

P0454 Widespread copper, mercury and arsenic tolerance genes among multidrug-resistant *Enterococcus* spp. from human, animal and environmental origins

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Background: *Enterococcus* spp. are opportunistic pathogens able to survive under diverse environmental stressors. Tolerance to metals might participate in the selection of antibiotic-resistant (ABR) strains and/or genetic elements, but has been scarcely explored among *Enterococcus* spp. This study aimed to evaluate copper-CuT, mercury-HgT and arsenic-AsT tolerance in well-characterized multidrug-resistant-(MDR) *Enterococcus* spp. from different species and sources, and to assess their metal tolerance genetic context.

Materials/methods: We include 124 *E. faecium*-(Efm), 131 *E. faecalis*-(Efl) and 58 *Enterococcus* spp.-(Ep) from human-103, animal-production-setting/food-157 and environmental-53 origins (1996-2012; Portugal), being 72% (n=225/313) MDR. The occurrence of CuT (*tcrB/cueO*), HgT (6 *merA* variants) and AsT (5 *arsA* variants) genes was assessed by PCR/sequencing. Susceptibility to CuSO₄ and Na₂HAsO₄ was determined by the agar dilution method (n=195 isolates). Metal tolerance (*tcrB/cueO/merA/arsA*) and tetracycline-resistance [*tet(M)*-widespread in diverse niches, bacterial hosts and genetic platforms; tetracycline is widely used in different settings] genes location was assessed by hybridization (S1-PFGE) in 48 isolates (different sources and species).

Results: Genes coding for metal tolerance were detected in 36% (n=112/313) of isolates from different species (*Efm*-n=67/112-60%, *Efls*-n=30/112-27%, *Ep*-n=15/112-13%) and sources (human-n=32/112-29%; animal/food-n=68/112-61%; environment-n=12/112-11%). They comprised CuT (*tcrB*-n=74/112-66%; *cueO*-n=85/112-76%), HgT (*merA*_IIA-n=19/112-17%; *merA*_IIB-n=1/112-1%; *merA*_III-n=7/112-6%; *merA*_V-n=2/112-2%; *merA*_VI-n=6/112-5%) and AsT (*arsA*_AI-n=15/112-13%; *arsA*_All-n=14/112-13%; *arsA*_BII-n=2/112-2%). The most often combinations of genes coding for tolerance to different metals were *tcrB+cueO+merA*_IIA-n=18-Efm/1-Ep and *tcrB+cueO+arsA*_AI-n=6-Efm. Isolates with CuT/AsT genes were more tolerant to CuSO₄ (MIC₉₀=25mM vs MIC₉₀=12mM with no genes) or Na₂HAsO₄ (MIC₉₀=32mM vs MIC₉₀=8mM). Plasmids (90-300kb) often had co-located more than one metal tolerance gene: *tcrB+cueO* (n=21; Efm/Efl/Ep), *tcrB+cueO+merA*_IIA (n=9; Efm); *tcrB+cueO+merA*_VI (n=3; Efm), *tcrB+cueO+merA*_IIA+*merA*_VI (n=1; Efm), *tcrB+cueO+merA*_IIA+*merA*_VI+*arsA*_II (n=1; Efm), *cueO+merA*_VI (n=1; Efm); *tcrB+merA*_IIA (n=1; Efm), only one gene (n=3; Efm). Plasmid hybridization with *arsA*_A variants was not detected for 13 of

16 isolates tested (Efm/Efl/Ep), suggesting their frequent chromosomal location. The *tet(M)* hybridized in the same plasmids as *tcrB*+*cueO* with/without *merAII_A/arsA_II* (n=10 isolates).

Conclusions: MDR-*Enterococcus* spp. often carry genes encoding tolerance to metals highly used in antropogenic activities. Their often co-integration in plasmids with widespread ABR [*tet(M)*] genes suggest an adaptation of *Enterococcus* to frequent stresses, potentially associated with a continuous selection of MDR strains in different contexts.