

# The Bio-Rad $\beta$ CARBA Assay Detects a Diverse Range of Carbapenemase-Producing Organisms (CPO)

BM Willey, X Trimi, R Iaboni, S Rajadurai, DN Grohn, DA Boyd, L. Mataseje, G Ricci, A Mazzulli, D Terenzi, P Lo, M. Mulvey, T Mazzulli, SM Poutanen  
Mount Sinai Hospital/UHN, University of Toronto, Toronto, Ontario; William Osler Health Sciences Centre, Brampton, Ontario; and the National Microbiology Laboratory, Winnipeg, Manitoba; CANADA

## ABSTRACT

**Background:** Although PCR-based confirmatory tests are now available, they typically do not detect all possible carbapenemases and are generally too costly for use on every suspected CPO. Phenotypic tests that rely on visualization of pH changes resulting from imipenem hydrolysis have been suggested in standards but have proven unreliable for detecting class D CPO in many clinical microbiology laboratories. This study determined the performance of Bio-Rad's  $\beta$ CARBA assay, which detects CPO via a pH independent colour-change, as it is the chromogenic substrate itself (a proprietary carbapenem) that changes colour.

**Methods:** 259 species-diverse isolates, highly-characterized by PCR/sequencing, were blinded to prevent bias. They included 221 CPO (108 class A: 99 KPC, 4 SME, 2 IMI-1, 2 GES, 1 NMC-A; 80 class B: 73 NDM, 6 VIM, 1 IMP7; 26 class D; OXA48; OXA181, OXA232, OXA244; 7 class B+D: NDM+OXA181, NDM+OXA232) and 38 non-CPO (derepressed/plasmid-mediated *ampC*, ESBL, *ompC/ompF*-mutants, *ompK35/ompK36*-mutants, *cphA*, OXA252). On/after recovery from -80°C, ertapenem discs were added to all plates for selective pressure. For  $\beta$ CARBA testing, 259/259 isolates were plated to Oxoid Columbia Sheep Blood agar (CSBA) while 76/259 were plated to Mueller-Hinton agar (MHA), these comprised 58 CPO (11 KPC, 3 SME, 2 GES, 1 IMI1, 1 NMC-A; 12 NDM, 6 VIM, 1 IMP7; 17 OXA48-like, 4 NDM+OXA48-like) and 18 non-CPO. All 335  $\beta$ CARBA tests were inoculated from growth surrounding ertapenem discs. Incubation, as per Bio-Rad, was at 37°C. Each tube was read independently at 30min by 5 readers for colour-changes from yellow (negative) to orange, red or purple (positives). Consensus reads were analyzed for sensitivity/specificity, and 95% confidence intervals were calculated in [www.graphpad.com](http://www.graphpad.com).

**Results:** Overall,  $\beta$ CARBA was 98.2% sensitive (95%CI: 95.8-99.3) and 100% specific (95%CI: 92.2-100) detecting 274/279 CPO in 335 tests from both agars combined.  $\beta$ CARBA performed equally well from CSBA and MHA detecting 218/221 (98.6%) and 56/58 (96.6%) of the CPO, respectively. Detection notably included 100% of the troublesome OXA48-like CPO (CSBA: 33/33; MHA: 21/21). False-negatives from both agars included the unique *nmcA*-positive *Enterobacter cloacae* and 1/2 GES *Klebsiella oxytoca* -  $\beta$ CARBA tests remained yellow >45min and again on repeat. The only  $\beta$ CARBA false-positive result was from an *Aeromonas hydrophila* (intrinsic carbapenem-resistance due to *cphA*). This species would be excluded from testing based on its identity, and thus was not included in specificity calculations.

**Conclusions:**  $\beta$ CARBA is low-complexity assay that is quick and easy to set-up and simple to interpret. When faced with a possible CPO,  $\beta$ CARBA provides a highly sensitive/specific results in 30min from 1 $\mu$ L-loop of organism, unlike similar tests that require larger inoculum and lengthier incubations.

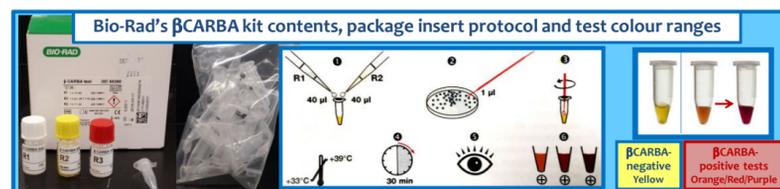
## INTRODUCTION

Rapid and accurate detection of CPO is critical to patient care and to limit dissemination. CPO are multidrug resistant Gram-negative bacilli (MDR-GNB) that present an urgent public health threat as they are typically resistant to all available antimicrobials including carbapenems which are generally considered among the last resort agents for treating serious infections.

Molecular methods are considered "gold standard" for confirming carbapenemase genes, yet no currently available PCR assay detects all CPO. This is because novel genotypes frequently arise without apparent warning, and known genotypes have been noted to diversify via genetic drift at rates which have reduced molecular test sensitivities. Availability of a routine PCR test for the limited number of common genotypes is important in certain situations to enable rapid detection (<4h) of CPO directly from specimens. But for day to day use, availability of a reliable rapid test capable of detecting phenotypic evidence of any carbapenemase in a suspicious surveillance or clinical isolate will reduce the overall risk of missing new or unusual genotypes, and enable improved patient management by allowing important infection control decisions to be made without delay. This was the idea behind the CARBA-NP assay, but in most labs, this pH indicator-based phenotypic test proved unreliable for detecting Class D type CPO.

Generally, specimens received for CPO surveillance are first screened using selective culture while CPO in clinical specimens are first detected via routine susceptibility testing. Suspected CPO are then ruled out depending on the laboratory by any of a number of methods including inhibitor testing, limited range PCR, or referral to reference laboratories for more extensive PCR. This may result in long reporting delays during which time dissemination of an unrecognized CPO may take place.

This study determined the utility of a low-complexity phenotypic CPO detection test, Bio-Rad's  $\beta$ CARBA assay. In contrast to the CARBA-NP, the  $\beta$ CARBA assay detects hydrolysis of a proprietary colorimetric carbapenem which undergoes a colour-change within 30min exposure to a CPO. This study was designed to assess the practicality, sensitivity, specificity, and cost-effectiveness of implementing the  $\beta$ CARBA into routine workflow of a large clinical microbiology laboratory to detect CPO from isolates grown on a variety of agars.



## METHODS

Table 1 describes the isolate characteristics and distribution by agar type used in the study. On and after recovery from -80°C, ertapenem discs were added to all subculture plates to maintain selective pressure. In preparation for blinded testing, all 259 isolates were plated to Oxoid's Columbia-based 5% Sheep Blood agar (Blood agar), of which 76 were also plated to Oxoid's Mueller-Hinton Plus agar (MHA). plates as before had an ertapenem disc placed on the inoculated area to maintain antibiotic pressure. Using fresh cultures of the total 335 isolates, Bio-Rad  $\beta$ CARBA tests were performed as per package inserts. Briefly, all liquid from Reagent 1 was transferred to dissolve the dehydrated contents of Reagent 2. Then 40 $\mu$ L ea. Regents 2 and 3 were pipetted into a micro-tube (provided in kit). A heavy 1 mL loop of colonies derived from growth closest to ertapenem discs on each plate was inoculated into each tube and mixed thoroughly. At 30 min incubation at 37°C (water-bath), tubes were read independently by 5 readers for colour-changes: Yellow = CPO-negative; Orange, Red or Purple = CPO-positive. Consensus reads were analyzed for sensitivity and specificity, 95% confidence intervals were calculated in [www.graphpad.com](http://www.graphpad.com).

**Table 1. Characteristics of 259 bacteria and agars used to evaluate Bio-Rad's  $\beta$ CARBA assay**

Ambler (No.)	CPO Genotypes (No.)	Species Identification	Blood agar (No.)	MHA subset (No.)
Class A CPO (108)	<i>blaKPC</i> (99)	<i>Citrobacter freundii</i>	2	0
		<i>Enterobacter aerogenes</i>	3	1
		<i>Enterobacter cloacae</i>	35	2
		<i>Escherichia coli</i>	12	3
		<i>Klebsiella oxytoca</i>	1	1
		<i>Klebsiella pneumoniae</i>	46	4
	<i>blaGES5</i> (2)	<i>Klebsiella oxytoca</i>	2	2
	<i>blaIM1</i> (1)	<i>Enterobacter cloacae</i>	1	1
	<i>blaNMC</i> (2)	<i>Enterobacter cloacae</i>	2	1
	<i>blaSME</i> (4)	<i>Serratia marcescens</i>	4	3
Class B CPO (80)	<i>blaIMP7</i> (1)	<i>Pseudomonas aeruginosa</i>	1	1
		<i>Acinetobacter baumannii</i>	1	1
	<i>blaNDM</i> (73)	<i>Citrobacter freundii</i>	1	1
		<i>Enterobacter cloacae</i>	3	1
		<i>Escherichia coli</i>	30	2
		<i>Klebsiella pneumoniae</i>	33	3
		<i>Morganella morganii</i>	4	3
		<i>Proteus mirabilis</i>	1	1
	<i>blaVIM</i> (6)	<i>Citrobacter freundii</i>	1	1
	<i>Enterobacter cloacae</i>	4	4	
<i>Pseudomonas putida</i>	1	1		
Classes B + D CPO (7)	<i>blaNDM+blaOXA181</i> (1)	<i>Escherichia coli</i>	1	1
		<i>Klebsiella pneumoniae</i>	3	2
	<i>blaNDM+blaOXA232</i> (1)	<i>Escherichia coli</i>	1	0
		<i>Klebsiella pneumoniae</i>	2	1
Class D CPO (26)	<i>blaOXA48</i> (15)	<i>Escherichia coli</i>	8	6
		<i>Klebsiella pneumoniae</i>	7	4
	<i>blaOXA181</i> (6)	<i>Escherichia coli</i>	2	0
		<i>Klebsiella pneumoniae</i>	4	3
	<i>blaOXA232</i> (4)	<i>Escherichia coli</i>	1	0
		<i>Klebsiella pneumoniae</i>	3	2
Non-CPO (38)	<i>ompC-ompF</i> (3)	<i>Enterobacter cloacae</i>	1	1
		<i>Escherichia coli</i>	3	3
	<i>ompK35-ompK36</i> (6)	<i>Klebsiella pneumoniae</i>	6	6
		<i>Klebsiella oxytoca</i>	1	1
	<i>Other mechanisms</i> (1)	<i>Enterobacteriaceae</i>	26	7
<i>blaOXA252</i> (1)	<i>Shewanella putrefaciens</i>	1	0	

## RESULTS

**Table 2: Summary of CPO detection results obtained using Bio-Rad's  $\beta$ CARBA assay when challenged by 259 distinct isolates cultivated on two common laboratory agars**

CPO Genotypes Tested	No. (%) Blood agar/ No. (%) $\beta$ CARBA-Pos	No. (%) Mueller-Hinton agar/ No. (%) $\beta$ CARBA-Pos	Tn. (%) Tested/ Tn. (%) $\beta$ CARBA-Pos
<i>blaKPC</i>	99 (38.2)/99 (100)	11 (14.5)/11 (100)	110 (32.8)/110 (100)
<i>blaNDM</i>	73 (28.2)/73 (100)	12 (15.8)/12 (100)	85 (25.4)/85 (100)
<i>blaOXA48-like</i>	26 (10)/26 (100)	17 (22.4)/17 (100)	43 (12.8)/43 (100)
<i>blaNDM+OXA48-like</i>	7 (2.7)/7 (100)	4 (5.3)/4 (100)	11 (3.3)/11 (100)
<i>blaVIM</i>	6 (2.3)/6 (100)	6 (7.9)/6 (100)	12 (3.6)/12 (100)
<i>blaSME</i>	4 (1.5)/4 (100; 1 weak)	3 (4)/3 (100)	7 (2.1)/7 (100)
<i>blaGES5</i>	2 (0.8)/0 (0)	2 (2.6)/1 (50)	4 (1.2)/1 (25)
<i>blaNMC<sub>A</sub></i>	2 (0.8)/1 (50)	1 (1.3)/0 (0)	3 (1.5)/1 (33)
<i>blaIM1</i>	1 (0.4)/1 (100)	1 (1.3)/1 (100)	2 (0.6)/2 (100)
<i>blaIMP7</i>	1 (0.4)/1 (100)	1 (1.3)/1 (100)	2 (0.6)/2 (100)
Non-CPO	37 (14.3)/0 (0)	18 (23.7)/0 (0)	55 (16.4)/0 (0)
Non-CPO (intrinsic <i>cphA</i> )	1 (0.4)/1 (100)	ND	1 (0.3)/1 (100)
Total CPO tested	221 (85.3)/218 (98.6)	58 (76.3)/56 (96.6)	279 (83.3)/274 (98.2)
Total non-CPO tested	38 (14.7)/1 (2.6)	18 (23.7)/0 (0)	56 (16.7)/1 (1.8)
Total isolates tested	259	76	335

## RESULTS

Table 2 (bottom left) summarizes  $\beta$ CARBA results obtained from blood and Mueller-Hinton agars, while Table 3 (below) displays overall  $\beta$ CARBA sensitivity and specificity with (95%CI) for detecting 274/279 98.2% CPO in 335 tests from both agars combined.

$\beta$ CARBA performed well from Blood/MHA, from which CPO detection was 218/221 (98.6%)/56/58 (96.6%) overall, and for OXA48-types, it was 33/33 (100%)/21/21 (100%), respectively.

False-negatives from both agars were rarely encountered in unusual class A genotypes only, namely in *K. oxytoca* with *blaGES5* and *E. cloacae* with *blaNMC<sub>A</sub>* which were not or only poorly detected by  $\beta$ CARBA from either agar (Table 2). In these isolates, the  $\beta$ CARBA tests reproducibly were yellow beyond 45 min. Sequencing confirmed their alleles to be *blaGES5* - a genotype with recognized carbapenemase activity. Similarly, sequences were determined for the 3 *E. cloacae* with closely related *blaNMC/IMI* genes. Although only represented by a few isolates in this study, it appeared that *blaIMI* readily hydrolyzed the proprietary carbapenem utilized in the  $\beta$ CARBA assay while *blaNMC<sub>A</sub>* encountered difficulties. In all cases, the  $\beta$ -lactamases produced by these isolates failed to hydrolyze the particular carbapenem used in the assay regardless of evidence of growth to ertapenem on both test agars, and phenotypic meropenem resistance as demonstrated by MHA using the screen breakpoint of  $\leq$ 25mm to indicate a suspect CPO.

The only  $\beta$ CARBA "false-positive" result was from an *Aeromonas hydrophila*. Since this species is intrinsically carbapenem-resistant due to chromosomal *cphA*, it is not considered a true CPO in the epidemiological sense. If encountered, this species should be excluded from further CPO testing based on its identity. For this reason, it was excluded from  $\beta$ CARBA specificity calculations. But as it mounted a reliably strong  $\beta$ CARBA-positive reaction, this species could be retained as a safe quality control isolate for this and equivalent tests, so to be readily discernable from potentially cross-contaminating CPO in clinical laboratories.

**Table 3: Overall Bio-Rad  $\beta$ CARBA test CPO detection performance from two common agars**

$\beta$ CARBA Sensitivity for any CPO	98.2% (95% CI: 95.8-99.3)
$\beta$ CARBA Specificity (excluding intrinsic mechanisms)	100% (95% CI: 92.2-100)

## DISCUSSION & CONCLUSION

### Bio-Rad's $\beta$ CARBA phenotypic CPO detection assay

- ✓ Simple to use and very easy to interpret (<1 minute hands-on time)
- ✓ Clear results available within 30 minutes
- ✓ Only 1 $\mu$ L loop of bacterial colonies required to set up test
- ✓ Accurate results from most common agars - Mueller-Hinton agar, Columbia-based sheep blood agar (insert precludes use of  $\beta$ CARBA from colonies on MacConkey-based agars citing unreliable results)
- ✓ Highly specific (100%) - no false-positive at 30min with the exception of genera with intrinsic resistance (exclude these based on identity)
- ✓ Highly sensitive (98.2%) for overall CPO detection, with 100% detection in the most common CPO such as *blaKPC*, *blaNDM*, *blaVIM*, and very notably in the troublesome *blaOXA48*-like genotypes
- ❖ Unreliable for detecting the rare *blaNMC-A* (1/9) and *blaGES5* (0/2) CPO

In conclusion,  $\beta$ CARBA is a cost-effective, low-complexity assay as it is relatively inexpensive, quick and easy to use, extremely simple to interpret, thus improves laboratory workflow. When faced with a possible CPO,  $\beta$ CARBA provides highly sensitive/specific results in 30min from only 1 $\mu$ L loop of organism, unlike similar phenotypic tests that require more colonies and lengthier incubations.

## Acknowledgements

$\beta$ CARBA test kits sufficient to complete this study were kindly provided by Bio-Rad