

# Evaluation of Molecular Assays for Norovirus Detection

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## INTRODUCTION

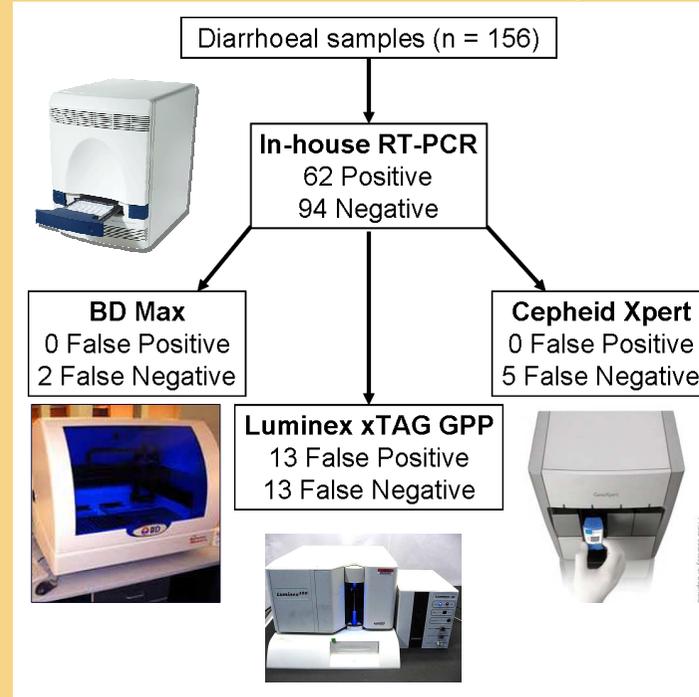
- Norovirus (NoV) exerts a considerable clinical and financial impact upon healthcare globally
- Laboratory diagnosis must be prompt and accurate to reduce the impact of NoV; molecular tests are best placed to provide this

● **AIM:** compare different molecular options and assess a prototype point-of-care assay for NoV detection

## METHODS

- One hundred and fifty six patient samples (87 female, 69 male) were tested by 3 methods and compared to the in house reverse-transcriptase real-time PCR (RT-PCR)
- All samples were faeces from individuals with diarrhoea; 137 were from hospital locations and 19 from primary care
- Samples were tested retrospectively during January to April 2013 (n = 131) and a small subset were tested prospectively from July 2013 (n = 25)
- An aliquot of liquid sample was suspended in 1 mL sterile water, centrifuged and then 200 µL of the supernatant was extracted on the easyMag (bioMérieux) and eluted into 100 µL. RT-PCR was then performed on the ABI 7500 (Applied Biosystems)
- The three evaluated test methods were:
  - BD MAX™ RNA Extraction Kit (Becton Dickinson) with PCR performed with Express one-step qRT-PCR kit (Life Technologies). Samples were processed as above prior to testing
  - xTAG® Gastrointestinal Pathogen Panel (GPP; Luminex). An aliquot of sample was placed in a Burton bead tube, vortexed and centrifuged prior to easyMAG extraction
  - Prototype GeneXpert® Norovirus Assay (Cepheid). A swab was inserted into the liquid stool, homogenised in sample reagent then transferred to the reaction cartridge and loaded onto the GeneXpert

**Figure 1** Results of testing diarrhoeal samples (n =156) on three platforms. In house real-time RT-PCR was considered gold standard



**Table 1** Performance\* of three commercial molecular Norovirus assays in patient samples (n = 156)

Test	Sensitivity	Specificity	NPV	PPV	Accuracy
<b>BD MAX</b>	<b>96.8</b> (92.4 - 100)	<b>100</b> (96.8 - 100)	<b>97.9</b> (95.1 - 100)	<b>100</b> (95.0 - 100)	<b>98.7</b> (97.0 - 100)
<b>xTAG GPP</b>	<b>79.0</b> (68.9 - 89.2)	<b>86.2</b> (79.2 - 93.1)	<b>86.2</b> (79.2 - 93.1)	<b>79.0</b> (68.9 - 89.2)	<b>83.3</b> (77.5 - 89.2)
<b>Xpert</b>	<b>91.9</b> (85.2 - 98.7)	<b>100</b> (96.8 - 100)	<b>94.9</b> (90.6 - 99.3)	<b>100</b> (94.7 - 100)	<b>96.8</b> (94.0 - 99.6)

\*Presented as percentage with 95% confidence interval

## RESULTS

- Results of laboratory testing are presented in Figure 1 and Table 1
- Of the 62 positive samples, 35 were GI and 27 were GII
- The BD Max had minimal hands-on-time, with results for 24 samples available 2h 30 min after receipt in the lab (2 h 10 min machine run time)
- The xTAG GPP required the most hands-on-time; results were available within 24 h for a full run of 86 test samples
- The Xpert required the least hands on time (3 min per sample), with machine run time of 1 h 30 min

## CONCLUSIONS

- The BD Max and Xpert assays gave acceptable performance for routine diagnosis of NoV
- The xTAG GPP was not considered acceptable for routine use, although the wide range of pathogens detected may prove beneficial in specific situations
- The Xpert assay offered simple processing and timely results which may enable point-of-care use in patient areas
- Point-of-care testing will likely reduce the time to results in high-incidence clinical areas, allowing rapid initiation of infection control measures and outbreak response

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