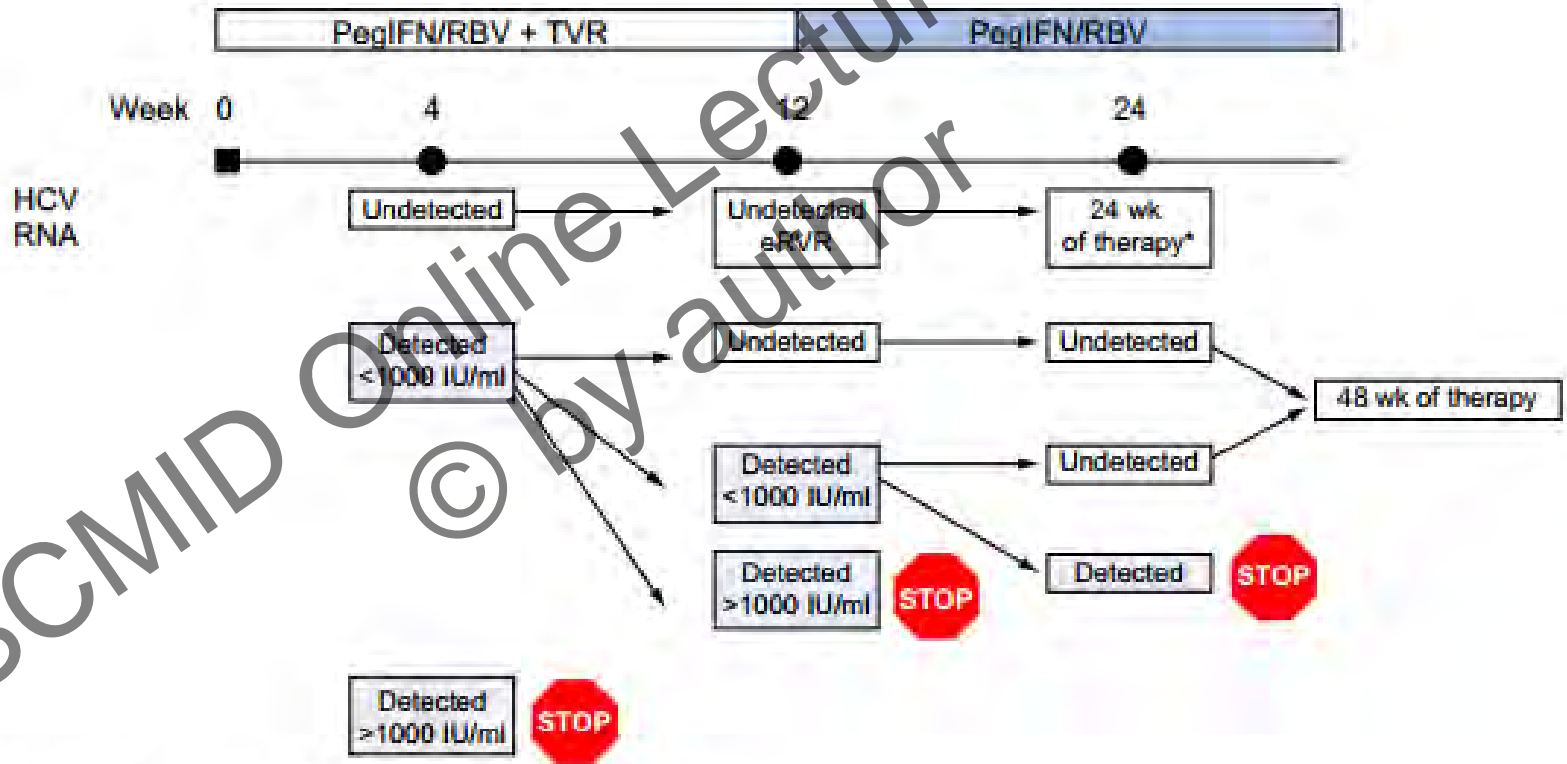
RGT: PEG + RV + TPR/BOC

- Shortened therapy (48 to 24 weeks) in g1 patients treated with triple therapy *if* eRVR was achieved

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Teleprevir: Stopping rules and RGT

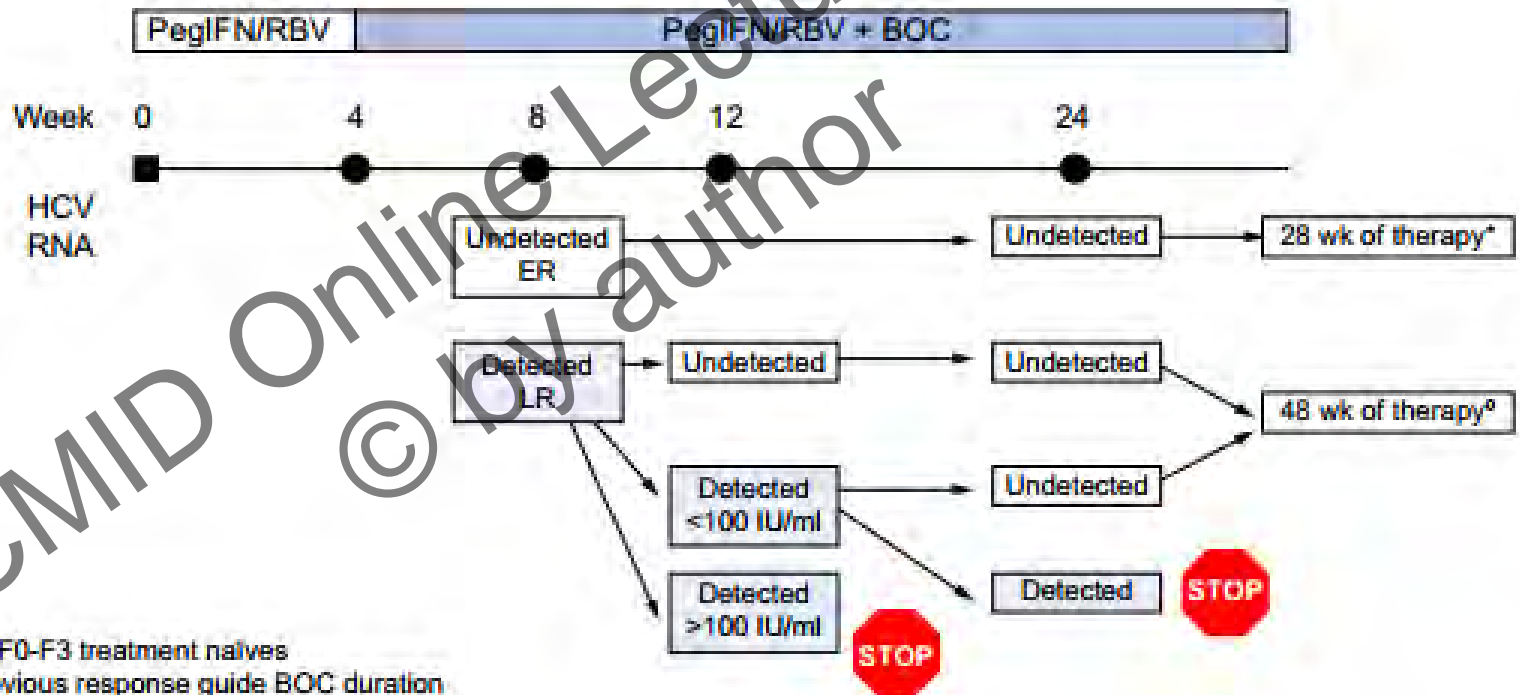
A



*only in fibrosis stage F0-F3 treatment naïves and relapsers

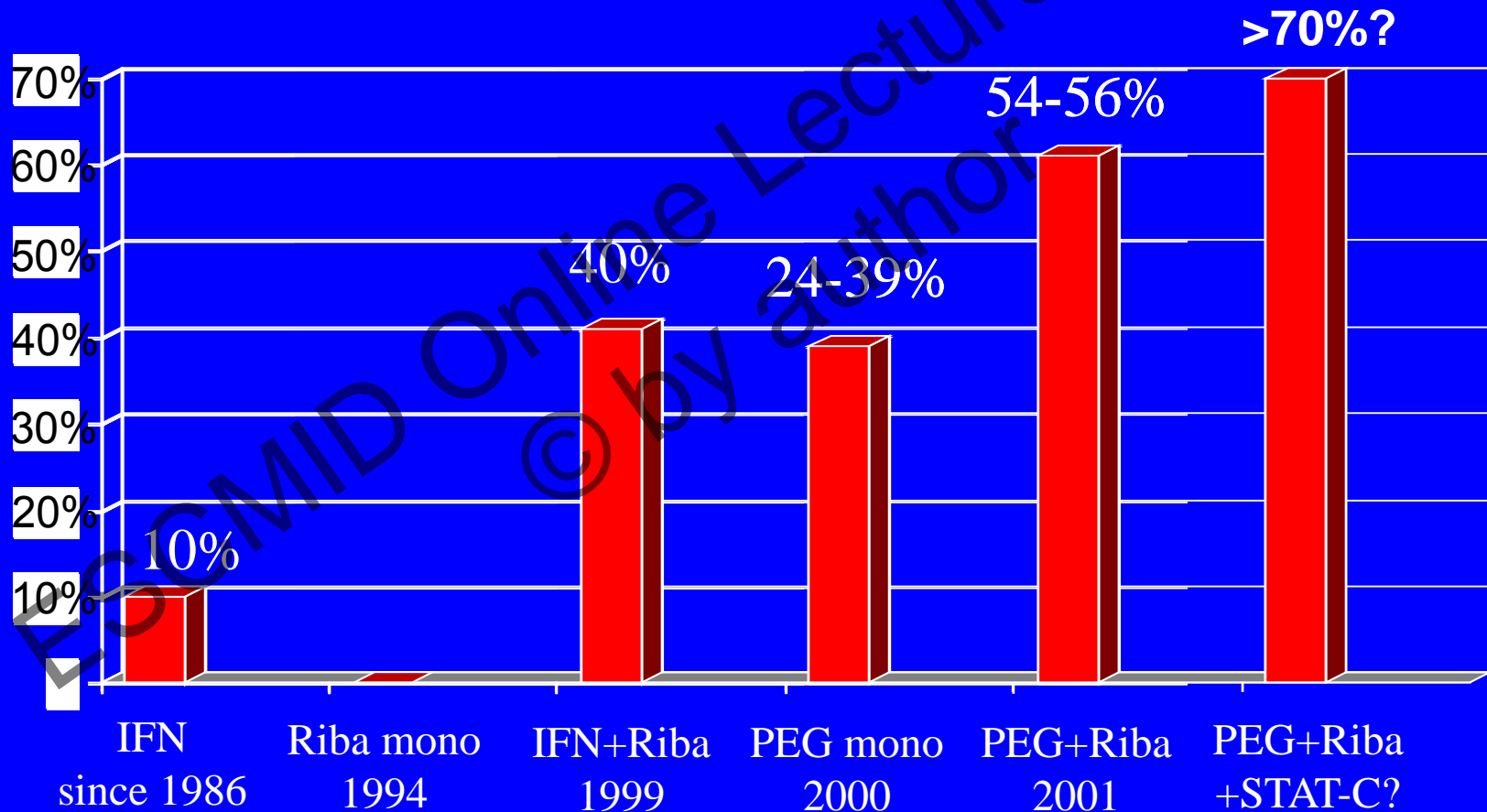
Boceprevir: Stopping rules and RGT

B



Treatment of Hepatitis C

Sustained Virologic Response (SVR): undetectable HCV RNA at least 6 months after the end of treatment



Hoofnagle et al. NEJM 1986

McHutchison et al., NEJM 1998

Manns et al., Lancet 2001

Brillanti et al. Gastroenterology 1994

Zeuzem et al., NEJM 2000

Fried et al., NEJM 2002

All Oral DAA therapy



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ORIGINAL ARTICLE

ORIGINAL ARTICLE

Ledipasvir and Sofosbuvir for Chronic HCV Genotype 1

Treatment of HCV with ABT-450/r-Ombitasvir and Dasabuvir with Ribavirin

Nezam Afdhal, M.D., Stefan Zeuzem, M.D., Paul Nelson, M.D., Norman Gitlin, M.D., Massimo Puoti, M.D., Manuel Romero-Gomez, M.D., Ph.D.,

Jordan J. Feld, M.D., M.P.H., Kris V. Kowdley, M.D., Eoin Coakley, M.D.,

The NEW ENGLAND JOURNAL of MEDICINE

ORIGINAL ARTICLE

ESTABLISHED IN 1812

APRIL 17, 2014

Ledipasvir and Sofosbuvir for Prevalent Genotype 1 Infection

Retreatment of HCV with ABT-450/r-Ombitasvir and Dasabuvir with Ribavirin

Nezam Afdhal, M.D., K. Rajender Reddy, M.D., David R. Nelson, M.D.,

Stefan Zeuzem, M.D., Ira M. Jacobson, M.D., Tolga Baykal, M.D.,

ORIGINAL ARTICLE

ORIGINAL ARTICLE

Ledipasvir and Sofosbuvir for 8 or 12 Weeks for Chronic HCV without Cirrhosis

ABT-450/r-Ombitasvir and Dasabuvir with Ribavirin for Hepatitis C with Cirrhosis

Kris V. Kowdley, M.D., Stuart C. Gordon, M.D., K. Rajender Reddy, M.D.,

Fred Poordad, M.D., Christophe Hezode, M.D., Roger Trinh, M.D., M.P.H.,

Stefan Zeuzem, M.D., Keshav Agrawal, M.D.,

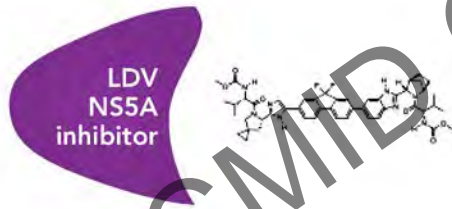
Lorenzo Boccia, M.D., David F. Bernstein, M.D., Eric Levitz, M.D.,

HARVONI[▼] is the first and only Single-Tablet Regimen^a for the majority of HCV GT1 adult patients^{1,b}

Excluding those with decompensated cirrhosis, or who are
pre- or post-liver transplant¹

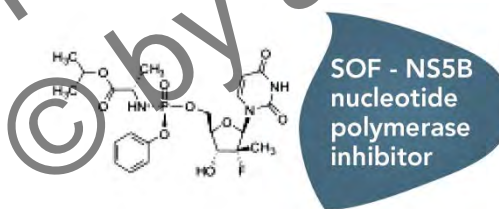
Ledipasvir

- Picomolar potency against HCV GT 1a and 1b¹
- Effective against NS5B RAV S282T²
- Once-daily, oral, 90 mg
- Half-life 47hrs²



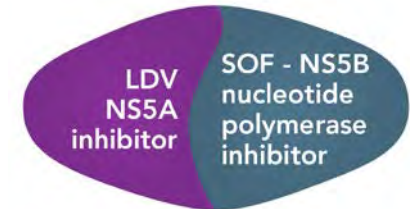
Sofosbuvir

- Potent antiviral activity against HCV GT 1–6
- Effective against NS5A RAVs³
- High barrier to resistance
- Once-daily, oral, 400 mg tablet
- Active metabolite half-life 27hrs³



Ledipasvir / sofosbuvir STR

- Once-daily, oral fixed-dose (90/400 mg) combination tablet
- No food effect
- 1,952 patients treated in the phase 3 programme



- Harvoni is indicated for the treatment of chronic hepatitis C (CHC) in adults¹ Genotypes 1, 3 & 4 (GT 3 + RBV)²

a. Single-tablet regimen is defined throughout this document as one tablet once daily

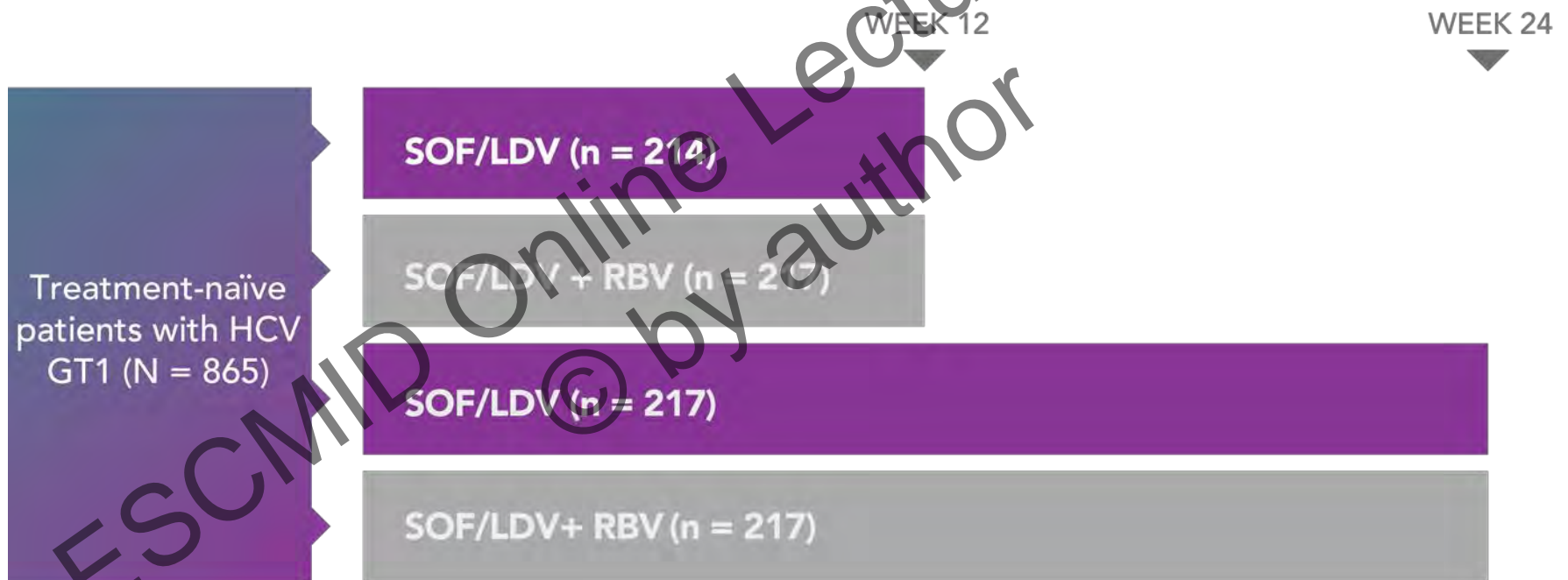
b. HARVONI offers a single-tablet, RBV-free regimen for the majority of HCV GT1 patients



Study Design – ION-11

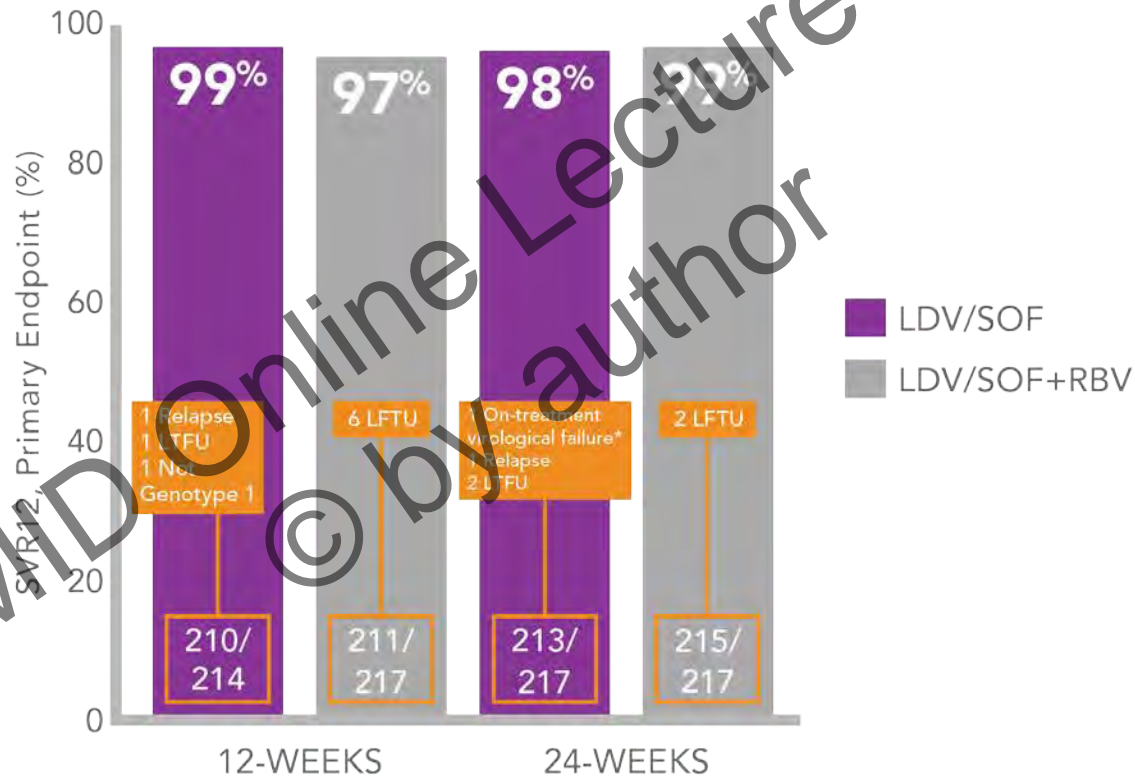
Randomised, open-label, Phase III trial in treatment-naïve patients with HCV GT1

Primary endpoints: SVR12



ION-1

Genotype 1 treatment naïve, including cirrhosis, 12 and 24 weeks¹

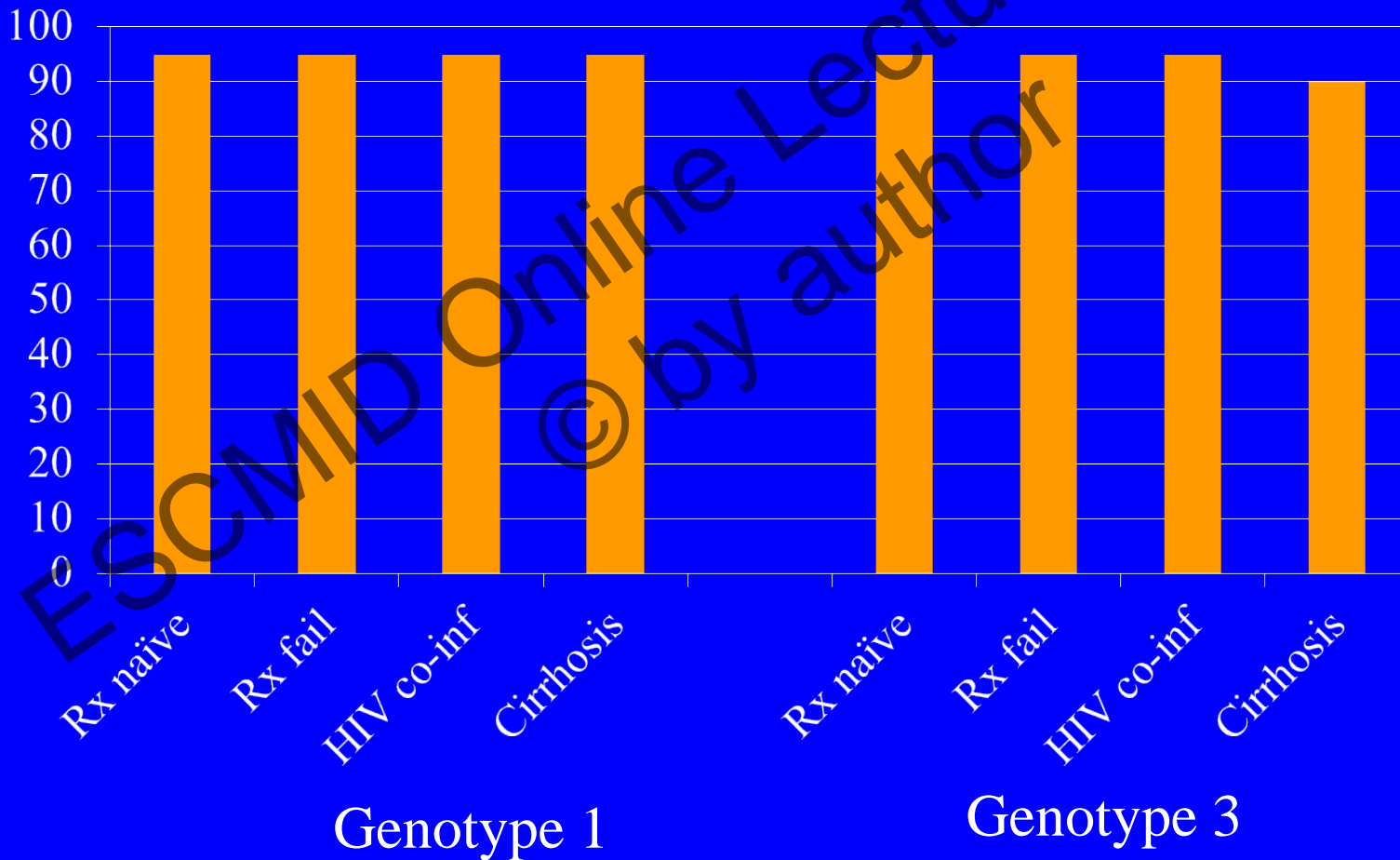


ION-1 Study Design: a randomised, open-label trial evaluating 12 and 24 weeks of treatment with HARVONI with or without RBV in treatment-naïve subjects with HCV GT1¹

*The on-treatment virological failure patient was confirmed as being non-compliant

Perfectovir

SVR



Monitoring therapy

- 1999 – IFN/RV, finish course, check for SVR
- 2005 – PEG/RV, apply stopping rule at week 12 for g1 patients
- 2008 – PEG/RV, assess for RGT (week 4), then apply stopping rules
- 2011 – PEG/RV/TPR/BOC – assess for RGT (weeks 4/8, 12/24) + stopping rules
- 2015 – DAAs, finish course, check for SVR