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MALDI-TOF mass spectrometry for the detection of antimicrobial resistance mechanisms

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31 March – 3 April 2012



Presentation Content

- Mechanisms of antibiotic resistance; possible applications of MALDI-TOF MS
- Detection of β -lactamases
- Detection of ribosomal RNA methyltransferase activity
- Resistance determination by the detection of specific peaks
- Proteomic approach
- Minisequencing
- Future perspectives



Mechanisms Of Antibiotic Resistance

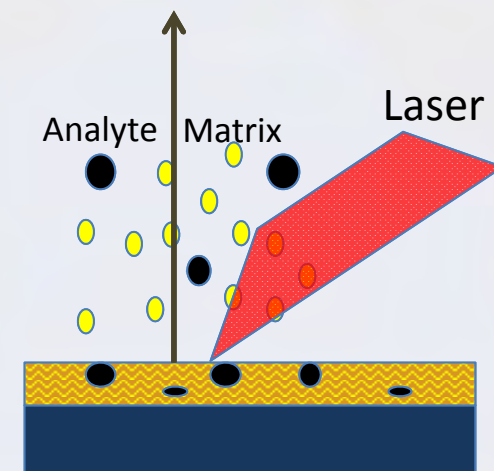
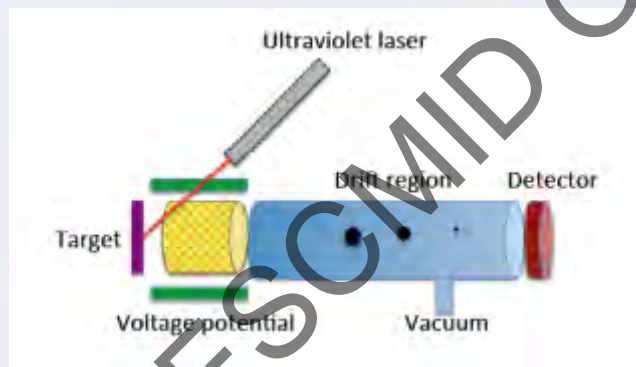
Possible Applications For MALDI-TOF MS

- Production of **enzymes** that inactivate antibiotic molecules (e.g., β -lactamases and aminoglycoside-modifying enzymes);
- Hyperproduction or production of novel **efflux pumps** and other changes in the **cell wall** (e.g., porin alteration);
- **Mutations** in target genes (e.g., in ribosomal protein genes or in genes coding for penicillin-binding proteins – PBPs);
- **Bypass** of a metabolic pathway (e.g., the expression of acquired PBPs with a low affinity for antibiotic molecules);
- Production of proteins that **preserve** the target site (e.g., quinolone resistance mediated by Qnr) or of **target site-modifying enzymes**



Already Published Applications For MALDI-TOF MS

- Analysis of **antibiotic molecules** and their **modified products**
- Analysis of **bacterial cell components**
- Analysis of **ribosomal RNA methylation**
- Detection of **mutations with mini-sequencing**





Detection Of β -Lactamases

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MALDI-TOF mass spectrometry for the detection of antimicrobial resistance mechanisms



Detection Of β -Lactamases

- MALDI-TOF MS is able to detect small molecules (e.g., antibiotics, hormones)
- **Could we detect β -lactams and their degradation products ?**

JOURNAL OF CLINICAL MICROBIOLOGY, Sept. 2011, p. 3222-3227
0095-1137/11/\$12.00 doi:10.1128/JCM.00984-11
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Vol. 49, No. 9

Carbapenemase Activity Detection by Matrix-Assisted Laser Desorption Ionization–Time of Flight Mass Spectrometry^v

Jaroslav Hrabák,^a Radka Walková, Vendula Študentová, Eva Chudíčková, and Tamara Bergerová

Department of Microbiology, Faculty of Medicine and University Hospital in Pilsen, Charles University in Prague, Pilsen, Czech Republic

JOURNAL OF CLINICAL MICROBIOLOGY, Sept. 2011, p. 3228-3234
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Vol. 49, No. 9

NOTES

Using Matrix-Assisted Laser Desorption Ionization–Time of Flight Mass Spectrometry To Detect Carbapenem Resistance within 1 to 2.5 Hours^v

Irene Burckhardt^a and Stefan Zimmermann

Department for Infectious Diseases, Microbiology and Hygiene, University of Heidelberg, Im Neuenheimer Feld 324, D-69115 Heidelberg, Germany

Matrix-Assisted Laser Desorption Ionization–Time of Flight Mass Spectrometry-Based Functional Assay for Rapid Detection of Resistance against β -Lactam Antibiotics

Katrin Sparbier,^a Sören Schubert,^b Ulrich Weller,^c Christiane Boogen,^c and Markus Kostrzewa^a

Bruker Daltonik GmbH, Bremen, Germany^a; Max von Laue Synchrotron-Institute, Munich, Germany^b; and Praxis für Laboratoriumsmedizin, Ärztliche Gemeinschaft für Diagnostik Köln-Born, Cologne, Germany^c

0095-1137/12/\$12.00 J. Clin. Microbiol. p. 927-937

jcm.asm.org 927



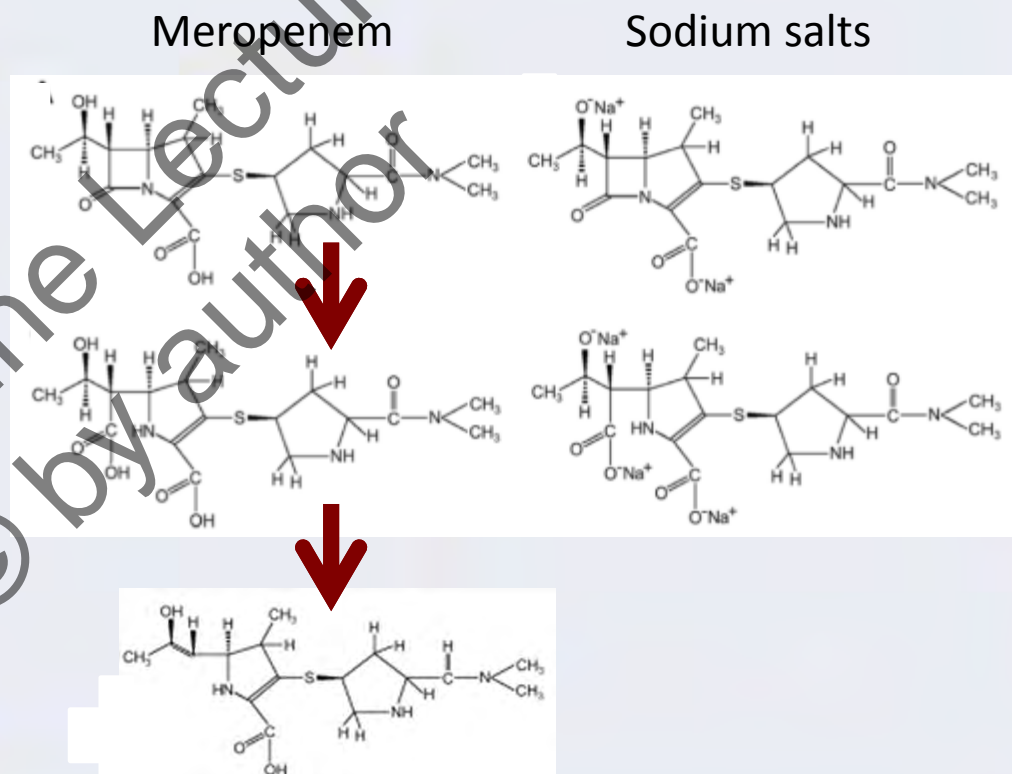
Carbapenemase Detection By MALDI-TOF MS

Principle Of the Meropenem Degradation

1. Hydrolysis of amide bound

2. Decarboxylation

Molecule	m/z
Meropenem MW	383,5
[Meropenem+H] ⁺	384,5
[Meropenem+Na] ⁺	406,5
[Meropenem+2Na] ⁺	428,5
[Meropenem _{hydr} +H] ⁺	402,5
[Meropenem _{hydr} +Na] ⁺	424,5
[Meropenem _{hydr} +2Na] ⁺	446,5
[Meropenem _{hydr} +3Na] ⁺	468,5
[Meropenem _{decarbox} +H] ⁺	358,5
[Meropenem _{decarbox} +Na] ⁺	380,5
[Meropenem _{decarbox} +2Na] ⁺	402,5

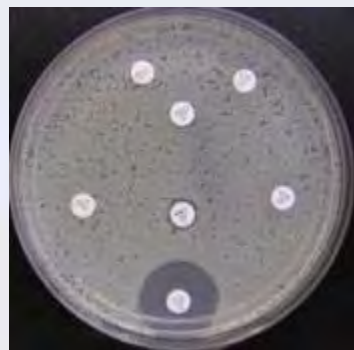


Walková *et al.* Carbapenemase identification by matrix assisted laser desorption/ionisation time-of-flight mass spectrometry P1714, 22nd ECCMID, London
 Hrabák *et al.*, MS in review



Carbapenemase Detection By MALDI-TOF MS

In *Enterobacteriaceae*, *Pseudomonas* spp., *Acinetobacter* spp.



3 McFarland

20 mM Tris-HCl
20 mM NaCl
pH 7.0



Centrifugation
of 1 ml of the
suspension



Resuspension
of pellet (in 50µl)

20 mM Tris-HCl
0.01% SDS
0.1 mM meropenem
pH 7.0

Incubation at
35°C for 2
hours

Centrifugation
of the mixture

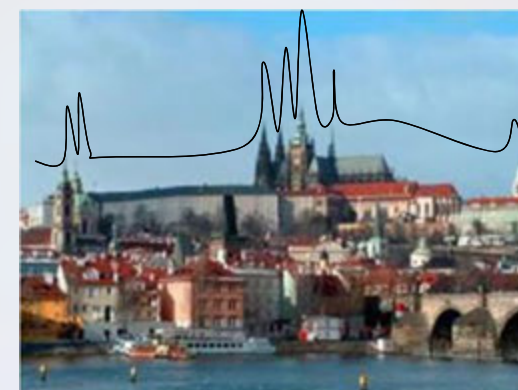
MALDI-TOF MS
(matrix: DHB
in 50% ethanol)

Walková *et al.* Carbapenemase identification by matrix assisted laser desorption/ionisation time-of-flight mass spectrometry P1714, 22nd ECCMID, London

Hrabák *et al.*, MS in review



Carbapenemase Detection By MALDI-TOF MS Spectra Analysis





MALDI-TOF mass spectrometry for the detection of antimicrobial resistance mechanisms

Carbapenemase Detection By MALDI-TOF MS

OPEN ACCESS Freely available online PLOS one

Rapid Detection of Carbapenem Resistance in *Acinetobacter baumannii* Using Matrix-Assisted Laser Desorption Ionization-Time of Flight Mass Spectrometry

Marie Kempf^{1*}, Sofiane Bakour^{1,2*}, Christophe Flaudrops¹, Meryem Berrazeg¹, Jean-Michel Brunel¹, Mourad Drissi³, Esma Mesli³, Abdelaziz Touati², Jean-Marc Rolain^{1*}

*Correspondence: Marie Kempf (marie.kempf@univ-poitiers.fr), Sofiane Bakour (sofiane.bakour@univ-poitiers.fr), Jean-Marc Rolain (jean-marc.rolain@univ-poitiers.fr)

1. UMR 1099 IGE, Université de Poitiers, France; 2. UMR 1099 IGE, Université de Poitiers, France; 3. UMR 1099 IGE, Université de Poitiers, France

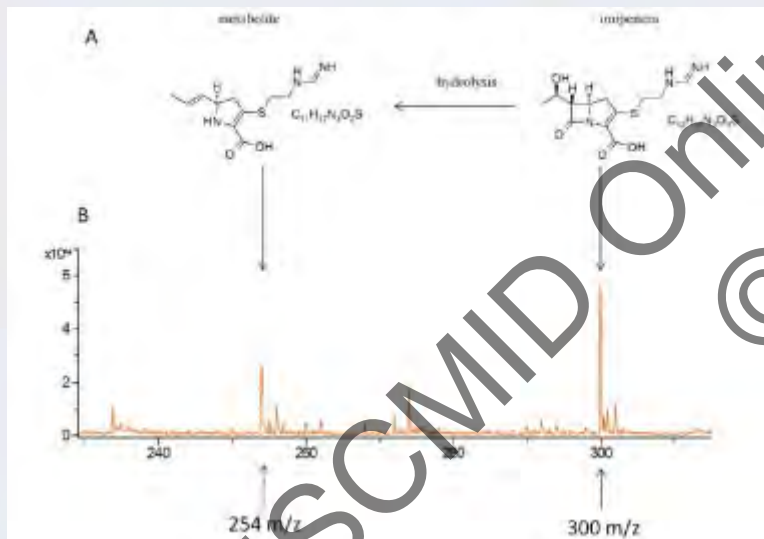


Table 1. Isolates used in the study.

Carbapenemase	Species (number of isolates)		
NDM-1	<i>Escherichia coli</i> (30)		
	<i>Enterobacter cloacae</i> (22)		
	<i>Citrobacter</i> spp. (6)		
	<i>Klebsiella pneumoniae</i> (5)		
	<i>Providencia rettgeri</i> (2)		
	<i>Acinetobacter baumannii</i> (2)		
	KPC-2, KPC-3	<i>Klebsiella pneumoniae</i> (10)	
		VIM-1	<i>Klebsiella pneumoniae</i> (16)
			<i>Enterobacter cloacae</i> (4)
	OXA-48	<i>Serratia marcescens</i> (3)	
<i>Escherichia coli</i> (3)			
<i>Enterobacter cloacae</i> (1)			
OXA-162	<i>Klebsiella pneumoniae</i> (2)		
	<i>Escherichia coli</i> (2)		
	<i>Raoultella ornithinolytica</i> (1)		
	<i>Citrobacter freundii</i> (1)		
Total number of carbapenemase-producing isolates:	110		
Non-carbapenemase-producing isolates resistant to carbapenems:			
	<i>Klebsiella pneumoniae</i> (28)		
	<i>Enterobacter cloacae</i> (4)		
	<i>Citrobacter</i> spp. (3)		
Total number of non-carbapenemase-producing isolates:	35		

Walková *et al.* Carbapenemase identification by matrix assisted laser desorption/ionisation time-of-flight mass spectrometry P1714, 22nd ECCMID, London
 Hrabák *et al.*, MS in review



β -Lactamase Detection By MALDI-TOF MS

- Cheap and quick method (turnaround time ca. 3 h!)
- **Detection of a real carbapenemase activity**
- Comparable with spectrophotometric assay
- Possible application to detect carbapenemases in *Acinetobacter baumannii*
- Detection of degradation products depends on a sample preparation (e.g., buffer system)



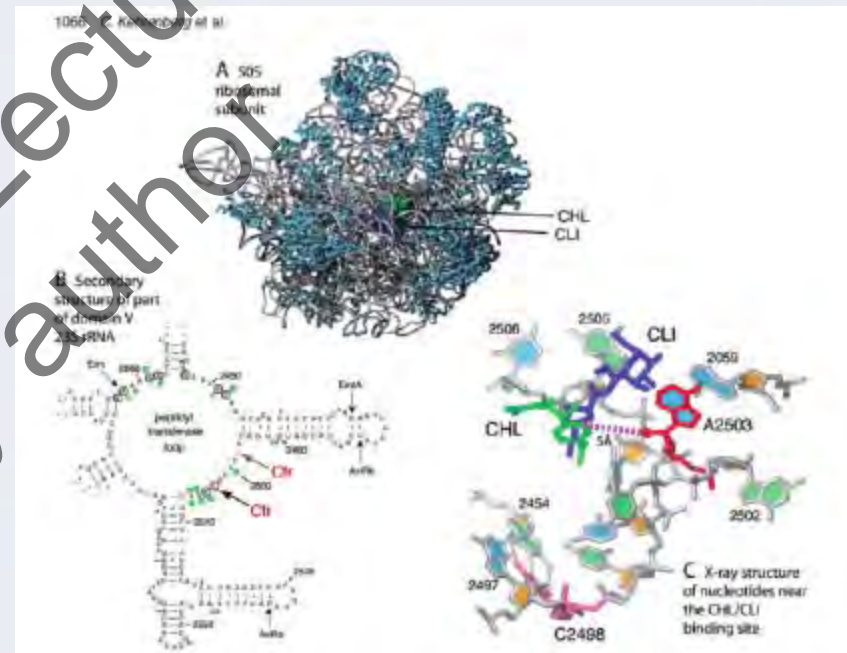
Detection Of Ribosomal RNA Methyltransferase Activity

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Detection Of Methyltransferase Activity

- Methylation of the RNA ribosomal subunit
- Resistance to:
 - Aminoglycosides
 - Chloramphenicol
 - Clindamycin



5420-5431 Nucleic Acids Research, 2009, Vol. 37, No. 16
doi:10.1093/nar/gkp375 Published online 9 July 2009

Determination of the target nucleosides for members of two families of 16S rRNA methyltransferases that confer resistance to partially overlapping groups of aminoglycoside antibiotics

Miloje Savic¹, Josip Lovric^{1,*}, Tatjana Ilic Tomic², Branka Vasiljevic² and Graeme L. Coan^{3,*}

Molecular Microbiology (2002) 67(4), 1044–1073

doi:10.1111/j.1365-2958.2002.04754.x

A new mechanism for chloramphenicol, florfenicol and clindamycin resistance: methylation of 23S ribosomal RNA at A2503



Detection Of Methyltransferase Activity

- In the assay, purified ribosomes and enzymes are used
- rRNA is then digested by a specific Rnase
- Small products are then analyzed with MALDI-TOF MS
- Methylation causes increasing of the mass by 14 Da
- **Still for research use only**



Antibiotic Resistance Determination By a Detection Of Specific Peaks In Mass Spectra



MRSA Detection

- Difference in mass spectra comparing MRSA and MSSA
- Direct detection of PBP2a has not been yet published

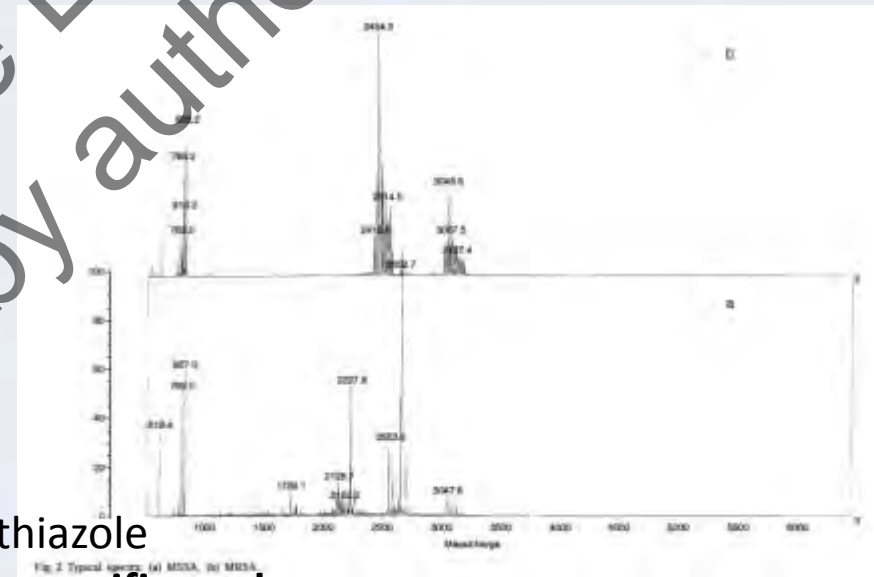
J. Med. Microbiol., Vol. 49 (2008), 295-300
© 2008 The Pathological Society of Great Britain and Ireland
ISSN 0950-2688

TECHNICAL NOTE

Rapid discrimination between methicillin-sensitive and methicillin-resistant *Staphylococcus aureus* by intact cell mass spectrometry

VALÉRIE EDWARDS-JONES*, M. A. CLAYDON*, D. J. EVASON*, J. WALKER†, A. J. FOX†, H. K. D. B. GORDON*

- Intact cells
- **Matrix:** 5-chloro-2-mercapto-benzothiazole
- **14 MRSA-specific peaks and 2 MSSA-specific peaks**





MRSA Detection



- **SELDI-TOF MS:** surface-enhanced laser desorption/ionization time-of-flight mass spectrometry
- Protein Chip Array is used

- **Specific preparation of sample via lysis solution**
- **Matrix:** sinapinic acid

MSSA specific peaks	MRSA specific peaks
3 081	5 709
5 893	7 694
9 580	15 308
	18 896



Antibiotic Resistance Determination By a Detection Of Specific Peaks In Mass Spectra

- **Specific resistance markers should be established for common resistant bacteria** (e.g., MRSA, VRE, Gram-negatives?, multidrug-resistant *Mycobacterium tuberculosis*?)
- Modification of sample preparation process
- **This methodology can be a useful tool for a quick resistance mechanism detection** (during bacterial identification?)



Proteomic Approach In the Determination Of Antibiotic Resistance

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Proteomic Approach

- **Proteome:**
the entire **PROTE**in complement expressed by a **genOME**,
or by a cell or tissue type
- **Methodology:**
 - Cell lysis – specific or non-specific
 - Electrophoretic separation (usually 2D – isoelectric focusing/SDS-PAGE)
 - Comparison of protein profiles
 - Identification of proteins by MALDI-TOF MS
 - peptide mass fingerprinting
 - amino acid sequencing



Proteomic Approach

Example of 2D gel electrophoresis:

Mass ↑

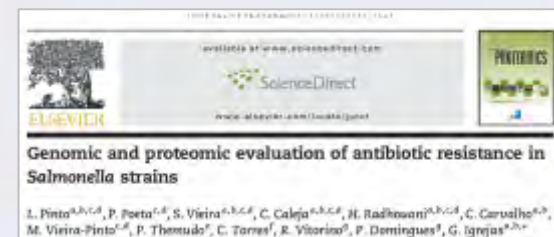
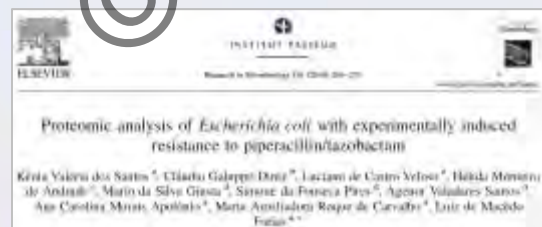
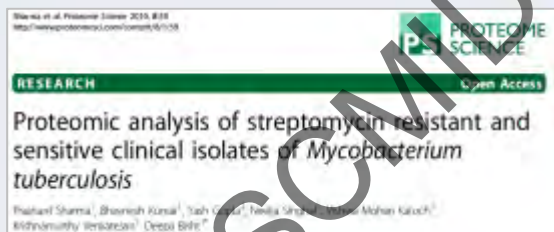
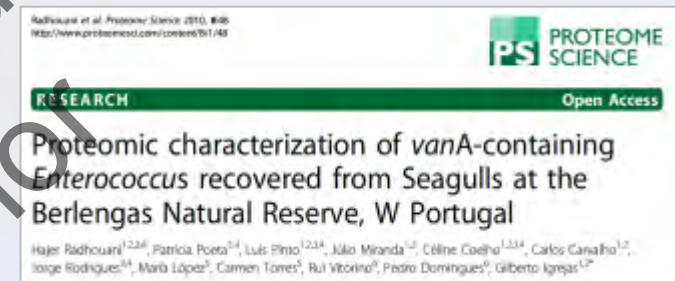


→ pI



Proteomic Approach In Antibiotic Resistance

- vancomycin-resistant *Enterococcus* spp. isolates recovered from seagulls
- ampicillin-resistant *Escherichia coli*
- ampicillin-resistant *Fusobacterium nucleatum*
- protein identification in resistant *E. coli*, *Salmonella enterica*, *Mycobacterium tuberculosis* strains.



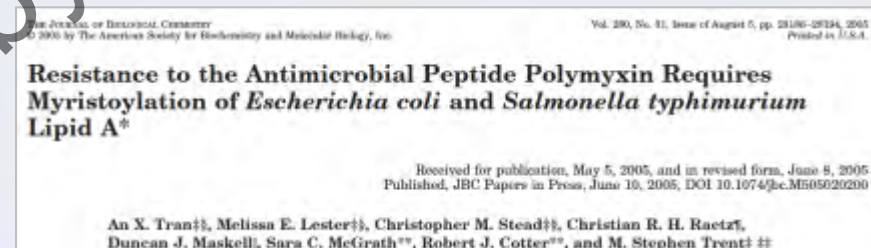
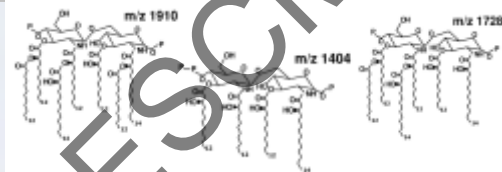
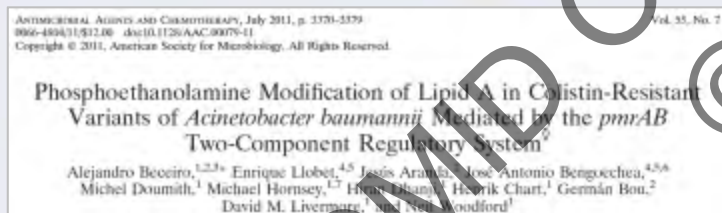
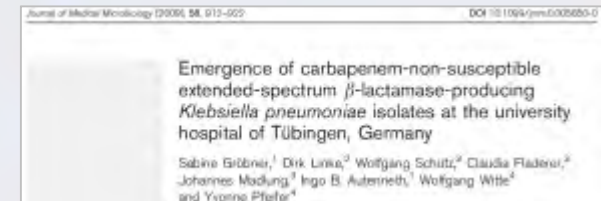


MALDI-TOF mass spectrometry for the detection of antimicrobial resistance mechanisms



Proteomic Approach – Cell-Wall Components

- **Porin identification** – resistance to carbapenems
 - Proteomic identification of porins
- **Efflux identification**
 - Proteomic identification of efflux pumps
- **Lipopolysaccharide** – resistance to polymyxins
 - Detection of Lipid A modification (activity of 4'-phosphatase)





Proteomic Approach In Antibiotic Resistance

- **Needs for a standardization of cultivation** (expression of resistance determinants)
- **Development of specific extraction methods** (e.g., for cell-wall components, lipopolysaccharide)
- **Labor intensive method**
- **More complex picture of resistant determinants**
- **Should be available in reference centres (???)**



MALDI-TOF MS-based DNA Sequencing (Minisequencing)

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MALDI-TOF mass spectrometry for the detection of antimicrobial resistance mechanisms



Minisequencing By MALDI-TOF MS

- SNP genotyping
- Labor intensive
- Length limitation

MALDI-TOF mass spectrometry-based SNP genotyping



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Pharmacogenomics (2002) 3(6) 537-540

537

Table 1. Mass differences resulting from single nucleotide exchanges.

Wild type	Mutant	Mass difference	Wild type	Mutant	Mass difference
A	C	+ 24 Da	C	A	- 24 Da
A	G	+ 16 Da	G	A	- 16 Da
A	T	+ 9 Da	T	A	- 9 Da
C	G	+ 40 Da	C	A	- 40 Da
C	T	+ 15 Da	T	C	- 15 Da
G	T	- 25 Da	T	G	+ 25 Da

DNA preparation

DNA amplification

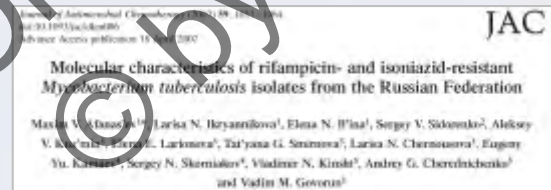
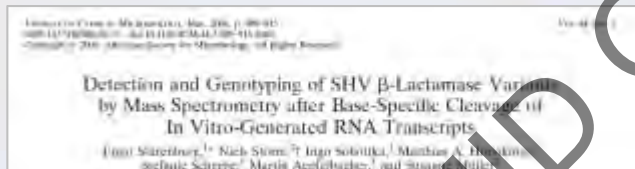
Purification

Allele-specific reaction

Purification

Data acquisition

Automated allele calling





Future Perspectives

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Future Perspectives

- Quantification of the molecules
- Selective extraction

Anal. Chem. 2007, 79, 3401–3408

Functionalized Magnetic Nanoparticles for Small-Molecule Isolation, Identification, and Quantification

Po-Chiao Lin,[†] Mei-Chun Tseng,[‡] An-Kai Su,[†] Yu-Ju Chen,^{*,†} and Chun-Cheng Lin^{*,†}

Option 1: Direct analysis of small molecule → MALDI analysis

Option 2: Affinity extraction of small molecule from biofluid → MNP@matrix-probe extraction → MALDI analysis

Legend: Magnetic Nanoparticle, Target Carbohydrate, Matrix, Probe protein

Analytica Chimica Acta 607 (2011) 1–7

Contents lists available at ScienceDirect

Analytica Chimica Acta

Journal homepage: www.elsevier.com/locate/aca

Nanoparticle-assisted MALDI-TOF MS combined with seed-layer surface preparation for quantification of small molecules

Yi-Chi Ho^a, Mei-Chun Tseng^b, Ying-Wei Lu^c, Chun-Cheng Lin^c, Yu-Ju Chen^{b,*}, Ming-Ren Fuh^{b,**}

1. dry
2. reconstituted in H₂O

Legend: Analyte, Endogenous Components, DHB@MNP

Fig. 1. Schematic representation of sample identification and quantification screening by MALDI-TOF MS.

Analytica Chimica Acta 681 (2011) 10–18

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Analytica Chimica Acta

Journal homepage: www.elsevier.com/locate/aca

Quantification in MALDI-TOF mass spectrometry of modified polymers

Zuzana Walterová, Jiří Horák^{*}



Future Perspectives

- **Detection of enzymes that degrade antibiotic molecules** through the direct detection of antibiotic modifications;
- **Detection of the resistance mechanism determinants** (e.g., Qnr proteins and modified PBPs) and **proteomic studies** on multi-resistant bacteria that allow the construction of a complete database of protein fingerprints of resistant isolates for detection of the main proteins responsible for resistance;
- **Analysis of the modification of target sites** (e.g., ribosomal methylation);
- **Quantification of antibiotics.** Developing a method for the determination of antibiotic concentrations could aid in the analysis of influx and efflux of an antibiotic.



Conclusion

- **Very fast and cheap detection of antimicrobial resistance**
- **New applications are expected**
- **More complex view on determining the resistance mechanisms (should be available in the national reference centres?)**

Applying this technique to determine resistance mechanisms cannot replace standard susceptibility testing.



Acknowledgements



MALDI-TOF MS team:

Tamara Bergerová, M.D.

Eva Chudáčková, M.D.

Radka Walková, M.D.

Vendula Študentová, BSc.

Thank you for your attention