

# ANTIMICROBIAL AND ANTI-BIOFILM ACTIVITIES OF BIOLOGICALLY SYNTHESIZED SELENIUM NANOPARTICLES

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## BACKGROUND

Tailored nanoparticles (NPs) with desired physico-chemical properties have been proposed as a new line in the battle against antibiotic-resistant microorganisms.

In this study, SeNPs generated by two bacterial isolates of environmental origin have been tested and compared with chemically synthesized NPs for their antibacterial, antifungal and cytotoxic activity. Antimicrobial and anti-biofilm characteristics of SeNPs were tested toward clinical strains of *Pseudomonas aeruginosa* and clinical isolates of *Candida* species. The antifungal activity was tested against clinical isolates of *Aspergillus spp.*

## METHODS

1. The gram-positive strain *Bacillus mycooides* SeITE01 and the gram-negative strain *Stenotrophomonas maltophilia* SeITE02 were used to produce biogenic SeNPs [(Bm-SeNPs(+)) and Sm-SeNPs(-)] respectively. Chemically synthesized SeNPs (Ch-SeNPs) were produced using a standard protocol.

2. *Pseudomonas* and *Candida* strains were treated for 48 hours at 37° C with different concentrations of Bm-SeNPs(+), Sm-SeNPs(-) and Ch-SeNPs to test their inhibition activity on biofilm formation. The biofilm degradation activity was evaluated by analyzing the quantity of the biofilm remaining after treatment of the synthesized exopolymer with different concentrations of biogenic and chemically synthesized SeNPs. The biofilm was quantified after methylene blue staining.

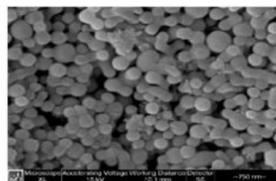


Figure 1. Sm-SeNPs (-).

3. The zone of inhibition (ZOI) technique was used to observe the activity of SeNPs on fungal species. The effect on mycelia growth was evaluated after filtering suspensions of *A. fumigatus* and *A. flavus* inoculated for 8 days with 64µg/ml (MIC) of Sm-SeNPs(-).

4. Evaluation of the Se-NPs cytotoxic activity and their effect on cell viability was determined by ELISA method, AlamarBlue® assay and cytochrome C reduction in human dendritic cells and fibroblast.

## RESULTS

Table 1. Inhibition of biofilm synthesis by Se-NPs. In bold, percentages of biofilm formation inhibition higher than 80%. (\*) Quantity of biofilm in arbitrary unit.

Bacterial/ Yeast strain (*)	<i>S. maltophilia</i> - SeNP (µg/ml)				<i>Bacillus mycooides</i> - Se NPs (µg/ml)				Chemically synthesized Se NPs (µg/ml)			
	50	100	250	500	50	100	250	500	50	100	250	500
<i>P. aeruginosa</i> PAO1 (1.10)	40±2.5	45±3	70±2.5	<b>96±1</b>	33±3	47±6	63±4.5	<b>95±1.2</b>	9±0.7	21±2.1	<b>94±0.7</b>	<b>96±1.4</b>
<i>P. aeruginosa</i> Pyo 27853 (1.00)	15±0.7	30±0.7	41±0.7	66±2.1	15±4.2	17±2	44±2.8	64±1	4±1.4	10±0.7	35±2.1	44±2
<i>P. aeruginosa</i> INT (1.00)	23±1	34±1	59±3.5	<b>95±1</b>	25±4.5	29±3.5	49±2.5	<b>94±1.5</b>	2±3.5	2±1.4	20±0.7	30±0.7
<i>P. aeruginosa</i> CFC21 (1.20)	66±5	<b>86±0.5</b>	<b>95±2</b>	<b>96±1.5</b>	37±5.5	66±3.5	<b>93±1.7</b>	<b>95±2</b>	10±3.5	33±3.5	71±1.4	65±1.4
<i>P. aeruginosa</i> CFC20 (1.10)	75±0.5	<b>82±1</b>	<b>86±1</b>	<b>93±1.5</b>	72±2	76±0.5	<b>85±1</b>	<b>91±2</b>	53±0.7	<b>90±0.7</b>	<b>95±1</b>	<b>95±1</b>
<i>P. aeruginosa</i> CFCA (2.75)	31±1	39±3	39±2	<b>88±4</b>	28±1.5	34±5	<b>81±4.5</b>	<b>85±5</b>	1±0.5	1±0.5	<b>97±0.7</b>	<b>98±1</b>
<i>P. aeruginosa</i> CFCB (3.00)	39±1	<b>94±1</b>	<b>94±3.5</b>	<b>96±1</b>	25±5	25±1.5	<b>94±0.5</b>	<b>96±1</b>	5±0.7	6±0.7	<b>96±1</b>	<b>97±1.5</b>
<i>C. albicans</i> (0.9)	61±0.5	60±3	60±1	<b>94±1</b>	60±6.5	69±2	74±2.5	<b>93±0.5</b>	0	0	0	9±0.7
<i>C. parapsilosis</i> (0.85)	72±1.5	79±0.5	73±1	<b>95±1</b>	75±1.5	73±5	72±3	<b>94±0.5</b>	0	0	0	5±0.7

Table 2. Percentages of biofilm disgregation caused by Se-NPs. In bold, percentages of disaggregation higher than 80% of the biofilm initial quantity. (\*) Quantity of biofilm in arbitrary unit.

Bacterial/ Yeast strain (*)	<i>S. maltophilia</i> - SeNP (µg/ml)				<i>Bacillus mycooides</i> - Se NPs (µg/ml)				Chemically synthesized Se NPs (µg/ml)			
	50	100	250	500	50	100	250	500	50	100	250	500
<i>P. aeruginosa</i> PAO1 (1.10)	73±5	72±2	76±2	73±5	52±1	62±4.5	57±1	73±6	16±0.7	18±4.2	37±0.7	53±1
<i>P. aeruginosa</i> Pyo 27853 (1.00)	49±1.4	53±1.4	53±1.4	43±0.7	31±4.9	43±1.4	51±7	45±7	23±1.4	35±1	40±2.5	46±0.7
<i>P. aeruginosa</i> INT (1.00)	63±5.5	53±0.5	61±1.5	41±4.5	65±4.5	44±0.5	58.2±3	32±1.5	15±6.3	15±6.3	8±2.8	8±4.5
<i>P. aeruginosa</i> CFC21 (1.20)	21±1	45±3.5	53±5.5	63±2	44±4.5	16±3	33±1.5	61±0.5	8±1.4	5±0.7	16±1.4	1±0.7
<i>P. aeruginosa</i> CFC20 (1.10)	<b>87±3</b>	<b>84±0.5</b>	<b>85±1</b>	<b>85±0.5</b>	<b>80±1</b>	<b>86±1</b>	<b>86±1.7</b>	<b>82±2</b>	17±3.5	13±2.8	29±4.2	50±2.8
<i>P. aeruginosa</i> CFCA (2.75)	53±4	58±3	56±3	64±2	25±1	27±5	51±2.5	47±3	0	4±1.4	57±1.4	72±0.7
<i>P. aeruginosa</i> CFCB (3.00)	44±2.5	44±2.5	39±1	53±2	28±4	20±2.5	40±2.5	53±1	0	2±0.7	66±0.7	73±0.7
<i>P. aeruginosa</i> Pyo 27853	49±1.4	53±1.4	53±1.4	43±0.7	31±4.9	43±1.4	51±7	45±7	23±1.4	35±1	40±2.5	46±0.7
<i>C. albicans</i> (0.9)	26±2.5	43±2.5	47±3.5	60±2	11±2.5	32±2	48±1.5	60±3.5	0	0	2	0
<i>C. parapsilosis</i> (0.85)	52±2	48±1.5	48±2.5	64±2	48±3	38±2	47±2	42±2.5	0	1	0	0

Table 3. MIC values for biogenic and chemically synthesized Se-NPs on different microbial strains.

Bacterial / Yeast strain	MIC (µg/ml)		
	<i>S. maltophilia</i> - SeNP	<i>Bacillus mycooides</i> - Se NPs	Chemically synthesized Se NPs
<i>P. aeruginosa</i> PAO1	128	128	128
<i>P. aeruginosa</i> Pyo ATCC27853	512	512	>512
<i>P. aeruginosa</i> INT	256	512	>512
<i>P. aeruginosa</i> FCF21	16	64	128
<i>P. aeruginosa</i> FCF20	8	32	128
<i>P. aeruginosa</i> CFCA	16	32	>128
<i>P. aeruginosa</i> CFCB	16	64	>128
<i>C. Albicans</i>	256	512	>512
<i>C. parapsilosis</i>	512	512	>512

Table 4. Mycelia quantity (in mg) after treatment with MIC doses of Sm- NPs (-) and Itraconazole compared with control.

<i>Aspergillus</i> strain	Mycelia growth (mg)		
	Control	Itraconazole 4µg/ml	<i>S. maltophilia</i> - SeNP 64µg/ml
<i>A. flavus</i>	25±0.1	8±0.7	5±1
<i>A. fumigatus</i> (A)	10±0.2	2±0.1	1±0.7
<i>A. fumigatus</i> (B)	10±0.1	2±0.7	2±0.7

## CONCLUSIONS

➤ Biogenic SeNPs are able to inhibit biofilm synthesis induced by both *Pseudomonas* and *Candida* as well as to disaggregate the mature exopolysaccharide matrix produced by these microorganisms.

➤ Some *P. aeruginosa* clinical strains (CFC20, CFC21 and CFCB) are particularly sensitive to the biogenic SeNPs: the lowest concentrations tested (50 and 100 µg/ml) caused a 70-90% inhibition of the biofilm synthesis (Table 1).

➤ The percentage of biofilm degradation (Table 2) upon treatment with 50-100 µg/ml NPs was about 50-70%, and did not increase with higher NP concentrations.

➤ As concerns the *Candida* strains, the lowest SeNP dose used (50 µg/ml) was capable of causing an inhibition of the biofilm formation or the disaggregation of the biofilm of about 50-60%.

➤ SeNPs produced by *S. maltophilia* show an antimicrobial and antifungal activity higher than that observed with *B. mycooides* SeNPs and chemically synthesized NPs.

➤ MIC values of Se-NPs varied widely with the different *Pseudomonas* clinical strains analysed: MIC values resulting from the treatments with Bm-SeNPs were significantly higher and those obtained with Ch-SeNPs indicated that these NPs had no effect on *Pseudomonas* growth (Table 3).

➤ SeNPs produced by *S. maltophilia* are able to interfere with the *Aspergillus* mycelia growth, causing a partial inhibition of mycelia growth (Table 4).

➤ Both biogenic and chemically synthesized SeNPs do not show cytotoxic activity as do not affect the viability of human dendritic cells and fibroblasts and do not elicit the substantial secretion of pro-inflammatory and immune-stimulatory cytokines (data not showed).

## REFERENCES

- ThR. Y. Pelgrift, A. J. Friedman AJ, Nanotechnology as a therapeutic tool to combat microbial resistance, *Adv. Drug. Deliv. Rev.* 2013, 65, 1803-1815.
- S. Lampis, E. Zonaro, C. Bertolini, P. Bernardi, C. S. Butler, G. Vallini, Delayed formation of zero-valent selenium nanoparticles by *Bacillus mycooides* SeITE01 as a consequence of selenite reduction under aerobic conditions, *Microb. Cell. Fact.*, 2014, DOI: 10.1186/1475-2859-13-35.
- Z. H. Lin, C. R. C. Wang, Evidence on the size-dependent absorption spectral evolution of selenium nanoparticles, *Mat. Chem. Phys.*, 2005, 92(2-3), 591-94.
- M. Shakibaie, H. Forootanfar, Y. Golkari, T. Mohammadi-Khorsand, M. R. Shakibaie, Anti biofilm activity of biogenic selenium nanoparticles and selenium dioxide against clinical isolates of *Staphylococcus aureus*, *Pseudomonas aeruginosa*, and *Proteus mirabilis*, *J. Trace Elem. Med. Biol.*, 2015, 29, 235-241.

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