

ECOFFs

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**MIC wild type distributions and
epidemiological cut-off values**

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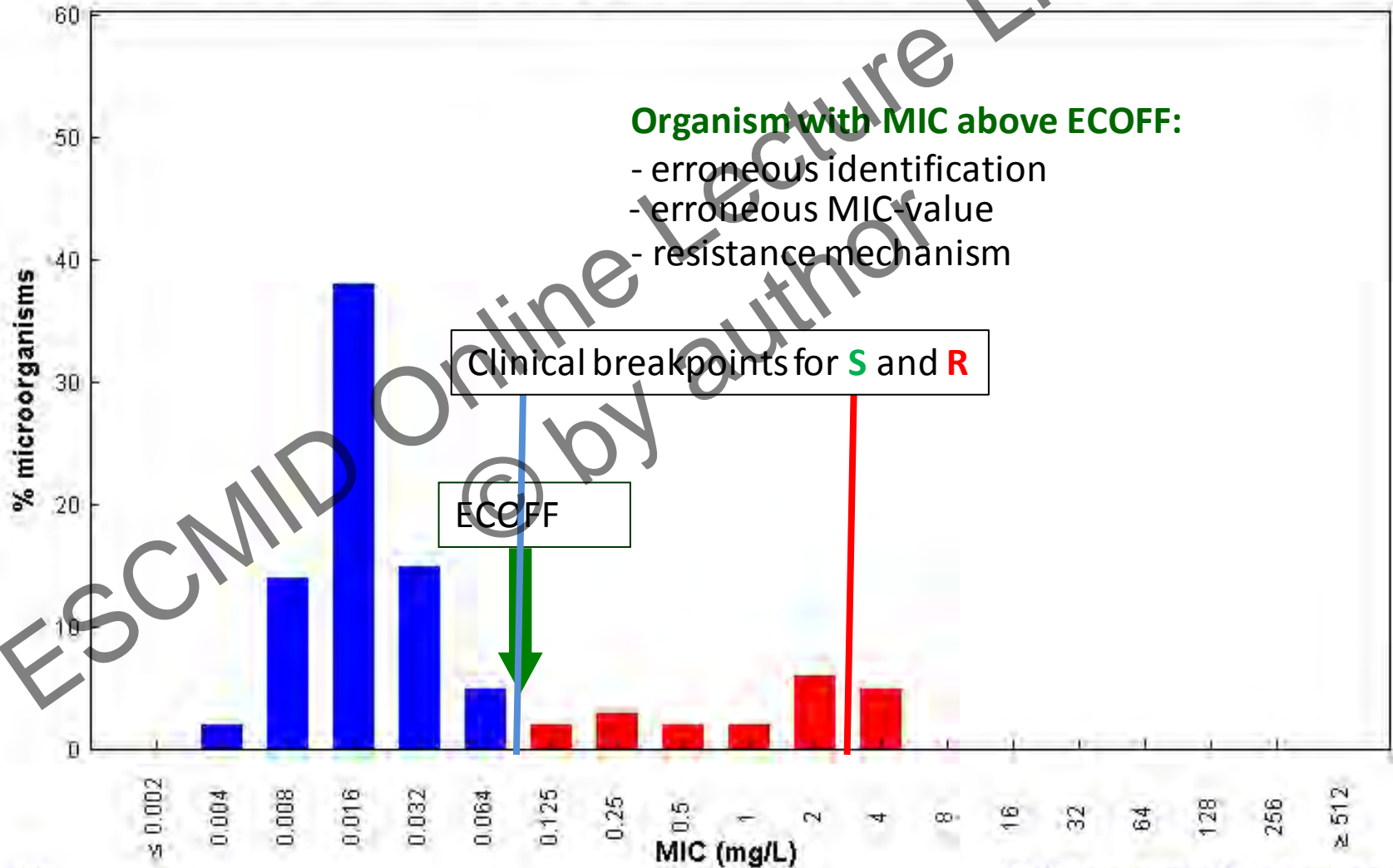
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Q1

Benzylpenicillin / Streptococcus pneumoniae

EUCAST MIC Distribution - Reference Database

MIC distributions include collated data from multiple sources, geographical areas and time periods and can never be used to infer rates of resistance



MIC
Epidemiological cut-off: WT ≤ 0.064 mg/L

EUCAST workshop, ECCMID 2013

37642 observations (32 data sources)
Clinical breakpoints: S ≤ 0.064 mg/L, R > 2 mg/L

Breakpoints

Clinical breakpoints

- MIC concentrations decided by man (based on clinical outcome data, MIC distributions, accepted dosing, PK/PD-data) to distinguish treatable from non-treatable organisms.
- should not divide wild type MIC distributions of target organisms.
- may render wild type organisms Susceptible (S), Intermediate (I) or Resistant (R)
- may change with a change in circumstances (indications, new resistance mechanisms, new dosing)

Breakpoints

ECOFF

- The ECOFF distinguishes between organisms **without** and **with** phenotypically expressed resistance mechanisms for a species and a drug in a defined test system.
- Within a species, it is the highest MIC for organisms lacking phenotypically expressed resistance.
- Organisms **without** resistance mechanisms (WT) are not by default treatable and organisms **with** resistance mechanisms (NWT) are not by default resistant.
- In a species considered susceptible (S) to the agent, the ECOFF is the lowest possible S breakpoint.

Establishing ECOFFs

- **ECOFFs should only be determined on**
 - MICs determined with methods calibrated to the internationally agreed standard method for broth microdilution (ISO)
 - large numbers of MICs ($n > 100$, ideally > 1000)
 - MICs from different places (minimum 3)
 - MICs performed by many investigators (minimum 3, ideally 10 or more)

S.pneumoniae vs ciprofloxacin

MIC-value	0.002	0.004	0.008	0.016	0.032	0.064	0.125	0.25	0.5	1	2	4	8	16	32	64	128	256	512
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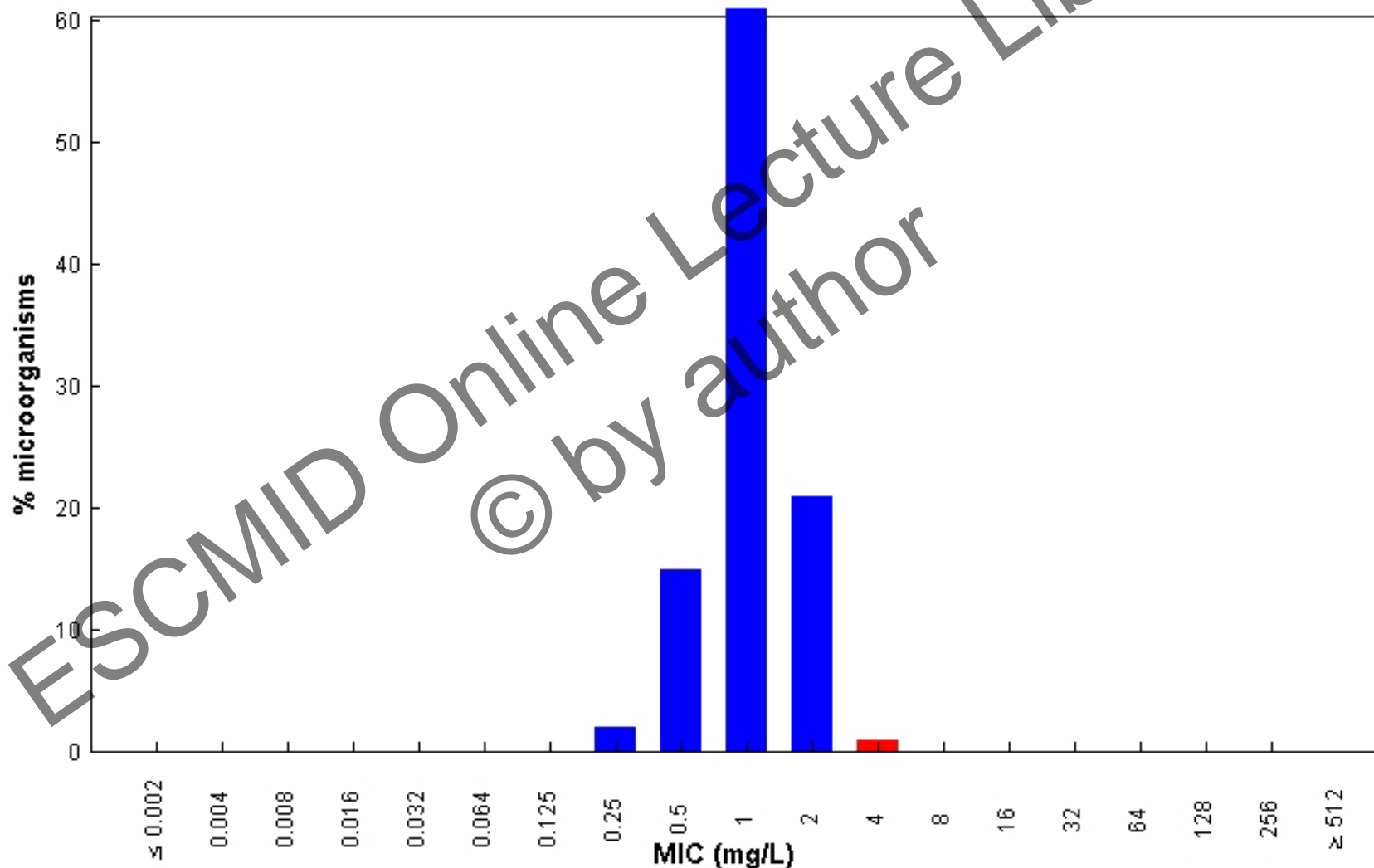
Tentative rules for aggregating MIC distributions and determine ECOFF

MIC distributions*	Distributions which disagree**	Action (none, aggregate or aggregate and determine ECOFF)
1	0	None***
2	0	None
3	0	Aggregate 3
3	1	None
4	0	Aggregate 4 and determine ECOFF
4	1	Aggregate 3
4	>1	None
5	0	Aggregate 5 and determine ECOFF
5	1	Aggregate 4
5	>1	None
6	0	Aggregate 6 and determine ECOFF
6	1	Aggregated 5 and determine ECOFF
6	>1	Aggregate 4
7	0	Aggregate 7 and determine ECOFF
7	1	Aggregate 6 and determine ECOFF
7	2	Aggregate 5 and determine ECOFF
7	>2	None
etc		

Ciprofloxacin / *Streptococcus pneumoniae*

EUCAST MIC Distribution - Reference Database 2011-02-04

MIC distributions include collated data from multiple sources, geographical areas and time periods and can never be used to infer rates of resistance



MIC
Epidemiological cut-off: WT ≤ 2 mg/L

73840 observations (50 data sources)
EUCAST workshop, ECCMID 2013 Clinical breakpoints: S ≤ 0.125 mg/L, R > 2 mg/L

Organization

Clinical breakpoints

Expert rules

MIC - distributions and QC

Zone diameter distributions

EUCAST disk diffusion test

Frequently Asked Questions (FAQ)

Meetings

EUCAST Presentations

Documents

Information for industry

Links

Website changes



search term
MIC - distributions

MIC distributions

[Link to the website with MIC distributions](#)

The website gives MIC distributions (and since 2010 inhibition zone diameter distributions generated with the new EUCAST disk diffusion method) for a wide range of organisms and antimicrobial agents in tables. The distributions are based on collated data from a total of close to 20000 MIC distributions from worldwide sources. The distributions include MICs from national and international studies such as resistance surveillance programs (Alexander, BSAC, ECO-SENS, MYSTIC, NORM and SENTRY), as well as MIC distributions from published articles, the pharmaceutical industry, veterinary programmes and individual laboratories. Histograms display wild type organisms, together with EUCAST clinical breakpoints and epidemiological cut-off values (ECOFFs). The distributions should never be referred to in any epidemiological context since data from many time periods and many countries have been aggregated.

MIC OC values

[MIC OC values](#)

[Recommend page](#)

EUCAST workshop, ECCMID 2013

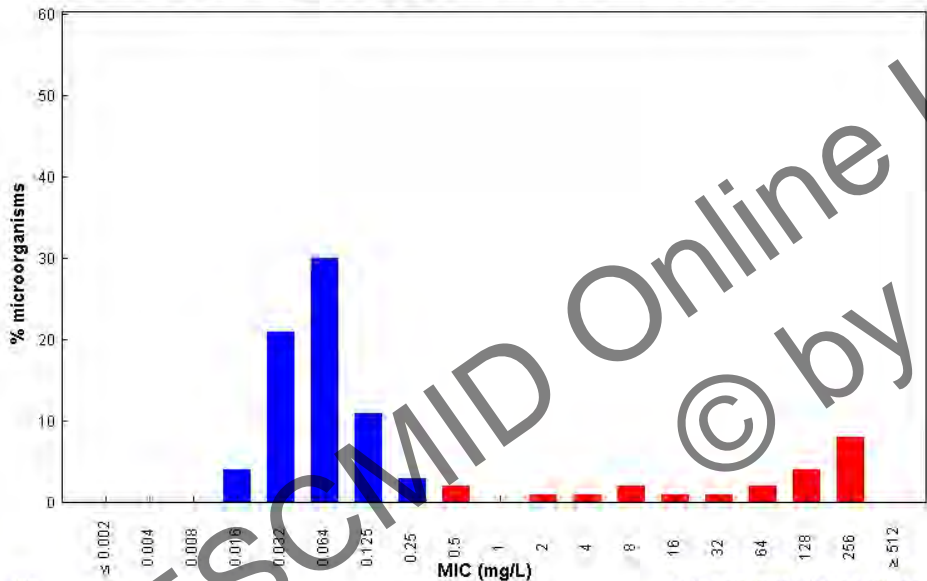
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Distribution of MICs and inhibition zone diameters defined by agent, species and test systems are freely available on the internet (www.eucast.org)

Cefotaxime / *Klebsiella pneumoniae*
EUCAST MIC Distribution - Reference Database 2011-02-05

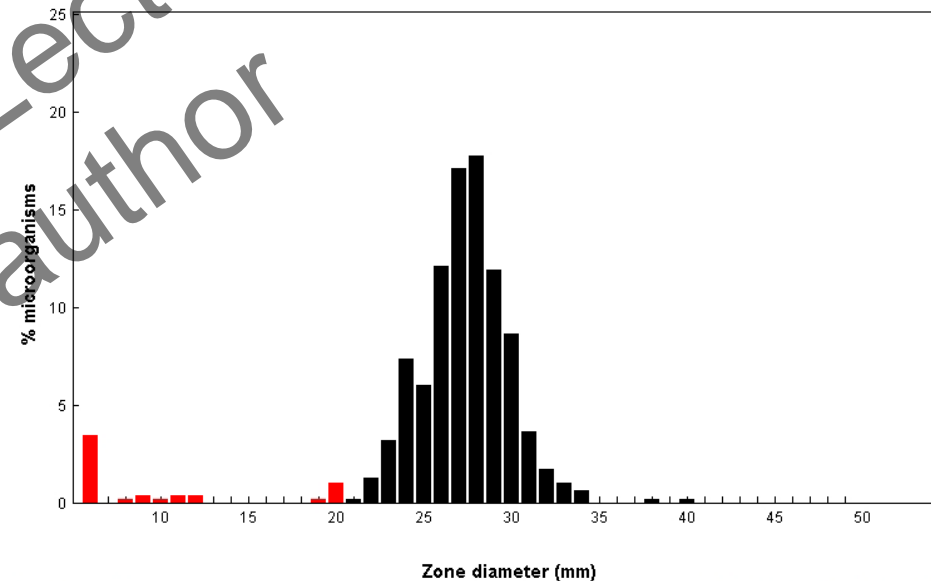
MIC distributions include collated data from multiple sources, geographical areas and time periods and can never be used to infer rates of resistance



MIC
 Epidemiological cut-off: WT ≤ 0.25 mg/L
 2415 observations (10 data sources)
 Clinical breakpoints: S ≤ 1 mg/L, R > 2 mg/L

Cefotaxime / *Klebsiella pneumoniae*
EUCAST zone diameter distribution - Reference database
EUCAST disk diffusion method

Distributions include collated data from multiple sources, geographical areas and time periods and can never be used to infer rates of resistance



Disk content: 5
 Epidemiological cut-off: WT ≥ 21 mm (MIC: ≤ 0.125 mg/L)
 461 observations (2 data sources)
 Clinical breakpoints: S ≥ 21 mm, R < 18 mm

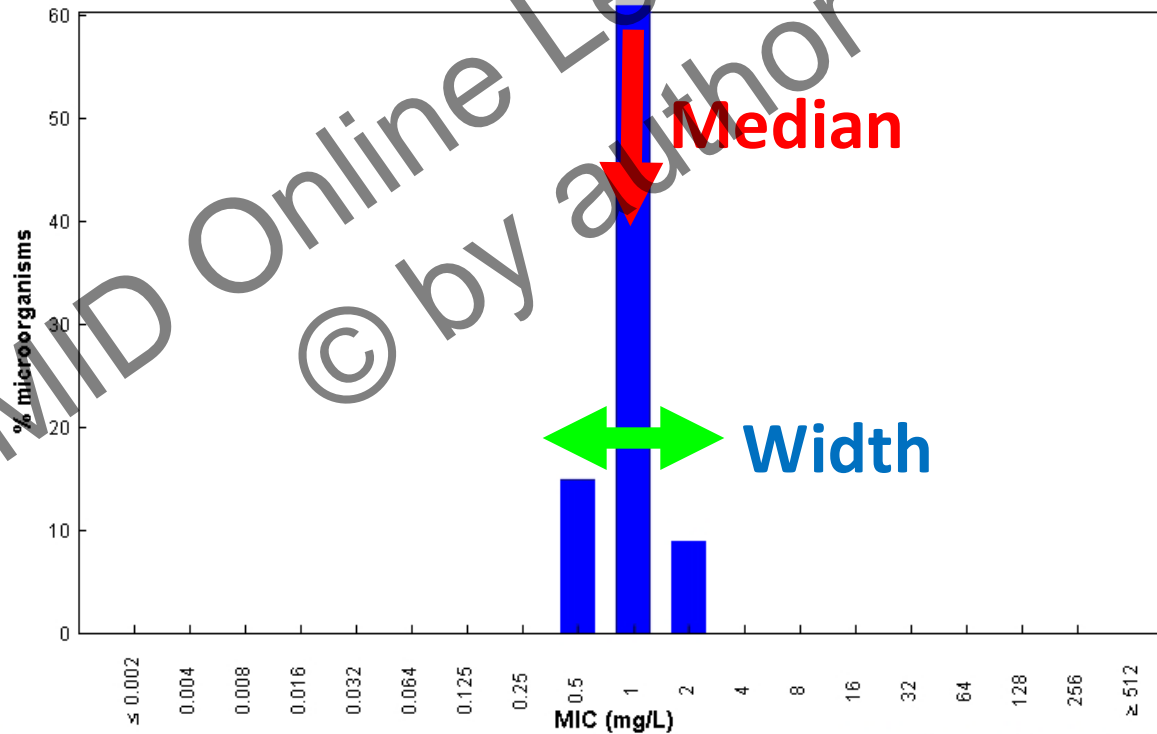
MIC distributions and ECOFFs on EUCAST website

- 25 000 MIC distributions
- Up to 100 000 MIC values per distribution
- Data from many investigators (1 – 100 per distrib.)
- Data from many time periods (1950 -)
- Data from many geographical areas and projects (USA, Europe, Australia, Far East, South America, Sentry, Mystic, etc)
- Data of multiple origin (Human clinical data, Surveillance programs, Veterinarian data, Wild life, Food safety programs)
- Database secure on three servers in different parts of Dusseldorf under the official responsibility of EUCAST and ESCMID.
- Ownership:
 - Software and administration: ECDC / ESCMID
 - Database: individual ownership of original data

Factors influencing the median and width of WT distributions

Vancomycin / *Staphylococcus aureus*
EUCAST MIC Distribution - Reference Database 2011-04-21

MIC distributions include collated data from multiple sources, geographical areas and time periods and can never be used to infer rates of resistance

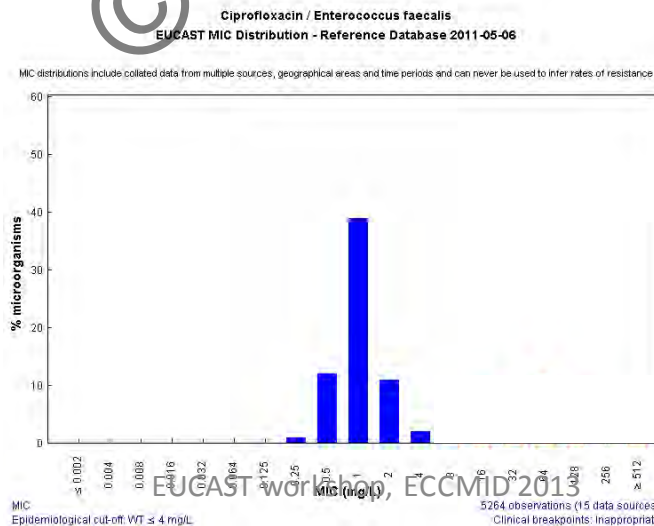
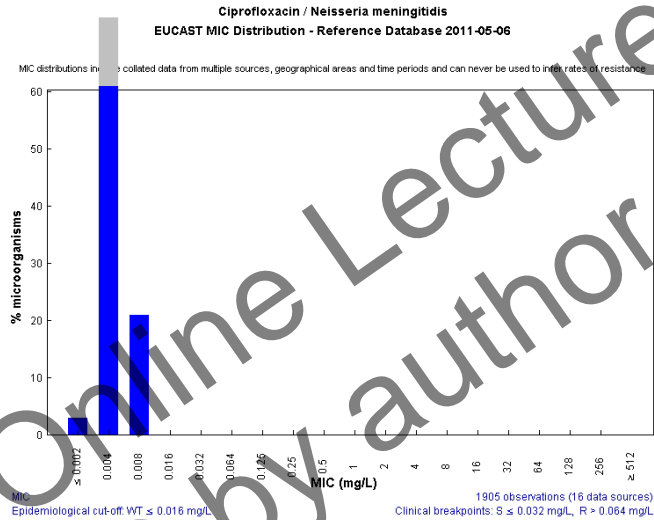


MIC
Epidemiological cut-off: WT ≤ 2 mg/L

87764 observations (33 data sources)
Clinical breakpoints: S ≤ 2 mg/L, R > 2 mg/L

The **median** of the MIC distribution:

- The inherent susceptibility of the species to the drug



The same applies to zone diameter distributions

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The **median** of the MIC distribution:

- The inherent susceptibility of the species to the drug
- Anything systematically influencing the activity of the drug:
 - Medium – variation in MICs depending on medium
 - Inoculum – increasing MICs with higher inocula
 - Incubation – increasing MICs with longer incubation
 - Atmosphere – affects the activity of some drugs
 - pH – some drugs are more active at high pH, others at low

The **median** of the MIC (or zone diameter) distribution:

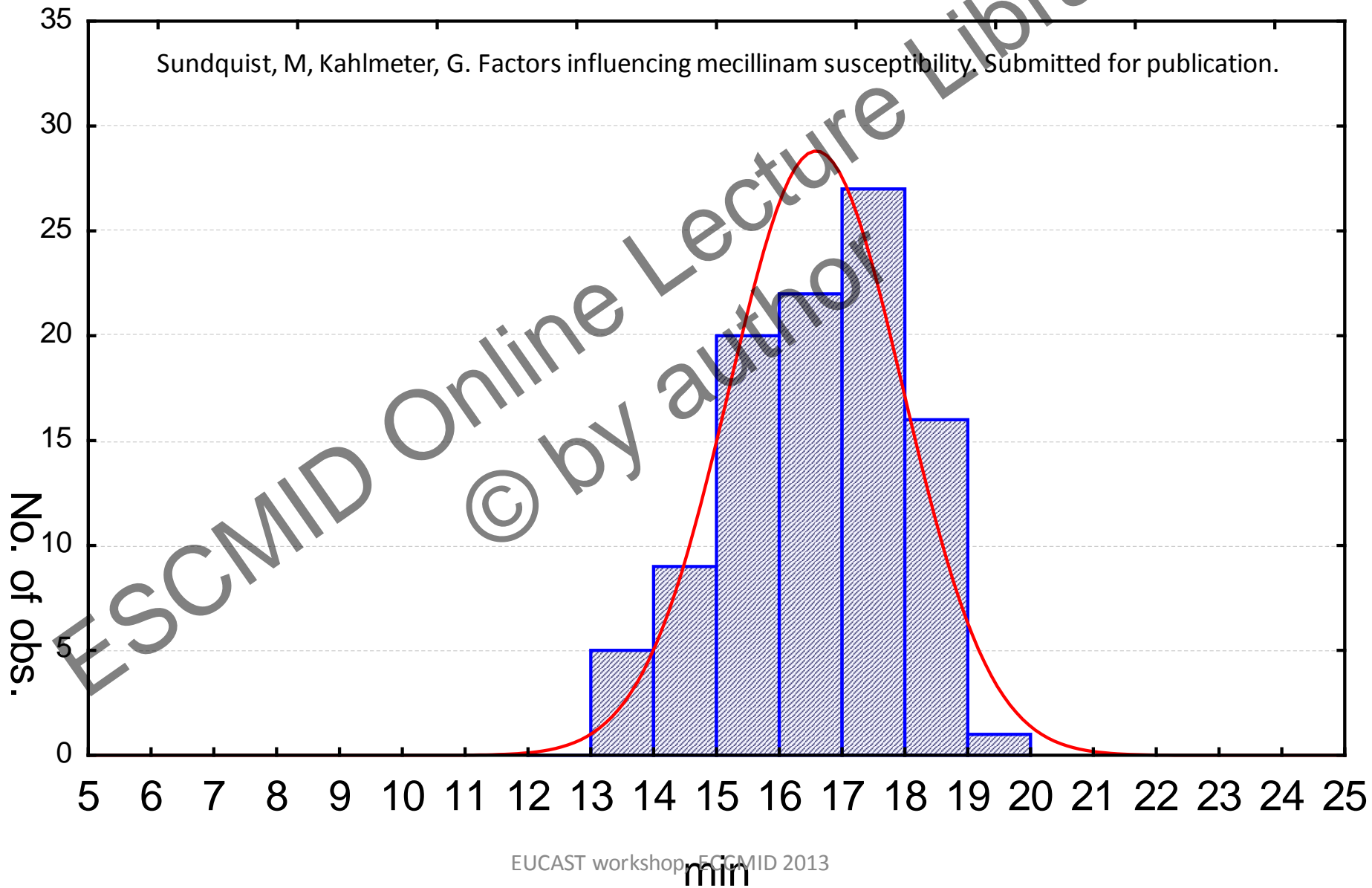
- The inherent susceptibility of the species to the drug
- Anything systematically influencing the activity of the drug
 - Medium, inoculum, pH, cations, incubation atmosphere and time,

The **width** of the MIC (or zone diameter) distribution:

- Inherent **variation in susceptibility** to the drug
- Biological **variation in other traits** that influence the MIC
 - any biological characteristic such as generation time, nutrient dependency, atmosphere dependency etc
- Exogenous **variation** randomly influencing the activity of the drug
 - pH, cations, incubation atmosphere and time, etc
- **Variation** in reading (between days, between readers, between systems)
- The stability of the molecule
- ...

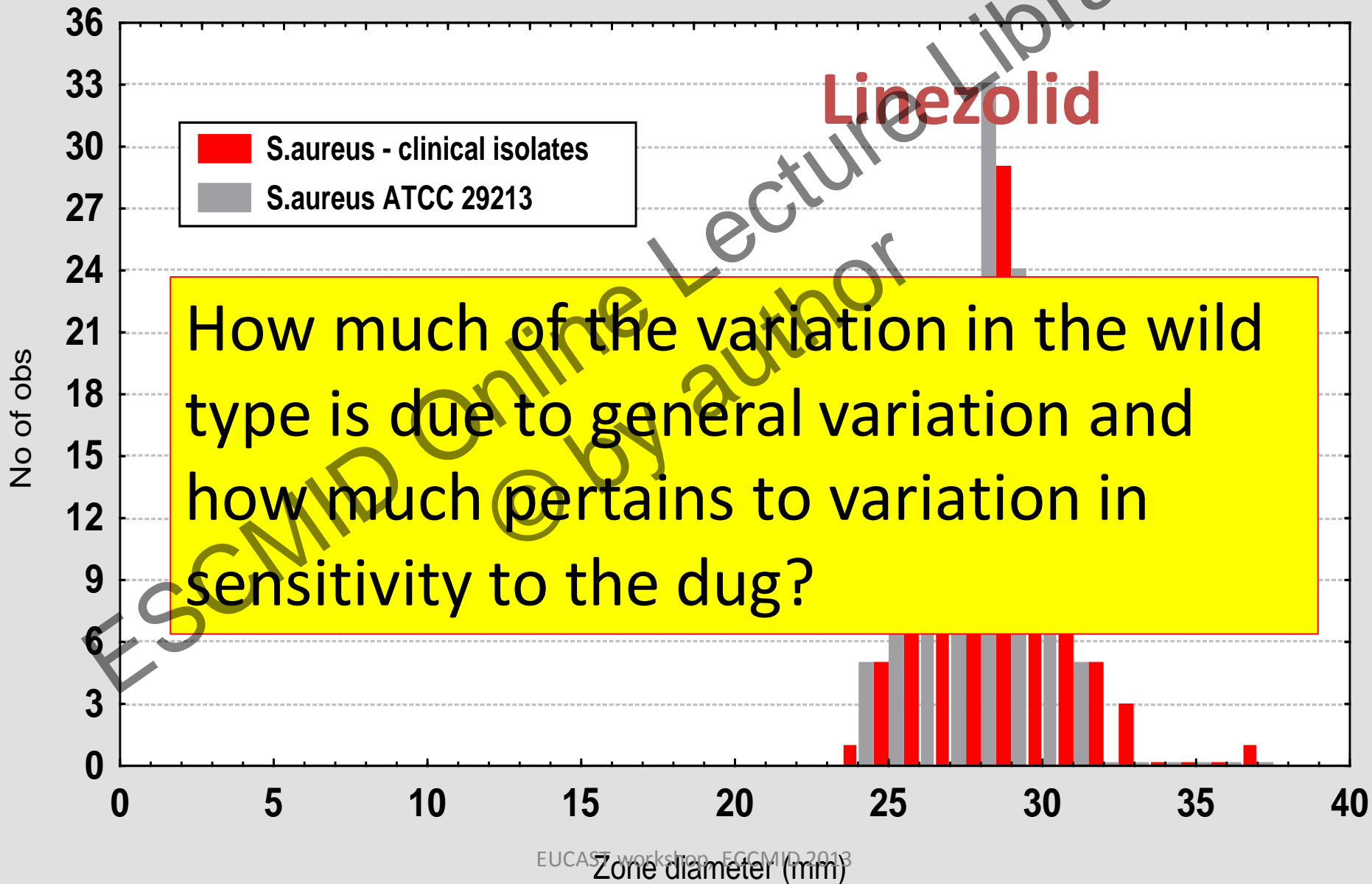
E. coli wild type; generation times (min) (n=100)

— Expected Normal

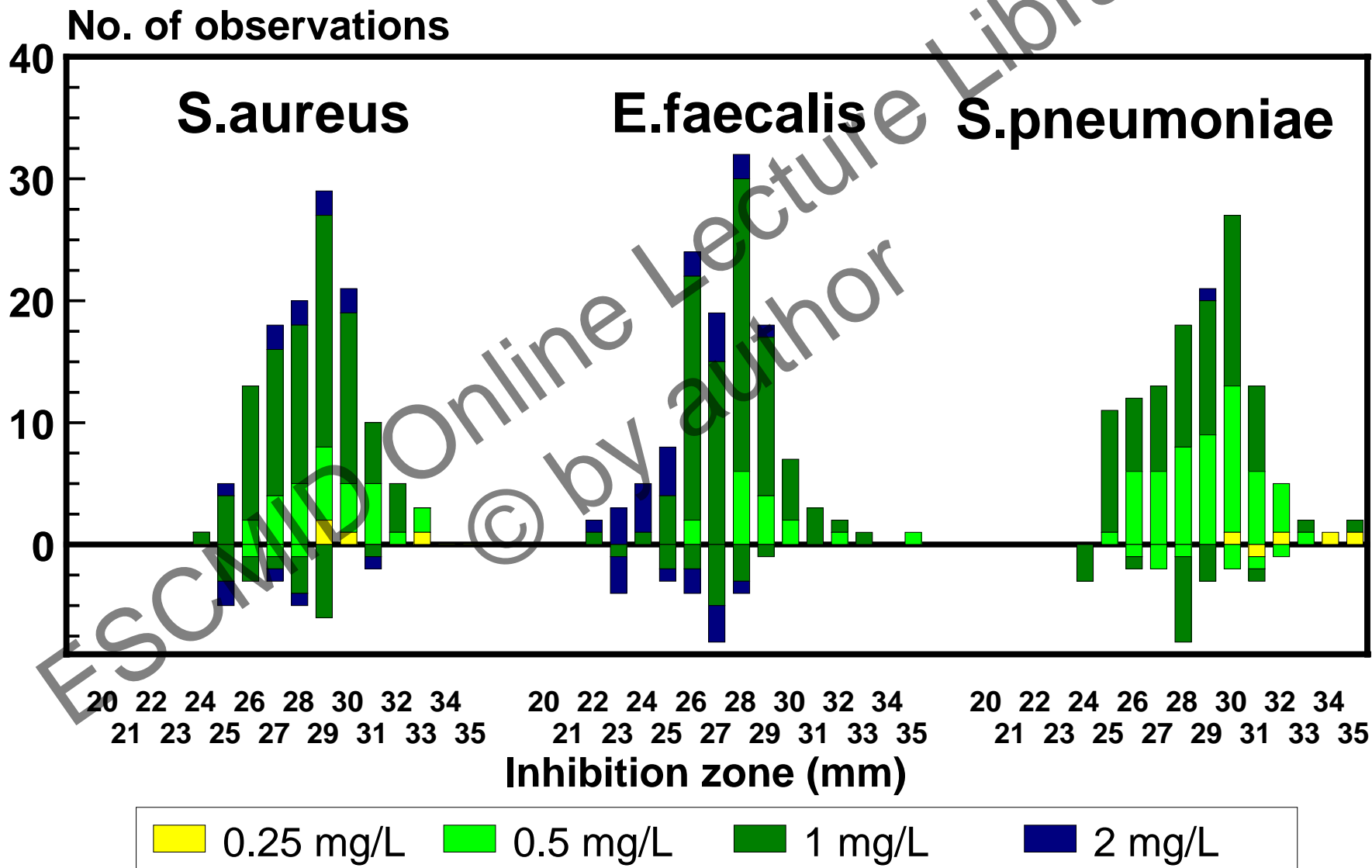


S.aureus - clinical isolates vs S.aureus ATCC 29213

125 clinical isolates vs. 125 determinations of one type strain



MIC distributions in zone diameter histograms Linezolid



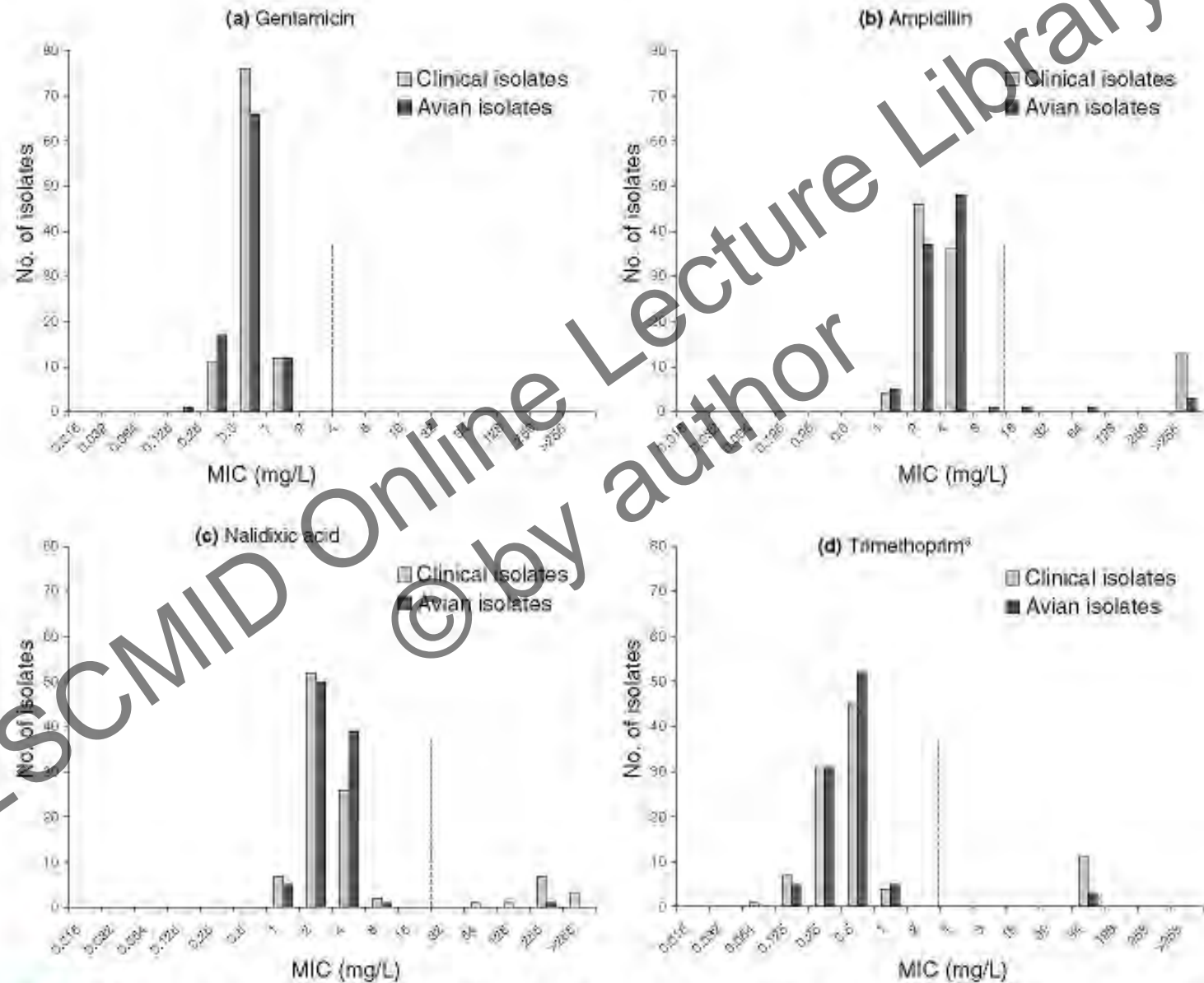
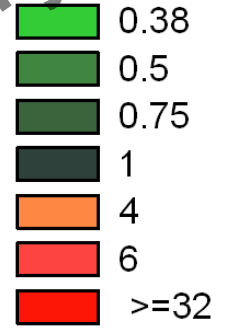


FIG. 2. MIC histograms for clinical and avian isolates. The dashed line indicates the epidemiological cut-off values (ECOFFs) for the wild-type distribution as determined by EUCAST (<http://www.eucast.org>). ^aTrimethoprim concentration test range: 0.016–32 mg/L.

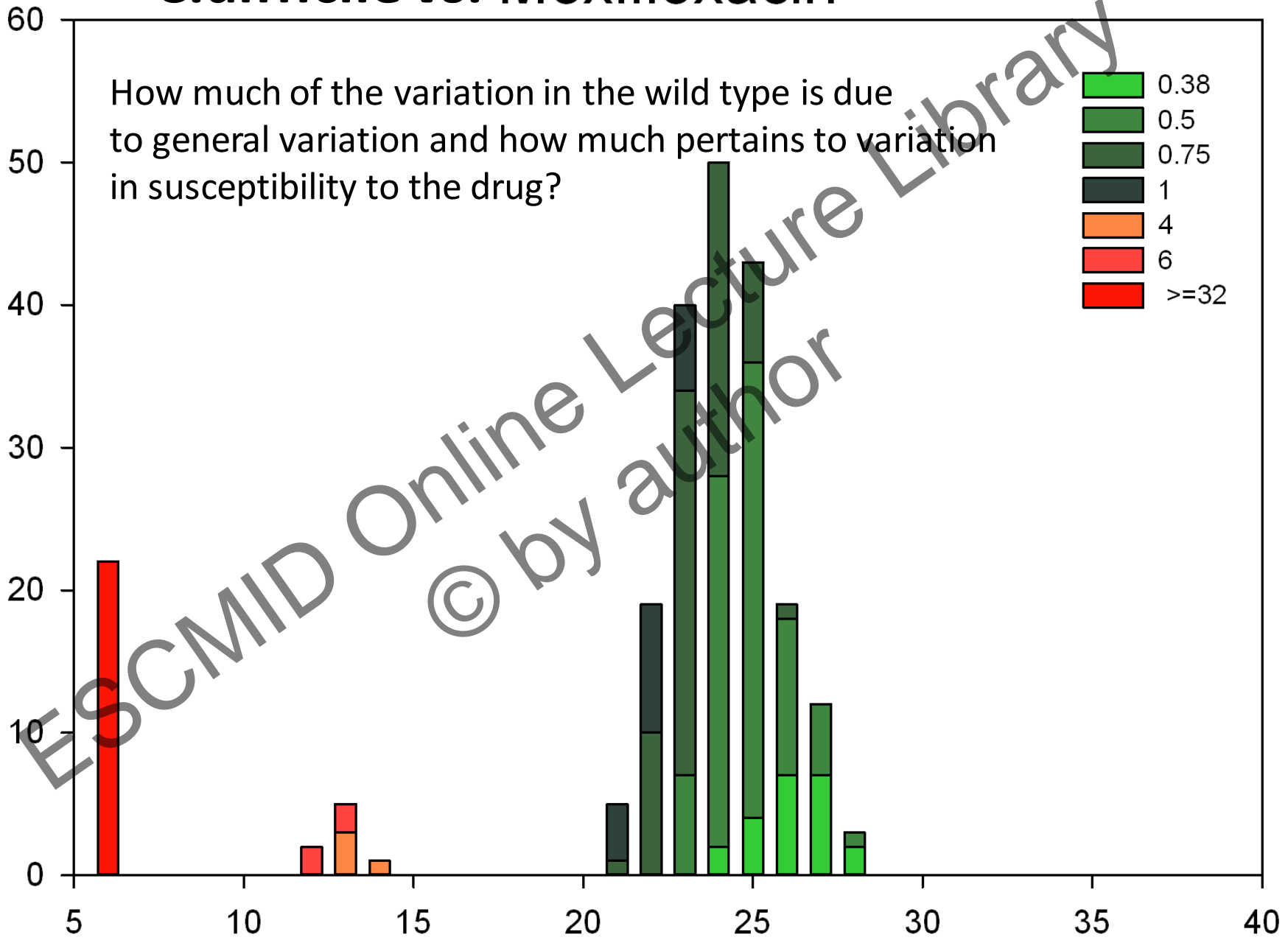
EUCAST workshop, ECCMID 2015

C.difficile vs. Moxifloxacin

How much of the variation in the wild type is due to general variation and how much pertains to variation in susceptibility to the drug?



No. of isolates



Methods for defining the wild type distribution (determining the ECOFF)

- The “eyeball” method (Kahlmeter)
- The 95% rule (Pfaller)
- The Normalised Resistance Interpretation (Kronvall)
- The iterative statistical method (Turnidge)
- Multimodal analysis (Meletiadis)

From John Turnidge, CLSI Workshop, Tampa, January 2013

The Normalized Resistance Interpretation

JOURNAL OF CLINICAL MICROBIOLOGY, Dec. 2010, p. 4445–4452
0095-1137/10/\$12.00 doi:10.1128/JCM.01101-10
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Vol. 48, No. 12

Normalized Resistance Interpretation as a Tool for Establishing Epidemiological MIC Susceptibility Breakpoints[∇]

Göran Kronvall*

*Department of Microbiology and Tumor Biology–MTC, Clinical Microbiology, Karolinska Institutet,
Karolinska University Hospital Solna, Stockholm, Sweden*

- Uses an adaptation of a method originally devised for “ECOFFs” for zone diameter distributions
 - Kronvall et al., Clin Micro Infect 2003

From John Turnidge, CLSI Workshop, Tampa, January 2013

Iterative Statistical Method

ORIGINAL ARTICLE

10.1111/j.1469-0691.2006.01377.x

Statistical characterisation of bacterial wild-type MIC value distributions and the determination of epidemiological cut-off values

J. Turnidge¹, G. Kahlmeter² and G. Kronvall³

¹Division of Laboratory Medicine, Women's and Children's Hospital, North Adelaide, South Australia, Australia, ²Department of Clinical Microbiology, Central Hospital, Växjö and ³Department of Microbiology and Tumour Biology, MTC, Clinical Microbiology, Karolinska Institute, Karolinska Hospital, Stockholm, Sweden

From John Turnidge, CLSI Workshop, Tampa, January 2013

Iterative Statistical Method - COFinder

Step 1. Population Data

CRGAT

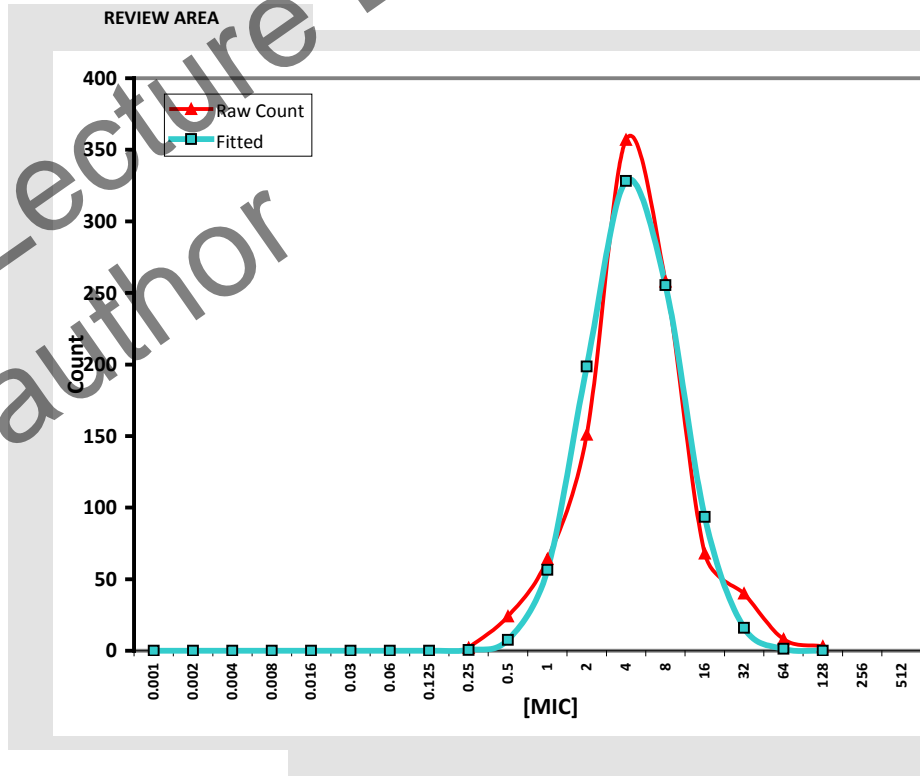
FLUC All

MIC	Log ₂ MIC	Raw Count	Cum. Count	Fitted
0.001	-10		0	0.0
0.002	-9		0	0.0
0.004	-8		0	0.0
0.008	-7		0	0.0
0.016	-6		0	0.0
0.03	-5		0	0.0
0.06	-4		0	0.0
0.125	-3		0	0.0
0.25	-2	2	2	0.5
0.5	-1	24	26	7.5
1	0	64	90	56.5
2	1	151	241	198.7
4	2	357	598	328.2
8	3	258	856	255.3
16	4	68	924	93.5
32	5	40	964	16.0
64	6	8	972	1.3
128	7	3	975	0.0
256	8		975	
512	9		975	
1024	10		975	

Modal MIC	4
Log ₂ MIC Mode	2
Max Log ₂ MIC	7

Selected Log ₂ Mean	1.6667 = 3.17
Selected Log ₂ SD	1.1145

Selected CO _{WT} Values		%>
CO _{WT} 95%	16	5.2%
CO _{WT} 97.5%	16	5.2%
CO _{WT} 99%	32	1.1%
CO _{WT} 99.9%	64	0.3%



?
Enter/paste
data
in this column

Data from 10 laboratories

Selected CO _{WT} Values		%>
CO _{WT} 95%	16	5.2%
CO _{WT} 97.5%	16	5.2%
CO _{WT} 99%	32	1.1%
CO _{WT} 99.9%	64	0.3%

From John Turnidge, CLSI
Workshop, Tampa,
January 2013

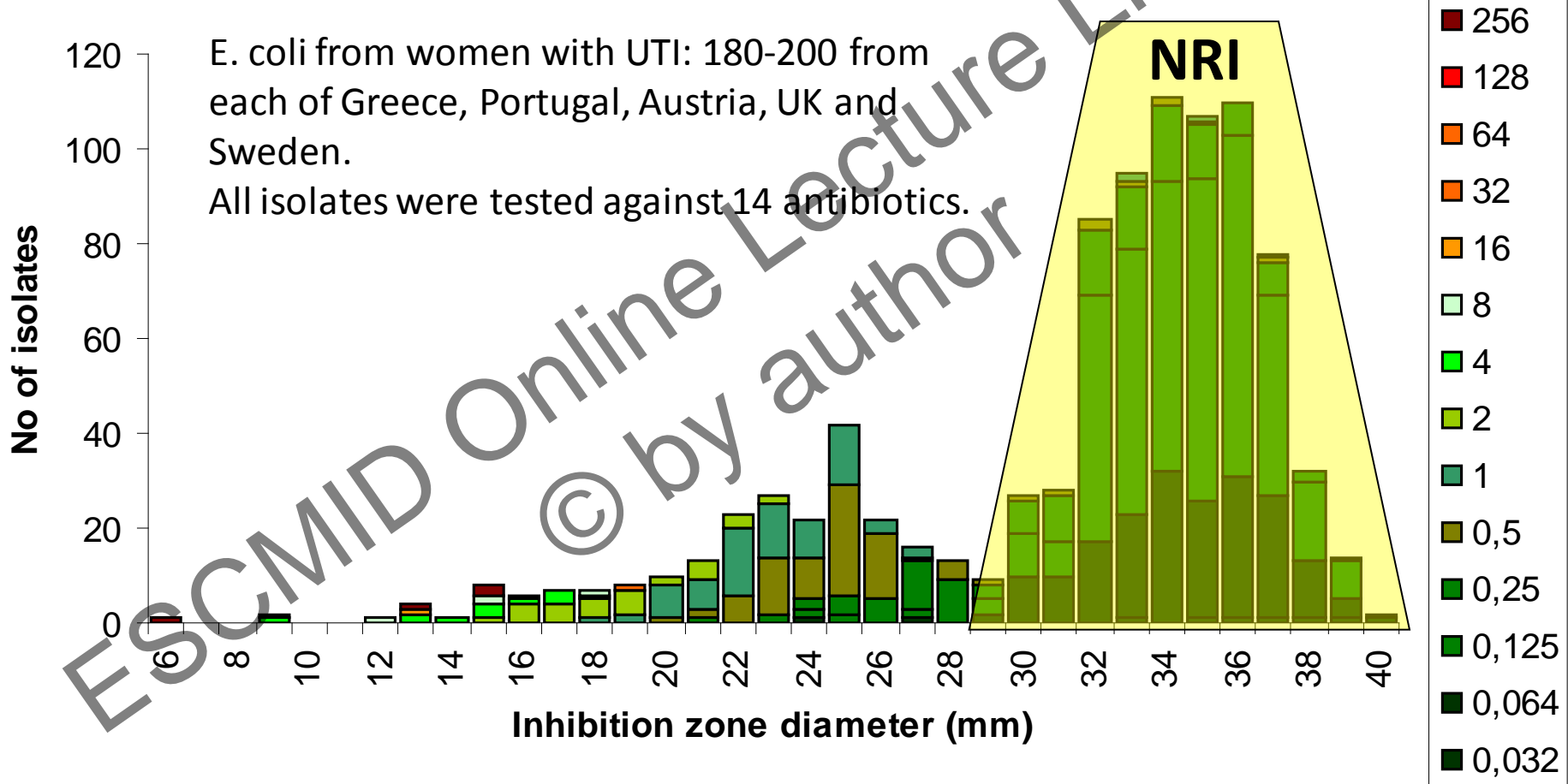
“A biological approach”

- Associated resistance – an organism resistant to one agent is very likely to carry resistance to other agents.
- By comparing the MIC distributions of
all isolates
and
isolates wild type to all other agents

The original distribution with all tested organisms

E. coli vs. *meccillinam*

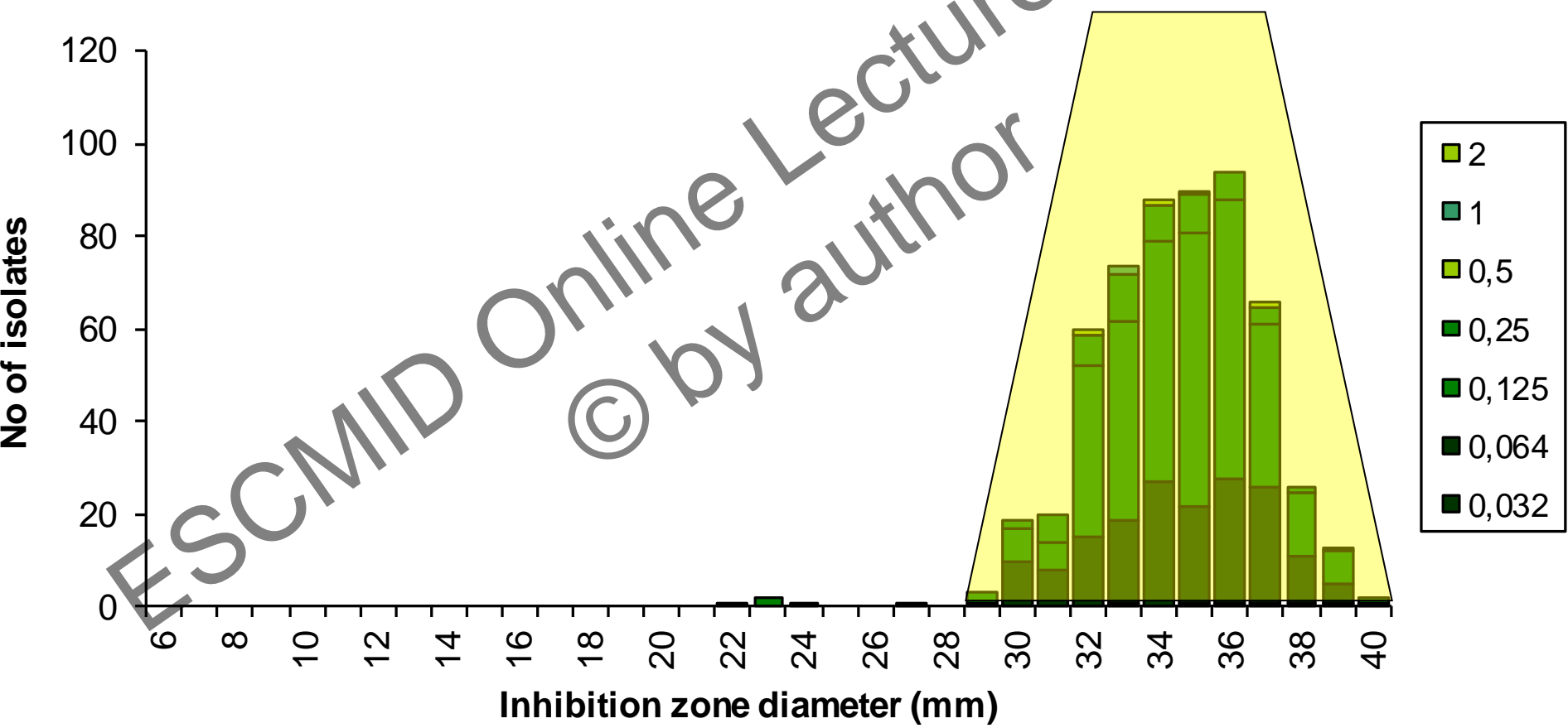
931 isolates from the ECOSENS II Study



The distribution when all organisms with resistance to other agents were removed.

E. coli vs. *mecillinam*

560 isolates with MICs below ECOFF for all other tested agents



The use of ECOFFs (1)

1. As a **tool in the determination of clinical breakpoints**
 - To avoid dividing wild type MIC distributions of target organisms
 - As a **surrogate clinical breakpoint when clinical data pertain only to wild type organisms (and when PK/PD data are incomplete).**
2. For **sensitive detection of (screening for) resistance**
 - oxacillin to detect all penicillin-R in *S. pneumoniae*
 - cefoxitin to detect methicillin resistance in *S. aureus*
 - benzylpenicillin to detect all betalactam resistance in *H. influenzae*
 - pefloxacin to detect quinolone resistance in *Salmonella* spp
 - meropenem to screen for KPC in Enterobacteriaceae


The use of ECOFFs (2)

3. For **surveillance of antimicrobial resistance** when clinical breakpoints...
 - have not been determined
 - are not sensitive enough
 - change over time
 - differ between systems (CLSI, FDA, EUCAST etc)
 - differ between humans, cows, pigs, birds, fish and camels.
4. To **exclude resistance**
 - To exclude methicillin resistance, carbapenem resistance etc
 - food safety – in the development of functional foods
5. For **clinical susceptibility reports?**

Is it possible to report the WT/NWT status of an isolate as supplementary information to S, I and R

- *E.faecalis* with MIC 16 mg/L to gentamicin would be reported: R^{WT}
- *E.coli* with MIC 0.5 mg/L to ciprofloxacin would be reported: S^{NWT}
- *E.faecalis* with MIC 1.0 mg/L to daptomycin would be reported: IE^{WT}
- *K.pneumoniae* with MIC 1.0 mg/L (and a KPC) to meropenem would be reported: S^{NWT}

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A large, bold, blue number '92' is centered on the page. The '9' has a thick stroke and a rounded top, while the '2' is also thick and has a horizontal base. The number is partially overlaid by a diagonal watermark.

