

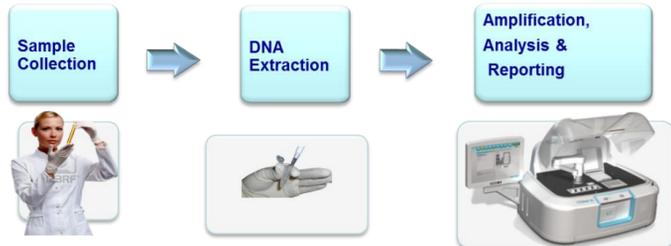
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Background

Infectious Gastroenteritis is a global health problem associated with extremely high morbidity and mortality rates. Accurate diagnosis is crucial to allow appropriate and timely treatment. Stool testing at the microbiology laboratory is currently a complex, time consuming and cumbersome process, demanding highly qualified personnel and application of a wide range of techniques. Thus, workload, lab space and turnaround time are high and costly. Savyon Diagnostics has recently finalized the development of a novel molecular-based diagnostic screening test for simultaneous detection of nine bacterial and protozoan parasitic pathogens tailored specifically according to the demands of a typical laboratory at community setting. The test was developed on Savyon proprietary NanoCHIP® molecular electronic microarray system. The bacterial panel includes *Salmonella*, *Shigella* and *Campylobacter* spp. The parasitic panel is composed of *Entamoeba histolytica*, *Entamoeba dispar*, *Giardia lamblia*, *Dientamoeba fragilis*, *Cryptosporidium* spp. and *Blastocystis* spp. The simultaneous detection of all these pathogens in both panels is enabled due to the high multiplex capabilities that characterize the NanoCHIP® platform, that together with testing multiple patient samples in the same run offers a powerful and unique medium-high throughput screening tool. The system demonstrates high performance, versatility in the panel composition according to lab specific needs, minimal hands-on time, vast reduction in the workload of the lab (detection of parasites and bacteria in the same runtime) and cost-effectiveness. The system is compatible with a variety of automatic DNA extraction systems and thus is capable to fully integrate in the routine workflow of clinical laboratories of various sizes.

The NanoCHIP® Workflow



Materials & Methods

- A clinical study composed of prospective and retrospective samples was conducted at Meuhedet Health Services, Rosh Haayin Laboratory, Israel, during February – April 2014 in symptomatic patients:
 - Retrospective samples: 46 positives
 - Prospective samples: 150 (31 positives; 119 negatives)
- The DNA was extracted by the Bullet PRO® system (Diasorin), amplified by PCR and processed by the NanoCHIP® NC400 instrument (Savyon)
- Results were compared to standard methods used routinely by the lab, i.e. culture and microscopy
- Discrepant analysis was carried out by Diagenode® RT-PCR assays (Mikrogen) and/or by sequencing

The performance parameters following the prospective study demonstrate the utility of The NanoCHIP GIP® test to be used routinely for screening purposes in the microbiology lab in the community

Objective

The aim of this work was to demonstrate the utility of the NanoCHIP® GIP test for screening of gastro-enteric bacterial and parasitic pathogens in stool specimens in a typical laboratory at community setting

The NanoCHIP® GIP Combi II Panel

| Parasites | LOD (CFU/mL) | Bacteria | LOD (CFU/mL) |
|--------------------------|--------------|----------------------|--------------|
| <i>Giardia lamblia</i> | 5.00E+03 | <i>Salmonella</i> | 1.00E+05 |
| <i>Cryptosporidium</i> | ND | <i>Shigella</i> | 1.00E+04 |
| <i>E. histolytica</i> | 5.00E+03 | <i>Campylobacter</i> | 1.40E+04 |
| <i>E. dispar</i> | 5.00E+03 | | |
| <i>D. fragilis</i> | 5.00E+03 | | |
| <i>Blastocystis</i> spp. | ND | | |

Results

Table 1. Results before and after analysis of discrepancies

| Pathogen | Results vs. Conventional Methods | | | | | | Discrepant Analysis Results | | | | | |
|------------------------|----------------------------------|----|-----|----|-------------|-------------|-----------------------------|----|-----|----|-------------|-------------|
| | TP | FP | TN | FN | Sensitivity | Specificity | TP | FP | TN | FN | Sensitivity | Specificity |
| <i>Giardia</i> | 34 | 2 | 157 | 3 | 92 | 99 | 35 | 1 | 159 | 1 | 97 | 99 |
| <i>Campylobacter</i> | 9 | 8 | 178 | 1 | 90 | 96 | 17 | 0 | 179 | 0 | 100 | 100 |
| <i>Shigella</i> | 15 | 16 | 164 | 1 | 94 | 91 | 31 | 0 | 165 | 0 | 100 | 100 |
| <i>Salmonella</i> | 3 | 1 | 190 | 2 | 60 | 99 | 4 | 0 | 192 | 0 | 100 | 100 |
| <i>Cryptosporidium</i> | 1 | 1 | 194 | 0 | 100 | 99 | 2 | 0 | 194 | 0 | 100 | 100 |
| <i>E. histolytica</i> | 3* | 0 | 193 | 0 | N/A | 100 | 3 | 0 | 193 | 0 | 100 | 100 |
| <i>D. fragilis</i> | 2 | 33 | 161 | 0 | 100 | 83 | 35 | 0 | 161 | 0 | 100 | 100 |
| <i>B. hominis</i> | 3 | 25 | 168 | 0 | 100 | 87 | 28 | 0 | 168 | 0 | 100 | 100 |

* Defined by the lab as *Entamoeba coli*

According to the discrepancy analysis, the NanoCHIP® GIP detected 87 positive samples that were defined originally as negatives by the conventional methods

7 samples that were detected as negatives by the NanoCHIP® GIP Combi II were confirmed as false positive detection by the conventional methods

Table 2. Prospective study results

| Pathogens | TP | FN | TN | FP | Sens | Spec | PPV | NPV |
|--------------------------|----|----|-----|----|------|------|------|------|
| <i>B. hominis</i> | 21 | 0 | 129 | 0 | 100% | 100% | 100% | 100% |
| <i>D. fragilis</i> | 27 | 0 | 123 | 0 | 100% | 100% | 100% | 100% |
| <i>Salmonella</i> * | 1 | 0 | 149 | 0 | 100% | 100% | 100% | 100% |
| <i>Shigella</i> | 19 | 0 | 131 | 0 | 100% | 100% | 100% | 100% |
| <i>Campylobacter</i> | 15 | 0 | 135 | 0 | 100% | 100% | 100% | 100% |
| <i>Giardia</i> * | 5 | 0 | 145 | 1 | 100% | 99% | 83% | 100% |
| <i>Cryptosporidium</i> * | 2 | 0 | 148 | 0 | 100% | 100% | 100% | 100% |
| <i>E. histolytica</i> * | 2 | 0 | 148 | 0 | 100% | 100% | 100% | 100% |
| <i>E. dispar</i> | 0 | 0 | 150 | 0 | N/A | 100% | N/A | 100% |

| Total GIP II assay | Prospective: total assay | | | | | | | |
|--------------------|--------------------------|----|----|----|------|------|-----|------|
| | TP | FN | TN | FP | Sens | Spec | PPV | NPV |
| | 92 | 0 | 57 | 1 | 100% | 98% | 99% | 100% |

* The amount of positives is not statistically significant

Results (cont.)

Table 3. Overall** Study Results

| Pathogens | Overall assay | | | | | | | |
|--------------------------|---------------|----|-----|----|------|------|------|------|
| | TP | FN | TN | FP | Sens | Spec | PPV | NPV |
| <i>B. hominis</i> | 28 | 0 | 168 | 0 | 100% | 100% | 100% | 100% |
| <i>D. fragilis</i> | 35 | 0 | 161 | 0 | 100% | 100% | 100% | 100% |
| <i>Salmonella</i> * | 4 | 0 | 192 | 0 | 100% | 100% | 100% | 100% |
| <i>Shigella</i> | 31 | 0 | 165 | 0 | 100% | 100% | 100% | 100% |
| <i>Campy</i> | 17 | 0 | 179 | 0 | 100% | 100% | 100% | 100% |
| <i>Giardia</i> | 35 | 1 | 159 | 1 | 97% | 99% | 97% | 99% |
| <i>Cryptosporidium</i> * | 2 | 0 | 194 | 0 | 100% | 100% | 100% | 100% |
| <i>E. histolytica</i> * | 3 | 0 | 193 | 0 | 100% | 100% | 100% | 100% |
| <i>E. dispar</i> | 0 | 0 | 150 | 0 | N/A | 100% | N/A | 100% |

| Total GIP II assay** | Overall assay | | | | | | | |
|----------------------|---------------|----|-----|----|------|------|-----|-----|
| | TP | FN | TN | FP | Sens | Spec | PPV | NPV |
| | 76 | 1 | 118 | 1 | 99% | 99% | 99% | 99% |

*The amount of positives is not statistically significant

** Overall results in retrospective and prospective samples
The rate of mixed infections: 36 out of 196 (18%) samples

Discussion & Summary

- The NanoCHIP® GIP Combi II has a higher detection yield compared to the conventional methods and the overall performance is better
- Improved detection is observed in all the pathogens composing the panel
- Proven differentiation between *histolytica* and *dispar* *Entamoeba* sub-types
- Reliable differentiation between *Blastocystis* / *D. fragilis* / *E. histolytica* and a higher positive rate in regard to these pathogens
- A procedure that is compatible with the laboratory needs in terms of time-to-result
- Efficient detection of mixed infections in one assay
- User-friendly, objective and clear interpretation of results without need for special skills
- Reduction in lab workload by avoiding separate procedures for bacterial and parasitic detection
- Overall, the NanoCHIP® GIP Combi II has demonstrated its utility in the community setting laboratory for reliable detection of bacterial and parasitic gastrointestinal infections and screening purposes
- The test presents significant advantages compared to currently used methods, mainly in terms of performance, minimal hands-on time, improved laboratory workflow, and potential assimilation as part of a fully automated process