

# Association between vancomycin MIC with virulence genes expression and clonal complexes of methicillin-susceptible *Staphylococcus aureus* (MSSA) strains isolated from left-sided endocarditis

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**Background.** The impact of a high vancomycin (VAN) minimum inhibitory concentration (MIC) phenotype (HVM; >1.5 mg/L) in methicillin-susceptible *Staphylococcus aureus* (MSSA) bacteraemia and infective endocarditis (IE) is poorly known. MSSA IE isolates are known to possess a distinct repertoire of virulence genes and clonal complexes (CC) that differentiates them from other type of MSSA isolates [1].

Several studies reported higher rates of complications and mortality in patients with MSSA bacteraemia caused by strains with HVM [2,3], as well as a correlation with *agr* dysfunction, *agr* type II polymorphism, and other specific findings shaping a repertoire of virulence factors in the HVM group [4-7]. Our group demonstrated the association between HVM and significantly higher mortality in a cohort of 93 patients with MSSA IE treated with cloxacillin [8], demonstrating higher mortality rates and systemic emboli in the HVM group. Mortality of MSSA IE was three-fold higher among patients with high VAN MIC isolates. Nonetheless, some recent studies did not find significant differences either on *agr* subgroup and function in MSSA bacteremia and IE [9,10] nor on left-MSSA IE treated with beta-lactams' clinical outcomes [10] according to vancomycin MIC.

The **aim of this study** was to investigate whether the strains belonging to the cohort of patients from the Cervera et al study [8] presented specific patterns of virulence factors, CC types or ability to form biofilms in the presence or not of vancomycin.

**Methods.** The cohort included 53 isolates with HVM and 40 with HVM collected from 2000 to 2006. Isolates underwent spa typing to infer CC, and multiplex polymerase chain reaction for the presence of virulence genes as described in [1]. Biofilm formation was determined following the methodology presented in [11]. The association between CC, virulence factors and VAN phenotype, as well as in-hospital mortality and symptomatic systemic emboli, were analyzed.

**Results.** We found no differences in adhesins [fibronectin-binding proteins (*fnbA*, *fnbB*), clumping factors (*clfA*, *clfB*), collagen-binding gene (*cna*), serine-aspartate repeat proteins for adhesion (*sdrC*, *sdrD*, *sdrE*), sialoprotein (*bbp*) and elastin-binding protein (*ebps*), and MHC class II analog proteins (MAP/EAP)], toxins [exfoliative toxins (*eta*, *etb*), enterotoxins (*tst*), staphylococcal enterotoxins (*sea*, *seb*, *sec*, *sed*, *see*, *seg*, *seh*, *sei*, *sej*), Panton-Valentin leucocidin (PVL), and hemolysin (*hlg*)] or other putative virulence [fibrinogen-binding protein (*efb*), adhesion intracellular protein A (*icaA*), chemotaxis-inhibiting protein (*chp*), and serine endopeptidase (*V8*)] genes between MSSA isolates according to VAN MIC (Table 1).

*Agr* subgroups I and III predominated, with no association with VAN MIC. Isolates with lower VAN MICs exhibited **higher ability to form biofilm with and without the presence of VAN** ( $p < 0.001$  and  $p = 0.022$ , respectively). **There was no association of CC type with VAN MIC** (CC30, CC34; CC45 = 50% of isolates).

**Table 1. Genotypic Characteristics, Clonal Complexes and Biofilm Productin of MSSA Isolates from Patients With IE According to Vancomycin (VAN) MIC**

	Vancomycin MIC < 1.5 µg/mL N=53	Vancomycin MIC ≥ 1.5 µg/mL N=40	P
<b>Adhesins:</b>			
▪ <i>fnbA</i>	53 (100%)	40 (100%)	-
▪ <i>fnbB</i>	41 (77%)	37 (93%)	0.049
<b>Toxins:</b>			
▪ <i>eta</i>	4 (8%)	7 (18%)	0.197
▪ <i>sei</i>	34 (64%)	26 (65%)	0.932
▪ <i>sej</i>	20 (38%)	13 (33%)	0.601
▪ <i>pvl</i>	0	1 (2.5%)	0.430
▪ <i>hlg</i>	41 (77%)	28 (70%)	0.422
<b>Other putative virulence genes:</b>			
▪ <i>efb</i>	35 (66%)	29 (73%)	0.505
▪ <i>icaA</i>	53 (100%)	40 (100%)	-
▪ <i>chp</i>	27 (51%)	20 (50%)	0.928
▪ <i>V8</i>	10 (19%)	8 (20%)	0.891
<b>Agr subgroup:</b>			
▪ I	17 (32%)	15 (38%)	0.586
▪ II	12 (23%)	9 (23%)	0.987
▪ III	20 (38%)	12 (30%)	0.437
▪ IV	2 (4%)	0	0.504
<b>Clonal complex:</b>			
▪ 5	3 (6%)	6 (15%)	0.166
▪ 30	12 (23%)	9 (23%)	0.987
▪ 34	7 (13%)	3 (8%)	0.507
▪ 45	8 (15%)	7 (18%)	0.755
▪ Other	23 (43%)	15 (38%)	0.567
<b>Biofilm formation</b>			
▪ without VAN (SD)	2,871 (0.685)	2,602 (0.417)	0.022
▪ with VAN (SD)	2,482 (0.509)	2,034 (0.421)	<0.001
<b>tPMP:</b>			
▪ 50 mcg/mL (SD)	64,442 (18,595)	72,764 (23,918)	0.062
▪ 25 mcg/mL (SD)	80,183 (21,271)	87,298 (20,607)	0.109
▪ 12.5 mcg/mL (SD)	76,665 (18,593)	80,845 (18,861)	0.289

Neither CC, biofilm formation nor virulence factors were identified as risk factors for **in-hospital mortality**, except for *efb*, which was associated with lower mortality. As for clinically evident **systemic emboli**, *sei*, *hlg*, *efb*, *V8*, and a lower ability for biofilm formation in the presence of VAN were associated with significantly higher rates of emboli (Table 2). On **multivariate analysis**, the presence of the *hlg* gene was associated with almost 20-fold higher risk of emboli (OR 19.5, 95%CI 1.78-212.51;  $P = 0.015$ ), whereas the ability to form biofilm in presence of VAN was associated with reduced risk of emboli (OR 0.21, 95%CI 0.06-0.80;  $P = 0.022$ ).

**Table 2. Univariate Analysis: In-hospital Mortality; Symptomatic Embolic Events**

	Survivors (n=56)	In-hospital mortality (n=37)	P	No embolic events (n=74)	Embolic events (n=19)	P
<b>Adhesins:</b>						
▪ <i>fnbA</i>	56 (100%)	37 (100%)	-	74 (100%)	19 (100%)	-
▪ <i>fnbB</i>	46 (82%)	32 (87%)	0.577	63 (85%)	15 (79%)	0.499
<b>Toxins:</b>						
▪ <i>sei</i>	38 (68%)	22 (59%)	0.407	44 (60%)	16 (84%)	0.044
▪ <i>sej</i>	16 (29%)	17 (46%)	0.087	27 (37%)	6 (32%)	0.792
▪ <i>pvl</i>	0	1 (3%)	0.398	0	1 (5%)	0.204
▪ <i>hlg</i>	42 (75%)	27 (73%)	0.827	50 (68%)	19 (100%)	0.002
<b>Other putative virulence genes:</b>						
▪ <i>efb</i>	43 (77%)	21 (57%)	0.041	47 (64%)	17 (90%)	0.029
▪ <i>icaA</i>	56 (100%)	37 (100%)	-	74 (100%)	19 (100%)	-
▪ <i>chp</i>	29 (52%)	18 (49%)	0.767	35 (47%)	12 (63%)	0.217
▪ <i>V8</i>	10 (18%)	8 (22%)	0.653	11 (15%)	7 (37%)	0.048
<b>Agr subgroup:</b>						
▪ I	20 (36%)	12 (32%)	0.744	25 (34%)	7 (37%)	0.802
▪ II	11 (20%)	10 (27%)	0.405	16 (22%)	5 (26%)	0.759
▪ III	22 (39%)	10 (27%)	0.223	26 (35%)	6 (32%)	0.771
▪ IV	2 (4%)	0	0.516	1 (1%)	1 (5%)	0.369
<b>Clonal complex:</b>						
▪ 5	4 (7%)	5 (14%)	0.316	6 (8%)	3 (16%)	0.467
▪ 30	12 (21%)	9 (24%)		19 (26%)	2 (11%)	
▪ 34	9 (16%)	1 (3%)		7 (10%)	3 (16%)	
▪ 45	9 (16%)	6 (16%)		13 (18%)	2 (11%)	
▪ Other	22 (39%)	16 (43%)		29 (39%)	9 (47%)	
<b>Biofilm formation</b>						
▪ without VAN (SD)	2.75 (0.66)	2.76 (0.49)	0.915	2.75 (0.58)	2.79 (0.66)	0.811
▪ with VAN (SD)	2.30 (0.50)	2.27 (0.55)	0.820	2.35 (0.53)	2.06 (0.41)	0.028
<b>tPMP:</b>						
▪ 50 mcg/mL (SD)	69.34 (21.7)	66.03 (20.9)	0.467	67.3 (21.1)	70.8 (22.3)	0.549
▪ 25 mcg/mL (SD)	84.59 (23.1)	81.20 (17.9)	0.453	82.5 (21.6)	86.1 (19.7)	0.508
▪ 12.5 mcg/mL (SD)	78.92 (18.6)	77.77 (19.1)	0.772	78.4 (19.0)	78.8 (18.2)	0.930

## Conclusions

- MSSA with **higher** vancomycin MICs had **lower ability to form biofilms**, which was associated with **higher embolic rates**.
- No association was found between adhesins, toxins, *agr* or other virulence gene expression and CC according to vancomycin MIC or **in-hospital mortality**, except for *efb*, which was associated with lower mortality.
- Hemolysin (*hlg*)** and **reduced biofilm formation** in the presence of vancomycin were associated with **systemic emboli**.

**References:** [1] Nienaber JJ et al, J Infect Dis 2011;204:704e13; [2] Holmes NE et al, J Infect Dis 2011;204:340e7; [3] Aguado JM et al, Emerg Infect Dis 2011;17:1099e102; [4] Viedma E et al, J Antimicrob Chemother 2014;69:51e8; [5] Bae IG et al, J Infect Dis 2009;200:1355e66; [6] Fowler VG et al, J Infect Dis 2007;196:738e47; [7] Holmes NE et al, J Clin Microbiol 2014;52:3384e93; [8] Cervera C et al, Clin Infect Dis 2014;58:1668e75; [9] López-Cortés LE et al, J Antimicrob Chemother 2015;70:2652e60; [10] Pericàs JM et al, Clin Microbiol Infect 2017 (ahead of print); [11] Abdelhady W et al, Antimicrobial Agents Chemother 2013; 57:1447-54.