

Diagnostic performance of a new multiplex PCR in patients with meningitis and/or encephalitis

G. Kapnisi¹, H. Sakkas¹, E. Priavali¹, A. Giannaki¹, A. Makis², S. Levidiotou¹, K. Gartzonika¹

¹ Department of Microbiology, ² Department of Pediatrics,
Faculty of Medicine, School of Health Sciences, University of Ioannina, Greece

Background

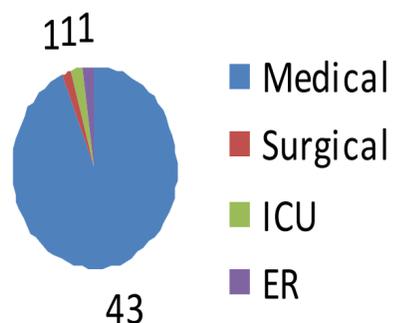
- Infectious meningitis and encephalitis, can lead to significant long-term morbidity, mortality and high health care costs.
- Rapid and accurate molecular diagnostic tests for the most common causes of these infections have the potential for high clinical impact.
- We present our experience using the FilmArray Meningitis/Encephalitis (ME) panel in comparison with standard diagnostic methods.

Material/methods I

Over an 8-month period (April-November 2016):

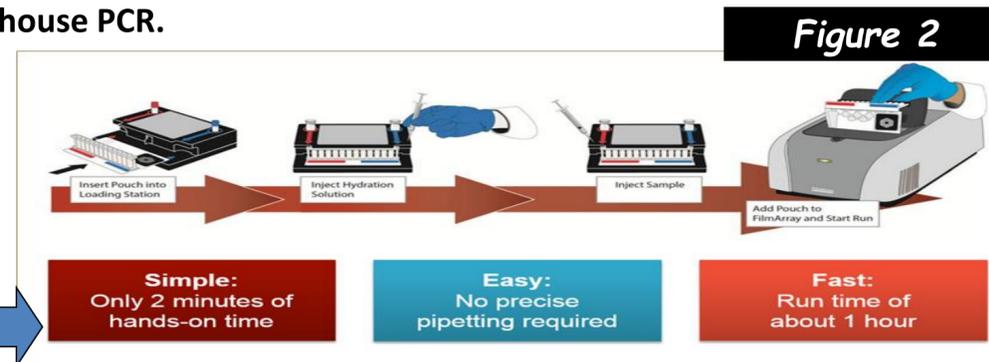
- Forty six (46) cerebrospinal fluid (CSF) specimens obtained from individuals suspected to have meningitis and/or encephalitis (**Figure 1**) were collected in the University Hospital of Ioannina
- The specimens were tested with the **FilmArray ME panel** (FilmArray, bioMerieux) (**Figure 2**) which is a qualitative nested-multiplexed nucleic acid-based diagnostic test for simultaneous detection and identification of the following 14 common agents capable of causing community-acquired ME.
 - ❖ **6 bacteria:** *E. coli* K1, *N. meningitis*, *H. Influenzae*, *S. pneumoniae*, *S. agalactiae*, *L. monocytogenes*
 - ❖ **7 viruses:** *CMV*, *Enterovirus*, *HSV1*, *HSV2*, *HHV-6*, *VZV*, *H. parechovirus*
 - ❖ **1 fungus:** *C. neoformans/gattii*

Figure 1 CSF specimen distribution by ward



Material/methods II

- Bacterial and fungal testing was also performed by **Gram stain** and **culture** on solid and broth media and isolates were identified using the automated **VITEK 2** system and **API** (bioMerieux, France).
- Testing for Enteroviruses and Herpesviruses group were in parallel carried out using the Enterovirus and Herpes **Consensus PCR** methodology respectively (Argene, BioMerieux, France). Nucleic acid extraction was performed using the QIAamp Viral RNA and DNA mini kit, respectively (Qiagen, Valencia).
- Samples with discrepant viral PCR results were retested by a **single house PCR**.



Results

- Pathogens were identified in 10 (21.7%) CSF samples by routine evaluation and in 11 (23.9%) by ME panel (Table 1).
- The sensitivity and specificity of FilmArray ME panel:
 - was 100% in detecting bacterial and fungal pathogens compared with conventional methods.
 - in viral detection, one discrepant result across FilmArray and Consensus PCR was observed. The FilmArray result was also confirmed by the in-house PCR (Table 1)
 - routine evaluation identified no pathogens in 35 FilmArray ME negative results.
- The time from receipt of CSF to report the organism identification was estimated for the FilmArray ME panel at 2.5 hours while for the standard methods at 16.5 hours.
- The distribution of microbial pathogens by age are shown in Table 2.

Table 1	Positive with standard diagnostic methods (Gram and/or culture, PCR)	Positive with FilmArray ME Panel	Positive with house PCR (discrepant results)
Pathogens detected			
<i>N.meningitis</i>	2 (1 only in Gram stain)	2	-
Enterovirus	2	2	-
VZV	5	6	1
<i>C.neoformans/gattii</i>	1	1	-

Table 2

Pathogens detected	<2 ms	2-23 ms	2-17 yrs	18-34 yrs	35-64 yrs	>64 yrs
<i>N.meningitis</i>				2		
Enterovirus	1		1			
VZV				1	2	3
<i>C.neoformans/gattii</i>					1	

Conclusions

- The FilmArray PCR panel offers a promising platform for the rapid diagnosis of ME and would be useful to overcome some of the challenges for conventional laboratory-based diagnosis of these infections.
- Further studies are needed to prospectively evaluate the clinical impact and cost-effectiveness of FilmArray ME panel.