

# In vitro activity of aztreonam-avibactam against isolates of *Enterobacteriaceae* collected in Europe as part of a global surveillance program, 2015

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## Abstract

**Background:** Aztreonam-avibactam is a  $\beta$ -lactam/ $\beta$ -lactamase inhibitor combination being developed for use against carbapenem-resistant *Enterobacteriaceae*, especially isolates producing metallo- $\beta$ -lactamases (MBL). Aztreonam is stable to hydrolysis by MBL but inactivated by many serine  $\beta$ -lactamases. Avibactam is active against serine  $\beta$ -lactamases often co-carried with MBLs, including extended-spectrum  $\beta$ -lactamases (ESBL), AmpC  $\beta$ -lactamases, and serine carbapenemases. This study evaluated the *in vitro* activity of aztreonam-avibactam and comparators against *Enterobacteriaceae* collected in 2015 in Europe.

**Materials/methods:** Non-duplicate clinical isolates of *Enterobacteriaceae* were collected from 67 centres in 17 European countries. Susceptibility testing was performed using CLSI broth microdilution and interpreted using EUCAST breakpoints. Aztreonam-avibactam was tested at a fixed concentration of 4 mg/L avibactam. Multidrug resistant (MDR) was defined as resistant by EUCAST breakpoints to sentinel agents from three or more drug classes (cephalosporins, monobactams,  $\beta$ -lactam/ $\beta$ -lactamase inhibitor combinations, carbapenems, fluoroquinolones, aminoglycosides, glycolycyclines, and polymyxins). PCR and sequencing were used to determine the  $\beta$ -lactamase genes present in isolates with meropenem MIC  $>1$  mg/L, ceftazidime MIC  $>8$  mg/L, and those phenotypically positive for ESBL activity.

**Results:** Aztreonam-avibactam demonstrated good activity against *Enterobacteriaceae*, with an MIC<sub>90</sub> of 0.12 mg/L against the overall population and MIC<sub>90</sub>s of 0.5–1 mg/L against aztreonam non-susceptible, meropenem non-susceptible, colistin-resistant, MDR, and MBL-positive subsets (Table).  $>99.9\%$  of isolates (6445 of 6449), including those that produced MBLs, were inhibited by  $\leq 8$  mg/L of aztreonam-avibactam. In comparison, 83.0–96.2% of overall isolates were susceptible to meropenem, tigecycline, and colistin. Activity of these comparator agents was greatly reduced against resistant subsets with the exception of colistin, which was active against MBL-positive isolates (MIC<sub>90</sub> 1 mg/L, 96.7% susceptible). VIM- and NDM-type MBLs were found in 60 isolates of 5 species of *Enterobacteriaceae* collected in 10 European countries. 95.0% (n=57) of MBL-producing isolates co-carried one or more plasmid- or chromosomally-mediated ESBL, AmpC, or serine carbapenemase, including CTX-M-15, KPC-2 and OXA-48. No IMP-type MBLs were found in *Enterobacteriaceae* from Europe.

Species/Phenotype (n)	Drug (MIC <sub>90</sub> % Susceptible)					
	ATM-AVI	ATM	MER	CST	TGC	TGC
<i>Enterobacteriaceae</i> , All (6449)	0.12 NA <sup>a</sup>	64 75.4%	0.12 96.2%	$>8$ 83.0%	2 88.3%	2 88.3%
ATM-NS (1590)	0.5 NA	$>128$ 0.0%	$\geq 8$ 85.6%	4 89.8%	2 87.8%	2 87.8%
MER-NS (248)	1 NA	$>128$ 7.7%	$>8$ 0.0%	$>8$ 75.4%	2 76.2%	2 76.2%
CST-R (125) <sup>b</sup>	1 NA	$>128$ 24.0%	$>8$ 51.2%	$>8$ 0.0%	2 84.0%	2 84.0%
MDR (1029) <sup>c</sup>	0.5 NA	$>128$ 6.4%	$>8$ 77.2%	$>8$ 82.1%	4 80.0%	4 80.0%
MBL-negative (6389)	0.12 NA	64 75.9%	0.12 97.0%	$>8$ 82.9%	2 88.4%	2 88.4%
MBL-positive (60)	1 NA	$>128$ 21.7%	$>8$ 10.0%	1 96.7%	2 70.0%	2 70.0%

ATM-AVI, aztreonam-avibactam; ATM, aztreonam; MER, meropenem; CST, colistin; TGC, tigecycline; MDR, multidrug resistant; MBL, metallo- $\beta$ -lactamase. EUCAST breakpoints were used to determine % susceptible and to define non-susceptible (NS) and resistant (R) subsets.  
<sup>a</sup> NA, no breakpoints available  
<sup>b</sup> Excludes isolates of *Proteus* and *Serratia* spp.  
<sup>c</sup> MDR, resistant by EUCAST breakpoints to sentinel agents from 3 or more drug classes.

**Conclusions:** Aztreonam-avibactam was highly potent *in vitro* against *Enterobacteriaceae* collected in Europe in 2015, including difficult-to-treat meropenem non-susceptible isolates (of which 21.8% carried MBLs), MDR, and colistin-resistant isolates. The promising activity of aztreonam-avibactam against MBL-producing isolates that often carry resistance mechanisms to multiple drug classes warrants further development of this combination for use against these pathogens.

## Introduction

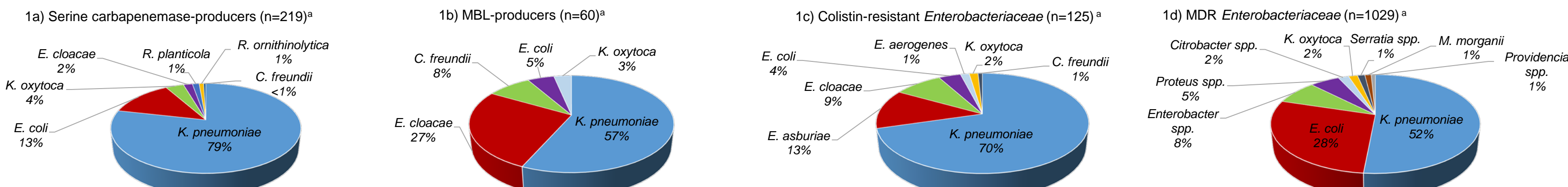
Aztreonam-avibactam is a  $\beta$ -lactam/ $\beta$ -lactamase inhibitor combination being developed for use against carbapenem-resistant *Enterobacteriaceae*, especially isolates producing metallo- $\beta$ -lactamases (MBL). Aztreonam is stable to hydrolysis by MBL but inactivated by many serine  $\beta$ -lactamases. Avibactam is active against serine  $\beta$ -lactamases, including extended-spectrum  $\beta$ -lactamases (ESBL), AmpC  $\beta$ -lactamases, and serine carbapenemases, often co-carried in isolates with MBLs. This study evaluated the *in vitro* activity of aztreonam-avibactam and comparators against *Enterobacteriaceae* collected in 2015 in Europe.

## Materials & Methods

- 6,449 non-duplicate clinical isolates of *Enterobacteriaceae* were collected from 67 medical centres in 17 European countries in 2015.
- Susceptibility testing was performed using CLSI broth microdilution [1] and interpreted using EUCAST 2017 breakpoints [2]. Aztreonam-avibactam was tested at a fixed concentration of 4 mg/L avibactam.
- Multidrug resistant (MDR) was defined as resistant by EUCAST breakpoints to sentinel agents from three or more drug classes (cephalosporins, monobactams,  $\beta$ -lactam/ $\beta$ -lactamase inhibitor combinations, carbapenems, fluoroquinolones, aminoglycosides, glycolycyclines, and polymyxins).
- PCR and sequencing were used to determine the  $\beta$ -lactamase genes (*bla*) present in isolates with meropenem MIC  $>1$  mg/L, ceftazidime MIC  $>8$  mg/L, and those phenotypically positive for ESBL activity.
- Isolates were screened for *bla* encoding MBLs (*bla*<sub>NDM</sub>, *bla*<sub>IMP</sub>, *bla*<sub>VIM</sub>, *bla*<sub>SPM</sub>, *bla*<sub>GIM</sub>), serine carbapenemases (*bla*<sub>KPC</sub>, *bla*<sub>OXA-48-like</sub>, *bla*<sub>GES</sub>), ESBLs (*bla*<sub>SHV</sub>, *bla*<sub>TEM</sub>, *bla*<sub>CTX-M</sub>, *bla*<sub>EB</sub>, *bla*<sub>PER</sub>, *bla*<sub>GES</sub>), and AmpC enzymes (*bla*<sub>ACC</sub>, *bla*<sub>ACT</sub>, *bla*<sub>CMY</sub>, *bla*<sub>DHA</sub>, *bla*<sub>FOX</sub>, *bla*<sub>MIR</sub>, *bla*<sub>MOX</sub>) as previously described [3].

## Results

Figure 1a-1d. Species distribution of serine carbapenemase-positive (KPC, OXA-48-like, GES), metallo- $\beta$ -lactamase-positive, colistin-resistant, and multidrug resistant (MDR) *Enterobacteriaceae* isolates.

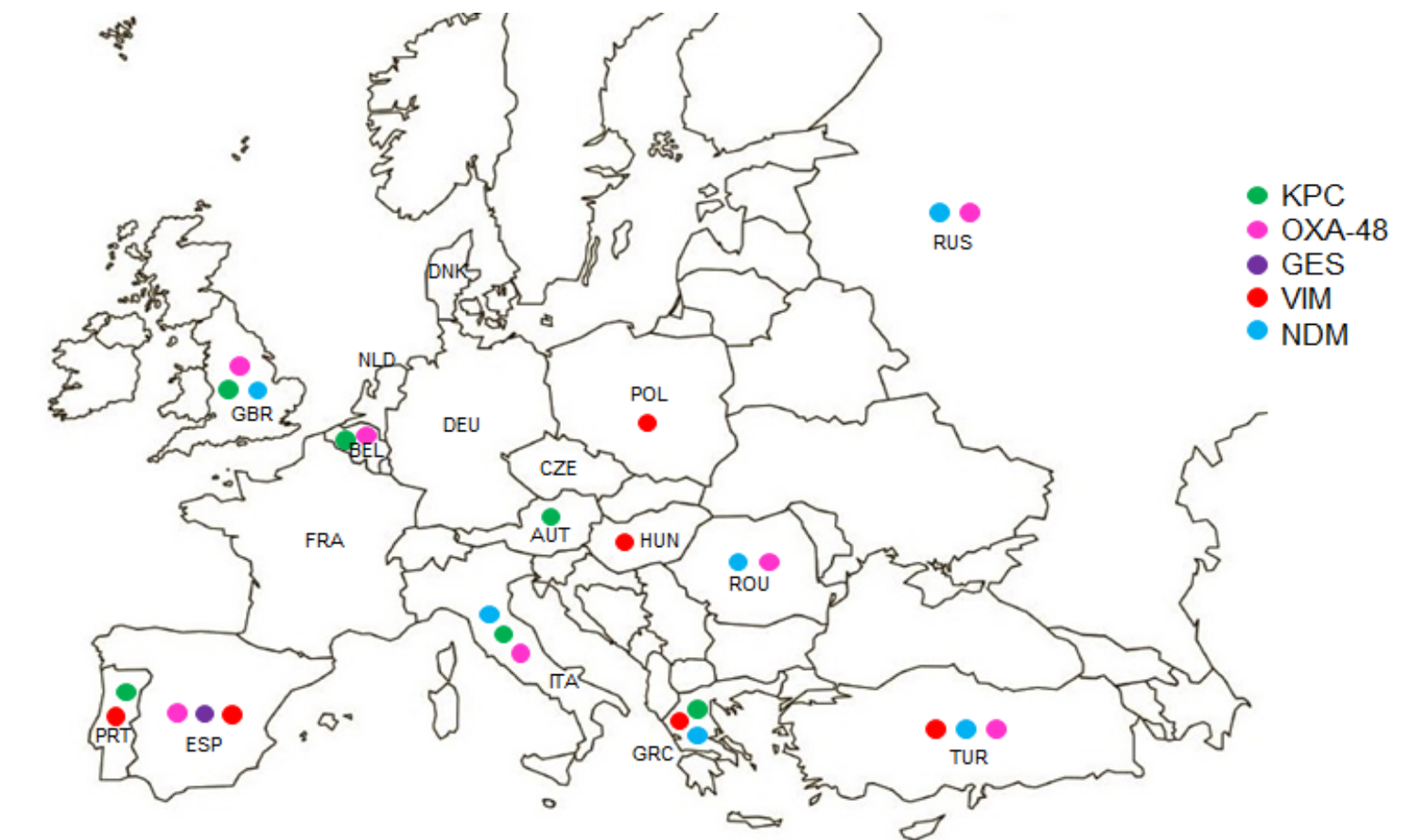


<sup>a</sup> Excludes two *K. pneumoniae* isolates carrying VIM-26 and KPC-2, five *C. freundii* carrying VIM-31 and OXA-48, and one *E. cloacae* and eleven *K. pneumoniae* carrying NDM-1 and OXA-48.  
<sup>b</sup>  $\beta$ -lactamase content of 60 isolates of *Enterobacteriaceae* (n): *C. freundii*, VIM-31, OXA-48 and TEM-OSBL (5); *E. cloacae*, VIM-1 and TEM-OSBL (7); VIM-1 (3); NDM-1 and CTX-M-15 (3); NDM-1 (1); NDM-1 and OXA-48 (1); NDM-1, CTX-M-15, SHV-12 and TEM-OSBL (1); *E. coli*, NDM-1 and TEM-OSBL (1); NDM-1, CTX-M-27 and CMY-2 (1); NDM-1, CTX-M-15, CTX-M-27, CMY-6, DHA-1 and TEM-OSBL (1); *K. oxytoca*, VIM-1 (1); VIM-1, CTX-M-3 and SHV-12 (1); *K. pneumoniae*, NDM-1, CTX-M-15 and SHV-OSBL (7); VIM-26 and SHV-5 (8); NDM-1, OXA-48, CTX-M-15 and SHV-ESBL (5); NDM-1, OXA-48 and CTX-M-15 (3); NDM-1, OXA-48 and SHV-OSBL (3); NDM-1, CTX-M-15, SHV-OSBL and TEM-OSBL (3); VIM-26, KPC-2, SHV-12 and TEM-OSBL (2); VIM-1 and SHV-OSBL (2); VIM-4, CTX-M-15, SHV-OSBL and TEM-OSBL (1); NDM-1, CTX-M-15, SHV-5, CMY-6, and TEM-OSBL (1); NDM-1, CMY-6 and SHV-OSBL (1).

<sup>a</sup> Excludes *Proteus* and *Serratia* spp. intrinsically resistant to colistin.

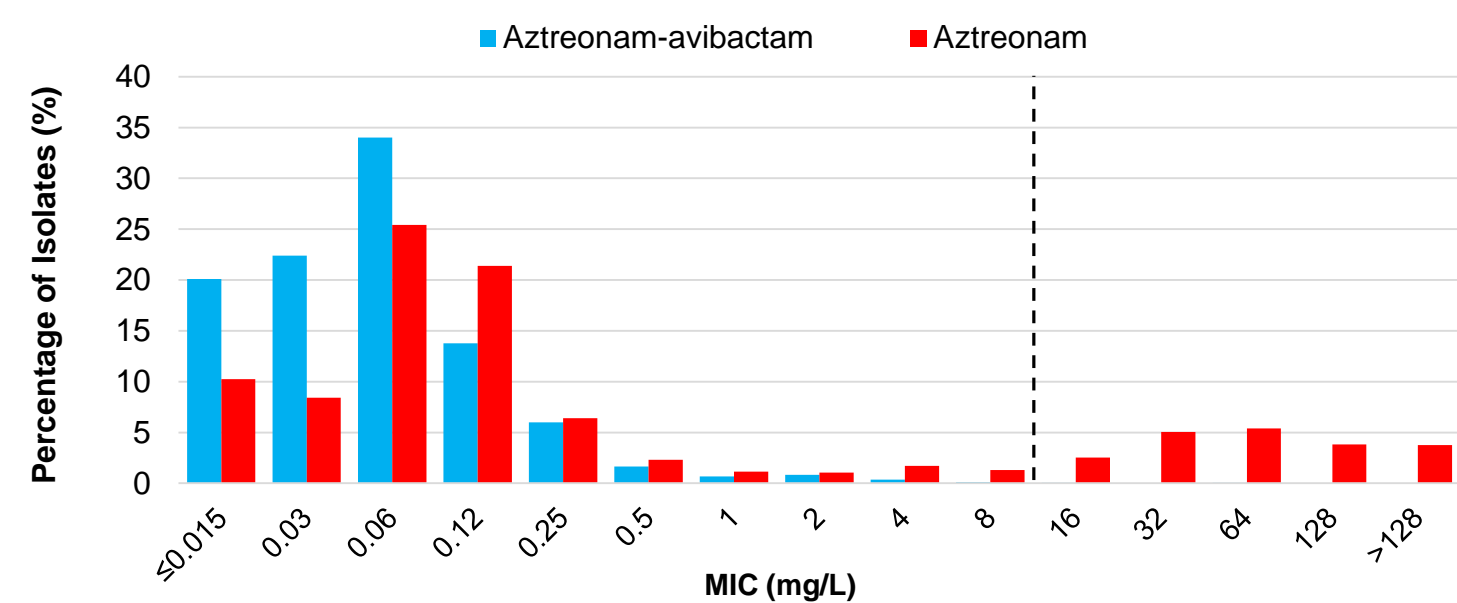
<sup>a</sup> *Citrobacter* spp. includes *Citrobacter freundii* (18), *Citrobacter braakii* (1), and *Citrobacter koseri* (1); *Enterobacter* spp. includes *Enterobacter cloacae* (61), *Enterobacter aerogenes* (15), *Enterobacter asburiae* (3), and *Enterobacter kobei* (2); *Proteus* spp. includes *Proteus mirabilis* (52) and *Proteus vulgaris* (4); *Providencia* spp. includes *Providencia stuartii* (7) and *Providencia rettgeri* (1); *Serratia* spp. includes *Serratia marcescens* (13) and *Serratia liquefaciens* (1).

Figure 2A. Country distribution of carbapenemases detected in *Enterobacteriaceae* clinical isolates collected in Europe in 2015.



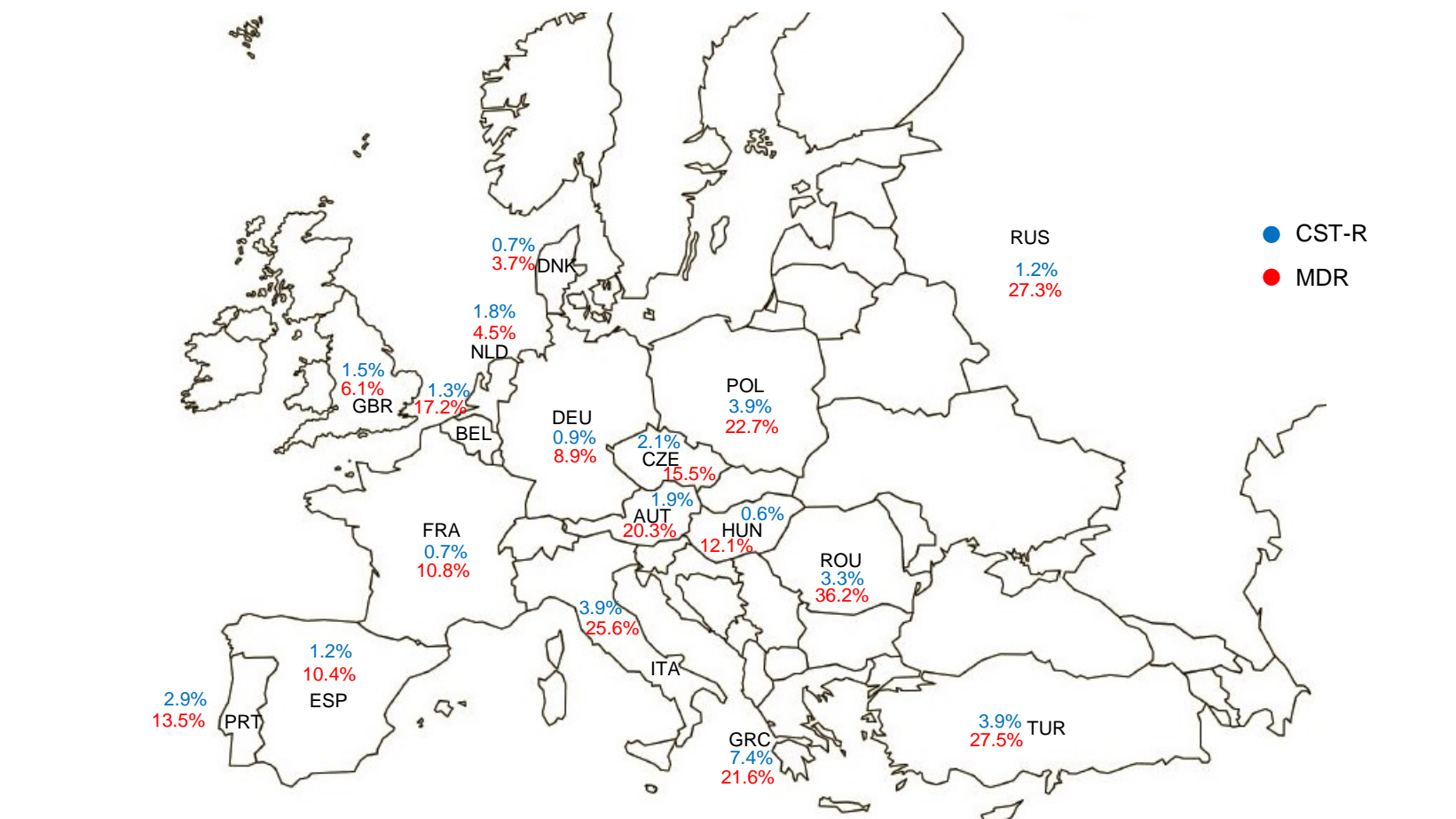
Participating countries: AUT, Austria; BEL, Belgium; CZE, Czech Republic; DNK, Denmark; FRA, France; DEU, Germany; GRC, Greece; HUN, Hungary; ITA, Italy; NLD, Netherlands; POL, Poland; PRT, Portugal; ROU, Romania; RUS, Russia; ESP, Spain; TUR, Turkey; GBR, United Kingdom.

Figure 3A. Aztreonam and aztreonam-avibactam MIC distributions against *Enterobacteriaceae* (n=6,449) collected in Europe.



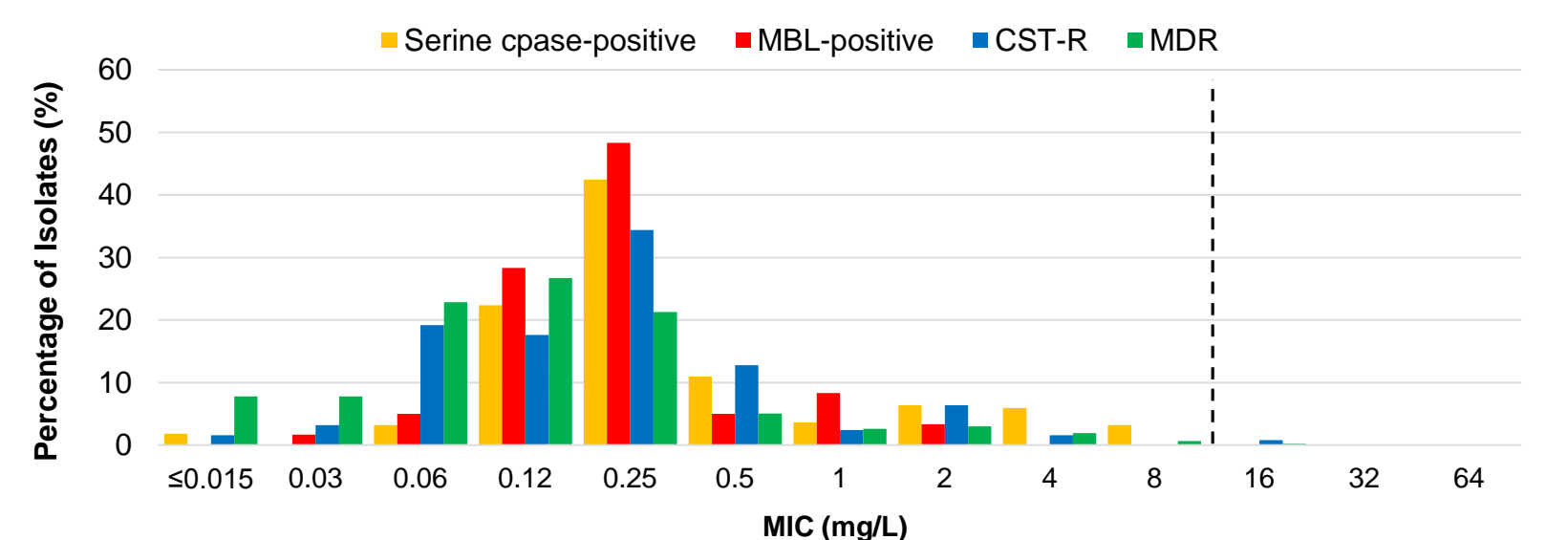
Dashed line represents the preliminary PK/PD cutoff of  $\leq 8$  mg/L for aztreonam-avibactam. Not visible due to low numbers and scale of graph: isolates with aztreonam-avibactam MIC values of 8 (0.11%), 16 (0.05%), and 64 mg/L (0.02%).

Figure 2B. Percentages of colistin-resistant (CST-R) and multidrug resistant (MDR) *Enterobacteriaceae* collected in 2015, by country.



Participating countries: AUT, Austria; BEL, Belgium; CZE, Czech Republic; DNK, Denmark; FRA, France; DEU, Germany; GRC, Greece; HUN, Hungary; ITA, Italy; NLD, Netherlands; POL, Poland; PRT, Portugal; ROU, Romania; RUS, Russia; ESP, Spain; TUR, Turkey; GBR, United Kingdom.

Figure 3B. Aztreonam-avibactam MIC distributions against serine carbapenemase-positive (Serine cpase-positive) (n=219), MBL-positive (n=60), colistin-resistant (CST-R) (n=125), and multidrug resistant (MDR) (n=1029) *Enterobacteriaceae* isolates collected in Europe.



Dashed line represents the preliminary PK/PD cutoff of  $\leq 8$  mg/L for aztreonam-avibactam. CST-R *Enterobacteriaceae* exclude isolates of *Proteus* and *Serratia* spp. intrinsically resistant to colistin. One CST-R *K. pneumoniae* isolate carrying SHV-12 and TEM-OSBL  $\beta$ -lactamases tested with an aztreonam-avibactam MIC of 16 mg/L. Not visible due to low numbers and scale of graph: MDR isolates with aztreonam-avibactam MIC values of 16 (0.19%) and 64 mg/L (0.10%).

Table 1. *In vitro* activity of aztreonam-avibactam and comparator agents tested against *Enterobacteriaceae* collected in Europe.

Group (n) <sup>a</sup> /Drug	MIC (mg/L)			% Susceptible <sup>b</sup>
	Range	MIC <sub>50</sub>	MIC <sub>90</sub>	
<i>Enterobacteriaceae</i> All (6,449)				
Aztreonam-avibactam	$\leq 0.015$ -64	0.06	0.12	NA <sup>c</sup>
Aztreonam	$\leq 0.015$ - $>128$	0.12	64	75.4
Cefepime	$\leq 0.12$ - $>16$	$\leq 0.12$	$>16$	79.1
Meropenem	$\leq 0.004$ - $>8$	0.06	0.12	96.2
Colistin	$\leq 0.06$ - $>8$	0.5	$>8$	83.0
Amikacin	$\leq 0.25$ - $>32$	2	8	93.4
Tigecycline	$\leq 0.015$ -8	0.5	2	88.3
Levofloxacin	$\leq 0.004$ - $>8$	0.06	$>8$	72.2
Aztreonam-NS (1,590)				
Aztreonam-avibactam	$\leq 0.015$ -64	0.12	0.5	NA
Aztreonam	2- $>128$	64	$>128$	0.0
Cefepime	$\leq 0.12$ - $>16$	$>16$	$>16$	19.9
Meropenem	0.008- $>8$	0.06	$>8$	85.6
Colistin	0.12- $>8$	0.5	4	89.8
Amikacin	$\leq 0.25$ - $>32$	4	32	79.5
Tigecycline	$\leq 0.015$ -8	0.5	2	87.8
Levofloxacin	0.008- $>8$	8	$>8$	30.4
Meropenem-NS (248)				
Aztreonam-avibactam	$\leq 0.015$ -16	0.25	1	NA
Aztreonam	0.06- $>128$	$>128$	$>128$	7.7
Cefepime	$\leq 0.12$ - $>16$	$>16$	$>16$	3.2
Meropenem	4- $>8$	$>8$	$>8$	0.0
Colistin	0.25- $>8$	1	$>8$	75.4
Amikacin	0.5- $>32$	32	$>32$	33.5
Tigecycline	0.12-8	1	2	76.2
Levofloxacin	0.06- $>8$	$>8$	$>8$	6.5
Colistin-R (125)				
Aztreonam-avibactam	$\leq 0.015$ -16	0.25	1	NA
Aztreonam	0.03- $>128$	128	$>128$	24.0
Cefepime	$\leq 0.12$ - $>16$	$>16$	$>16$	30.4
Meropenem	0.015- $>8$	1	$>8$	51.2
Colistin	4- $>8$	$>8$	$>8$	0.0
Amikacin	0.5- $>32$	8	$>32$	51.2
Tigecycline	0.12-8	1	2	84.0
Levofloxacin	0.015- $>8$	$>8$	$>8$	30.4
MDR (1,029)				
Aztreonam-avibactam	$\leq 0.015$ -64	0.12	0.5	NA
Aztreonam	$\leq 0.015$ - $>128$	64	$>128$	6.4
Cefepime	$\leq 0.12$ - $>16$	$>16$	$>16$	6.8
Meropenem	0.008- $>8$	0.06	$>8$	77.2
Colistin	0.12- $>8$	0.5	$>8$	82.1
Amikacin	0.5- $>32$	8	$>32$	68.5
Tigecycline	$\leq 0.015$ -8	0.5	4	80.0
Levofloxacin	0.03- $>8$	$>8$	$>8$	8.2
MBL-negative (6,389)				
Aztreonam-avibactam	$\leq 0.015$ -64	0.06	0.12	NA
Aztreonam	$\leq 0.015$ - $>128$	0.12	64	75.9
Cefepime	$\leq 0.12$ - $>16$	$\leq 0.12$	$>16$	79.8
Meropenem	$\leq 0.004$ - $>8$	0.06	0.12	97.0
Colistin	$\leq 0.06$ - $>8$	0.5	$>8$	82.9
Amikacin	$\leq 0.25$ - $>32$	2	8	94.0
Tigecycline	$\leq 0.015$ -8	0.5	2	88.4
Levofloxacin	$\leq 0.004$ - $>8$	0.06	$>8$	72.8
MBL-positive (60) <sup>d</sup>				
Aztreonam-avibactam	0.03-2	0.25	1	NA
Aztreonam	0.06- $>128$	128	$>128$	21.7
Cefepime	1- $>16$	$>16$	$>16$	3.3
Meropenem	0.25- $>8$	$>8$	$>8$	10.0
Colistin	0.25- $>8$	0.5	1	96.7
Amikacin	1- $>32$	32	$>32$	28.3
Tigecycline	0.12-4	0.5	2	70.0
Levofloxacin	0.03- $>8$	$>8$	$>8$	10.0

<sup>a</sup> NS, non-susceptible; R, resistant; MDR, multidrug resistant; MBL, metallo- $\beta$ -lactamase.  
<sup>b</sup> % Susceptibility was determined according to EUCAST 2017 interpretive criteria. Values  $\geq 90\%$  are indicated in bold.  
<sup>c</sup> NA, no breakpoints available.  
<sup>d</sup> Includes isolates carrying Class A (KPC, ESBL, OSBL) and/or Class C and/or Class D (OXA-48-like) enzymes.

## Results Summary

- 279 carbapenemase-producing isolates (206 *K. pneumoniae*, 32 *E. coli*, 20 *E. cloacae*, and 21 other isolates of *Enterobacteriaceae*) of a total of 6,449 tested were collected in 12 of 17 participating European countries in 2015, including isolates positive for KPC (n=111), OXA-48-like (n=107), VIM (n=21), NDM (n=20), GES (n=1), NDM and OXA-48-like (n=12), VIM and OXA-48-like (n=5), and VIM and KPC (n=2) enzymes (Figure 1 and Figure 2).
- Though no *Enterobacteriaceae* carrying IMP were found, other carbapenemases were widely disseminated, with OXA-48-positive isolates found in 7 countries and KPC-positive, VIM-positive, and NDM-positive isolates each found in 6 countries. Multiple carbapenemase types were found in 9 European countries (Figure 2).
- Aztreonam-avibactam showed good activity against all *Enterobacteriaceae* (MIC<sub>90</sub>, 0.12 mg/L) collected in Europe.  $>99.9\%$  (6,445 of 6,449) of isolates tested with aztreonam-avibactam MIC values  $\leq 8$  mg/L, including isolates carrying MBLs and/or serine  $\beta$ -lactamases, aztreonam-NS, meropenem-NS, colistin-R, and MDR isolates (MIC<sub>90</sub>s, 0.5–1 mg/L) (Figure 3 and Table 1).
- Among meropenem-NS, colistin-R, MDR, and MBL-positive isolates, susceptibilities to other tested agents were reduced (cephalosporins, 3.2–30.4% susceptible; meropenem, 0–77.2% susceptible; amikacin, 28.3–68.5% susceptible; tigecycline, 70.0–84.0% susceptible). Colistin was active against MBL-positive isolates (MIC<sub>90</sub>, 1 mg/L; 96.7% susceptible) but showed decreased activity against other difficult-to-treat subsets of isolates (0–82.1% susceptible) (Table 1).

## Conclusions

- Aztreonam-avibactam was highly potent *in vitro* against *Enterobacteriaceae* collected in Europe in 2015, including difficult-to-treat meropenem non-susceptible isolates (of which 21.8% carried MBLs), MDR, and colistin-resistant isolates.
- The promising activity of aztreonam-avibactam against MBL-producing isolates that often carry resistance mechanisms to multiple drug classes warrants further development of this combination for use against these pathogens.

### References and Acknowledgments:

- Clinical Laboratory Standards Institute. 2015. Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically: approved standards -- Tenth Edition. CLSI document M07-910. Wayne, PA.
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