

Future MDx ESGMD course | Paul Savelkoul | January 22th, 2016

### Future developments in MDx

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### First microbiologist

1632-1723



Antoni van Leeuwenhoek

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### Start discipline of medical microbiology

1822-1895      1843-1910



Louis Pasteur

Origin of many of the currently laboratory techniques

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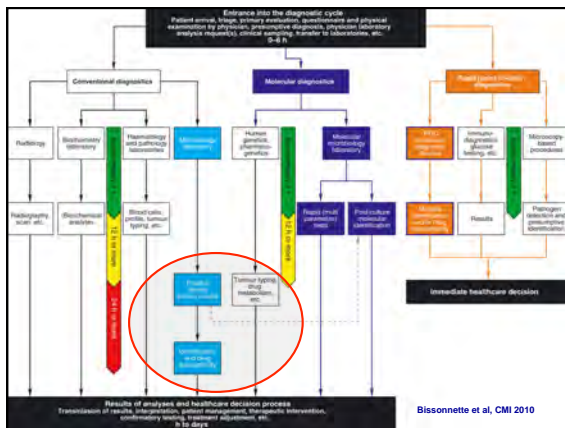
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### Current situation in the medical microbiological laboratory

**Classical technics:**

- culturing
- blood culture
- serological testing
- identification & AST
- staining

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### Current changing situation in the routine microbiological laboratory

**Molecular technics:**

- DNA isolators
- Real Time PCR
- MaldiToF
- Sanger sequencer (larger/university labs)

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**Consequences in the routine microbiological laboratory**

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- Viruses: virusculture replaced by RTPCR
- Parasites: more and more RTPCR
- Fungi: Increasing molecular testing
- Bacteriology: MaldiToF & RTPCR & BRPCR
- Resistance: More molecular detection

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**What is the most common situation on the sample level?**

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**Hospital:**

High throughput: Large sample size, planned, sample driven (e.g. hpv, CT, NG, HIV)

Middle throughput: Complex, speed, combined tests, patient driven (e.g. all clinical diagnostics)

Low throughput: Urgence, rapid result, patient driven (e.g. ICU patient)

**General practitioners :**


Most samples outsourcing to laboratories

Insourcing rapid easy to go POC tests

**Testing at home:**

Patient at home: self sampling and/or self testing

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**The new era in Medical microbiology**

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Crossing points with regard to spread of resistance, new molecular technologies, bioinformatics



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### A new era in medical microbiology

**MDx: enabling analysis of total human microbiome**

**Determining health & disease**

- Diagnostics of chronic infectious diseases
- Prevention of exacerbations
- Personalized diagnosis & therapy
- Monitoring of therapy
- New (preventive) treatment strategies

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### The new paradigm of medical microbiology

**One disease : One pathogen?**

**From pathogen detection toward microbiome detection:**

**Specific -> Syndrom -> Systemic**

**one bacterium -> known set -> microbiome**

**And beyond.....**

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### New approaches

- From sample driven towards patient driven analysis**
- From one test towards monitoring infection**
- From single pathogens towards communities**
- From pathogen detection towards commensal detection**
- From testing towards testing based patient specific treatment**
- From acute infections towards chronic inflammatory diseases**

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### MDx future lines

- New rapid RTPCR applications
- POC systems
- Massspectrometry systems (Maldi/Esi tof) (metabolites & DNA)
- Broad Range DNA amplification (gutflora, biofilm) (NGS, bacterial profiling)
- Next generation DNA sequencing (species & strain typing/resistome)
- Electronic nose (VOC)



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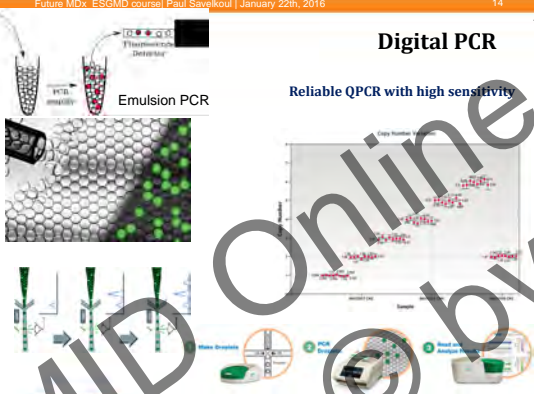
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### Digital PCR

Emulsion PCR

Reliable QPCR with high sensitivity



The diagram illustrates the digital PCR process. On the left, it shows 'Emulsion PCR' where a sample is mixed with PCR reagents in a microfluidic device to create individual droplets. On the right, a graph shows 'Copy Number' on the y-axis and 'Fluorescence Intensity' on the x-axis, with a peak indicating a positive result. Below the graph, a schematic shows the detection of PCR products and the resulting fluorescence signal.

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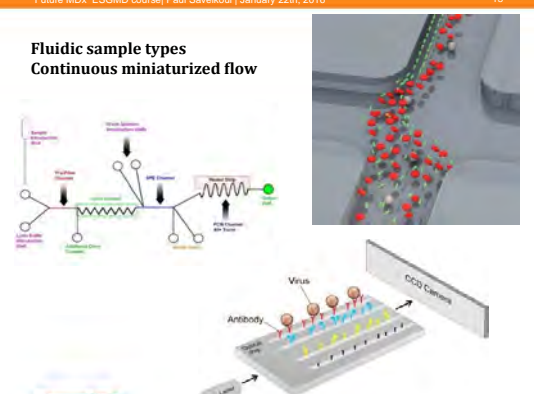
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### Fluidic sample types

Continuous miniaturized flow



The schematic shows a continuous miniaturized flow system. A sample containing 'Virus' and 'Antibody' is introduced into a microfluidic channel. The system includes a 'Virus' reservoir, a 'Virus' reservoir, a 'Virus' reservoir, and a 'Virus' reservoir. The flow is controlled by a 'Virus' reservoir and a 'Virus' reservoir. The final detection is performed using a 'CCD Camera'.

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### Miniaturisation

#### IC-CHIP POWERED NUCLEIC ACID MD

MDx technology based on electrical readout lab on a chip cartridge

FRIZ Biochem's Cycle® technology

Figure 2  
The cartridge for automated analysis is equipped with lyophilised chemicals for lysis and PCR. The MDx test kit further comprises a way to pick the sample, e.g. a swab of BIRSA PoC (VD)

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### Blood cultures

Direct detection PCR, Maldi of

Dag 0 1 2 3 4 5 6

Positive bloodculture

Negative bloodculture

Gram stain, Maldi of identification, PCR

Identification & AST

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### Direct detection of sepsis

5-10 ml fresh blood

standard lab equipment

POC system:  
Diagnosis of sepsis from 3- 6 days to 2 hrs

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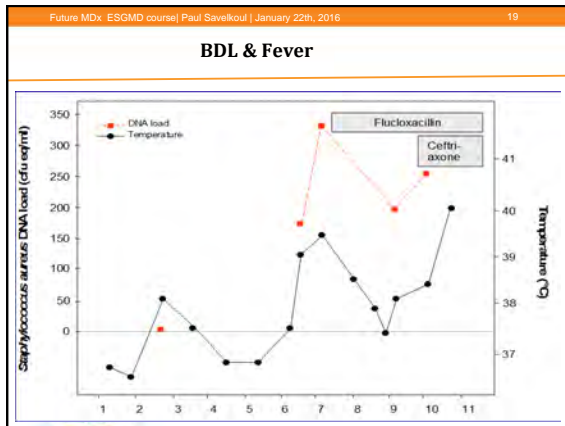
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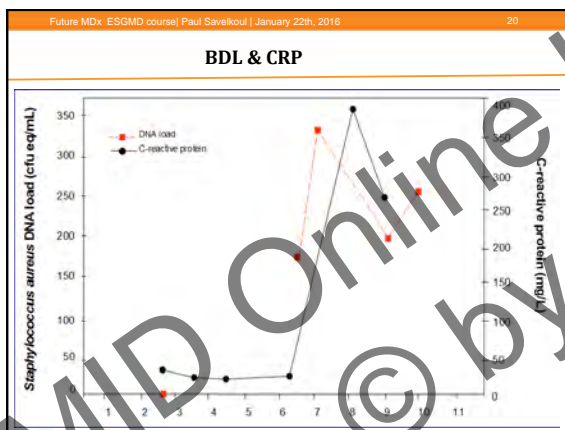
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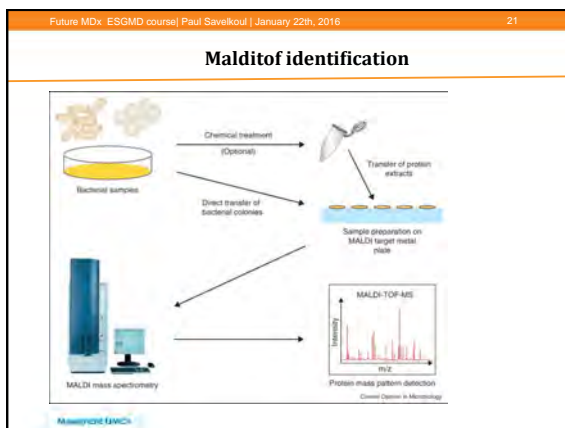
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### Next gen seq applications in the field of medical microbiology



14,870,316	bacterial genomes
10,627	viral genomes
8,641	plasmids
767,967	fungi
84,984	archaea

**Single cell sequencing**  
Identification and determination  
Detection of resistance genes  
Identification of virulence genes  
Discriminating epidemical & non-epidemical strains  
Strain specific characteristics (pathotyping)  
Transmission routes of mobile DNA elements

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### Resistome

Micro-organisms may carry resistance genes

Resistome = the collection of antibiotic resistance genes in both pathogenic and non-pathogenic bacteria

- Why focus on the resistome?
- Microbial communities – potential reservoirs of exchangeable resistance genes



Wright, G.D. Nature Reviews Microbiology 2007;5:175-186

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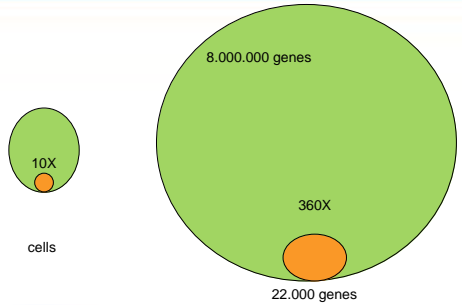
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8.000.000 genes

10X

cells

22.000 genes

360X

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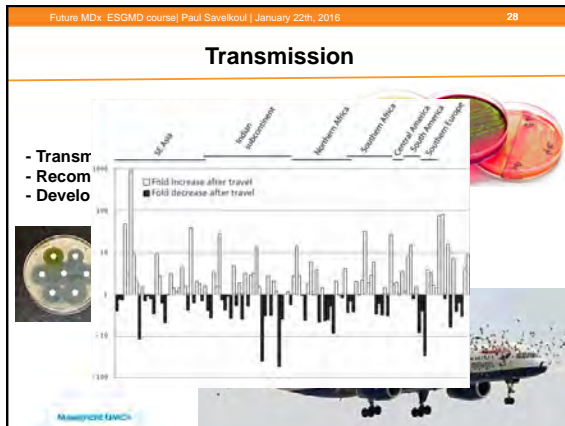
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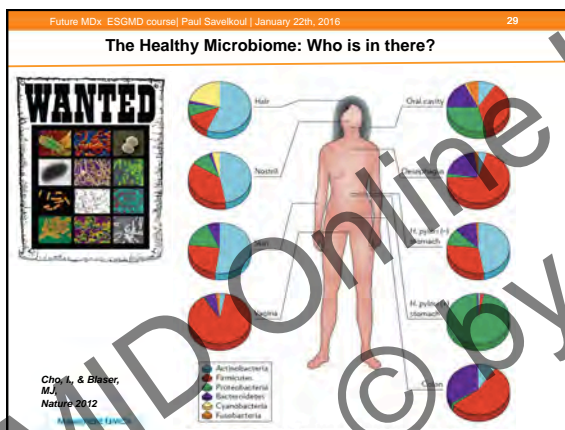
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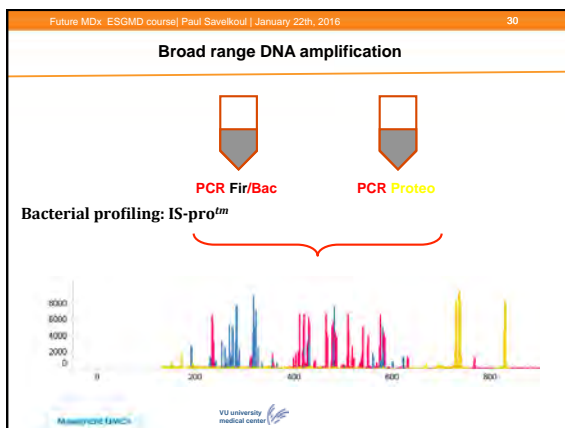
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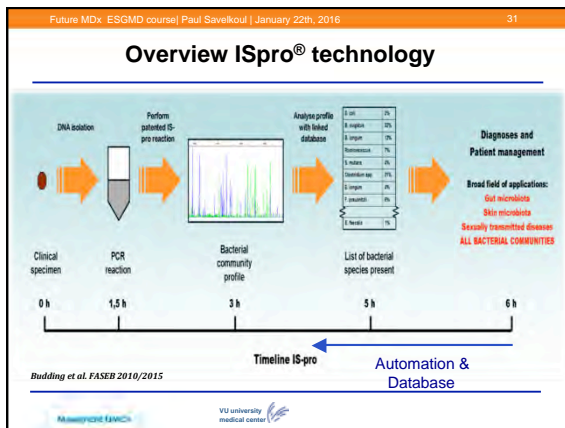
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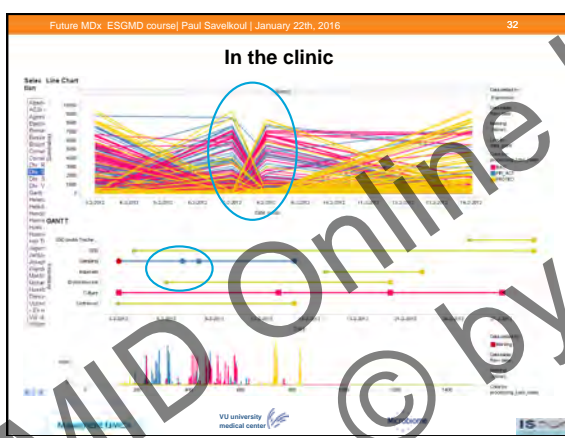
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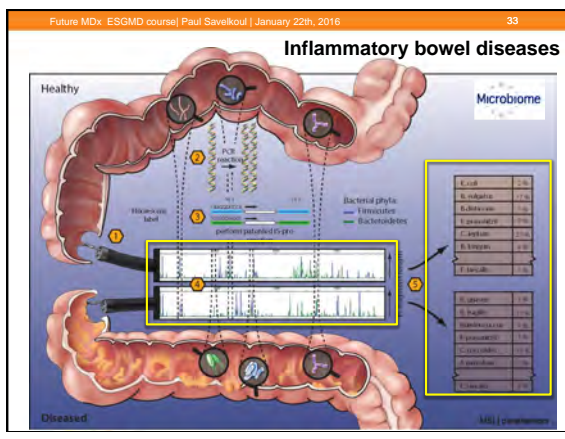
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### Medical applications of VOCs

VOCs in medicine

	Prevention	Diag	Progn
Neoplastic	Exposure assessment	Screening	Therapy response
Infectious	Pathogen identification	Host response	Monitoring
Inflammatory	Exposure assessment	Stratification and therapy titration	Monitoring for exacerbations

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### Pneumonia - Host response assessment

VOCs in medicine

Bean et al. ERJ, 2015

OWLSTONE MEDICAL

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### NEC - Monitoring for disease onset

VOCs in medicine

Table III. Performance characteristics of fecal VOC analysis for the discrimination of NEC, sepsis, and controls

Time window	AUC ± 95% CI	P value	Sensitivity	Specificity	LR+	LR-
<b>NEC vs controls</b>						
T <sub>0-1</sub>	0.98 ± 0.04	>201	88.9	88.9	8.1	0.1
T <sub>1-2</sub>	0.77 ± 0.25	0.04	62.5	75.0	2.3	0.2
T <sub>2-3</sub>	0.69 ± 0.25	0.27	60.0	60.0	1.5	0.7
<b>NEC vs sepsis</b>						
T <sub>0-1</sub>	0.84 ± 0.18	0.03	88.9	88.9	3.1	0.25
T <sub>1-2</sub>	0.80 ± 0.17	0.04	83.3	75.0	3.3	0.2
T <sub>2-3</sub>	0.82 ± 0.23	0.08	80.0	80.0	2.8	0.2

LR+, positive likelihood ratio; LR-, negative likelihood ratio. Sensitivity, specificity, and positive and negative likelihood ratios are reported for the optimum cut point.

de Meij et al. J Ped, 2015

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### Crohn's Disease - Disease activity?

VOCs in medicine

Table 2: Performance characteristics for the discrimination of ulcerative colitis, Crohn's disease and healthy controls by faecal VOC analysis.

	AUC ± 95% CI	p-Value	Sensitivity	Specificity	+LR	-LR
<b>Discriminating ulcerative colitis from healthy controls</b>						
Active disease	1.00 ± 0.00	< 0.001	100	100	N.A.	N.A.
Remission	0.94 ± 0.00	< 0.001	94.4	94.4	16.88	0.06
<b>Discriminating Crohn's disease from healthy controls</b>						
Active disease	0.89 ± 0.05	< 0.001	86.3	86.7	2.99	0.21
Remission	0.94 ± 0.06	< 0.001	93.8	91.4	16.75	0.07
<b>Discriminating ulcerative colitis from Crohn's disease</b>						
Active disease	0.96 ± 0.03	< 0.001	96.4	93.0	12.08	0.04
Remission	0.81 ± 0.08	0.002	87.8	77.2	16.86	0.06

de Meij, J Crohn Colitis, 2014

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### Current challenges in VOC research

VOCs in medicine

- Selecting the right population
- Lack of standardisation
- Potential confounders
- Aspecificity of some markers
- Reliability of sensor platforms
- Selectivity of analytical approach
- Lack of biological validation

Intention to diagnose population  
Lack of reproduction and data sharing  
Usability versus scientific rigor  
Carefully select clinical application  
Intra- / Inter-device variability, drift  
Balancing selectivity (local, systemic, exo) with portability  
More insights into biomarker origins needed

Agnes Boots, Trends in Molecular Medicine, 2015

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### Direct detection from clinical samples

Detection of resistance  
Detection of virulence factors  
Typing of strains

```
graph TD
    Saliva --> DNA_Ext[DNA extraction  
15 min, 2 h]
    Saliva --> Culture[Culture  
6-18 h]
    Culture --> PCR[PCR for MLST  
1 h]
    Culture --> MALDI[MALDI-TOF MS  
identification  
20 min, 24 h]
    Culture --> Phage[Phage typing  
6-18 h]
    PCR --> PLEX[PLEX-Microarray  
16-24 h]
    PCR --> NGS[NGS-MBT  
4-8 h]
    MALDI --> Protein[Protein  
expression  
20 min, 24 h]
    Phage --> Antigen[Antigen test  
5-18 h]
```

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### MDx: Automation

- Need for speed
- Need for connective databases
- Need for accessibility
- Need for specialised molecular microbiologists
- Need for molecular epidemiologists
- Need for bioinformaticians

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### The future routes of MDx

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### The coming years....

- More rapid MDX systems (shift from culture -> MDx)
- Fully automation (culture + MDx)
- Focus from pathogen detection towards microbiome detection
- Focus from infection towards prevention (typing & transmission)
- New treatment strategies to prevent disease and spread of resistance
- From disease towards personalised health (& systemic medicine)

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