

# Pre Clinical In Vitro Models

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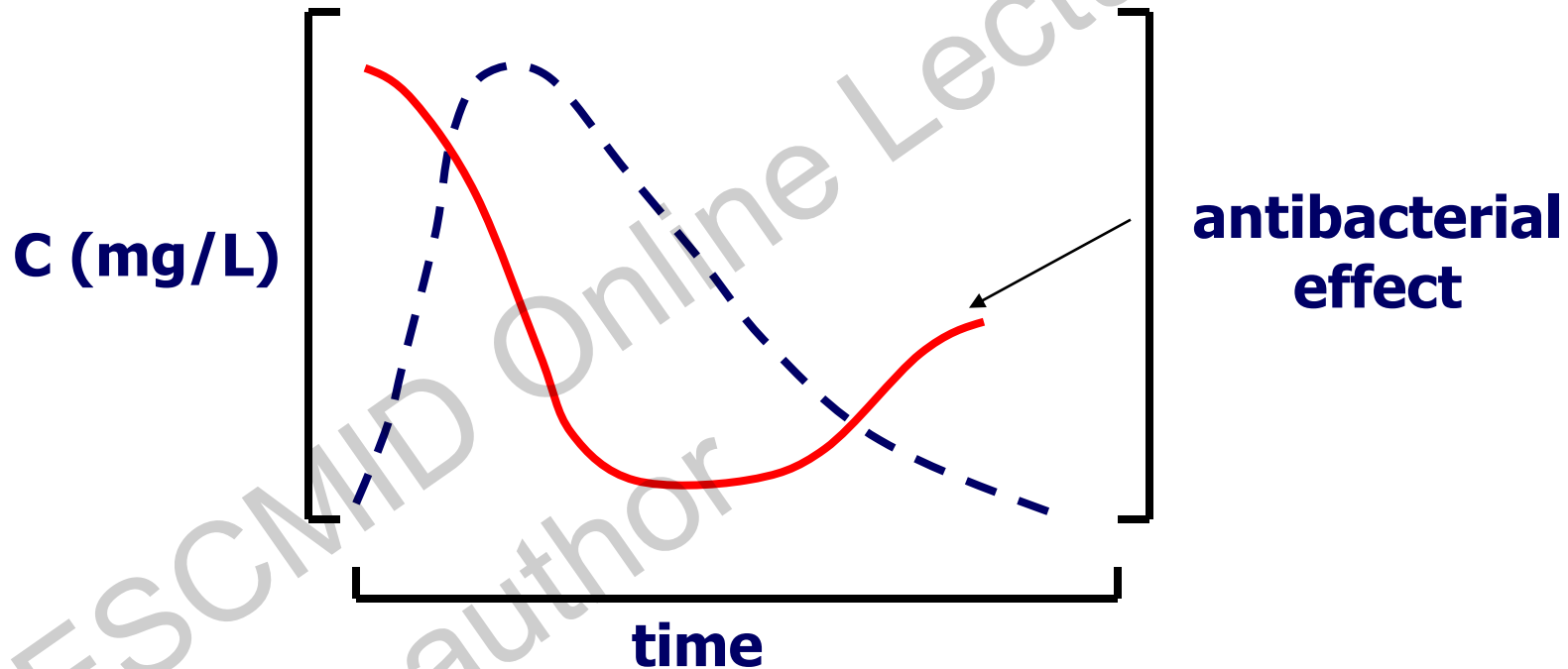
Bristol, UK

# In vitro models

- What are they
- What they can't do
- What are the pk/pd outputs
- Resistance
- Combinations
- In vivo correlates

# In vitro pK/pD modelling

study of changing drug concentration and antibacterial effect (ABE)



# What can in vitro models do?

Use pre-clinical models to understand drug exposure in relation to

- species
- inoculum
- exposure; pattern & duration
- impact of combination therapy
- pre-existing resistance within drug class
- risk of emergence of resistance

# Advantages v animals

- Easier to simulate human pk profile -free/total fractions
- Multiple sampling
- Extended duration of therapy
- High inoculum
  
- Strains difficult to grow in animals –Str.pneumoniae, H.influenzae
- Simulate diseases not established in animals eg TB, anthrax, Acinetobacter, anaerobes
- Combination therapies

# What in vitro models can't do

- Simulate immune response
- Tissue concentrations - protein binding

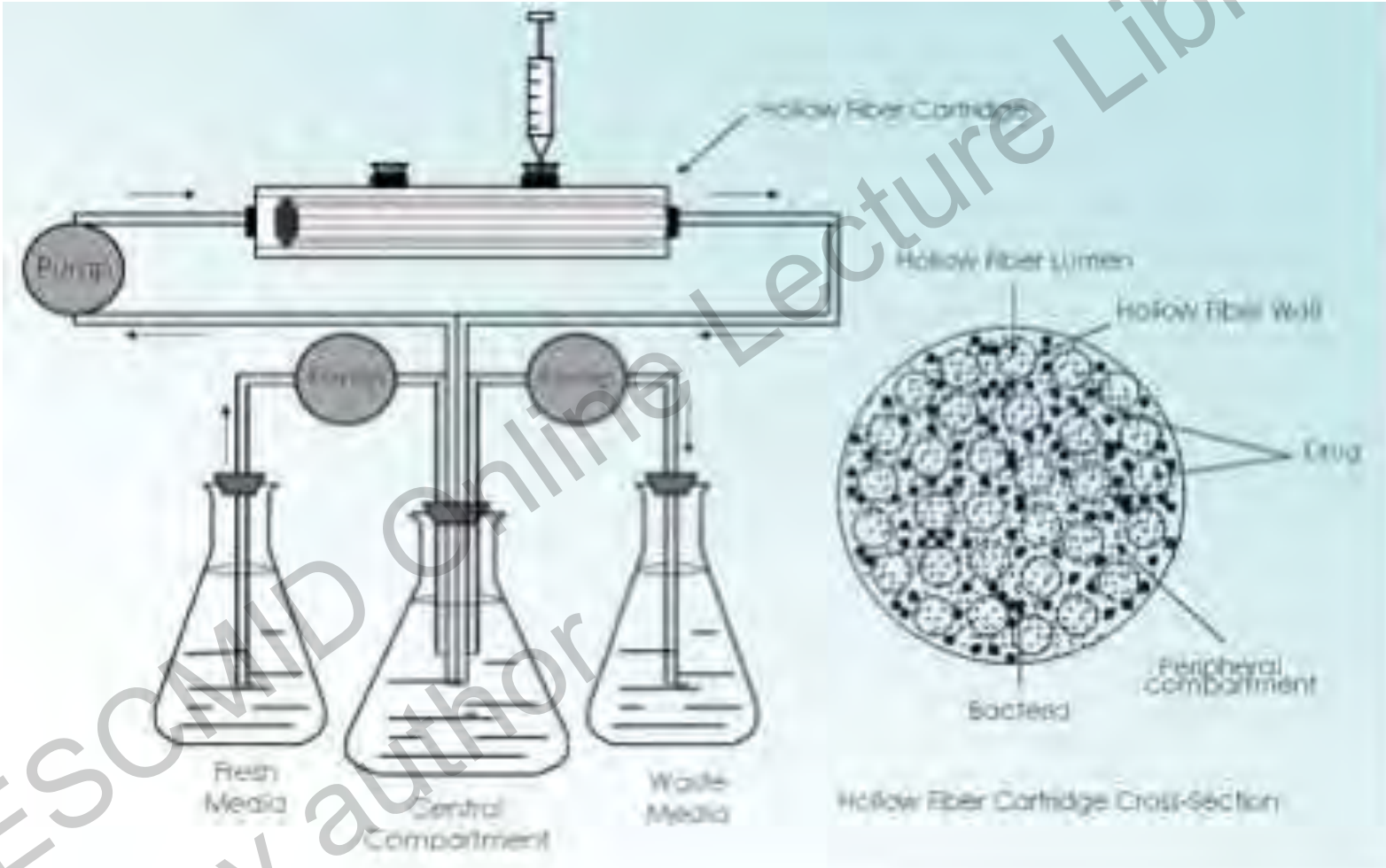
## Technical issues

- Adherence
- Contamination
- Dialysis models - blocked filters
- Labour intensive

# Types of IVMs

- Dilutional - with and without filter membrane (glass/plastic)
- Dialysis - hollow fibre
- Foreign body - Simulated Endocarditis Vegetations SEVs, Hanging Clot (fibrin)
- Intra-cellular
- Biofilms e.g. - mimic short and long term infection by *S.pneumoniae*/*P.aeruginosa* & pathophysiology of infection

# Types of in vitro models - Hollow fibre





# In vitro pK model - dilutional

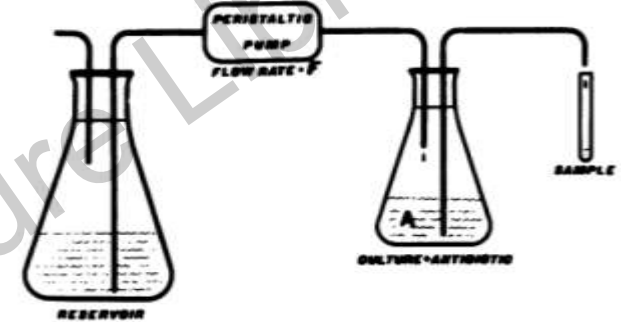


FIG. 1. Model I apparatus for simulation of monoexponential decrease in antibiotic concentration.

$$C = C_0 \times e^{-kt}$$

$C_0$  = conc in the vessel at Time 0

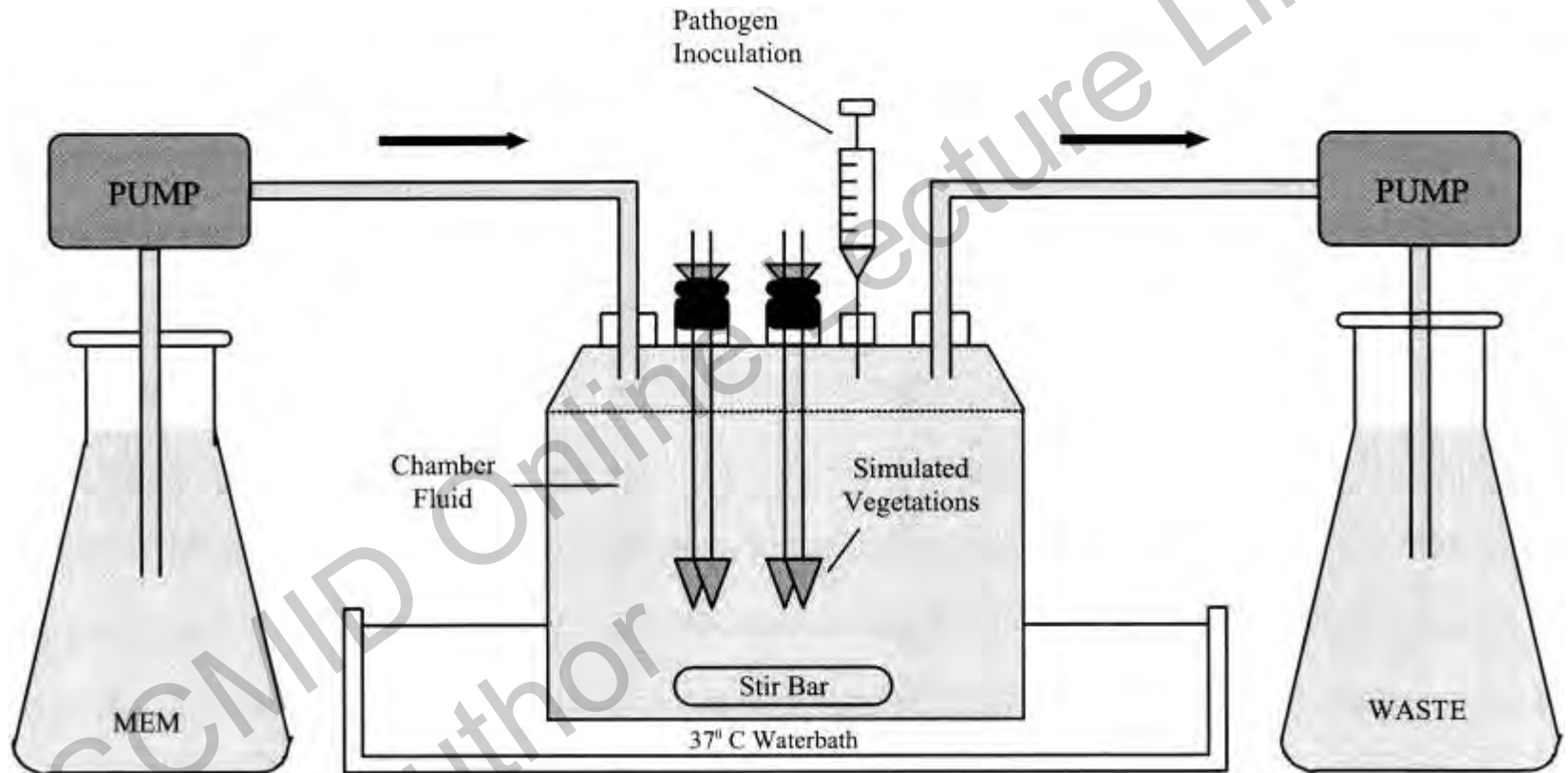
$$K = F/V$$

$$F = V \times 0.693/t_{1/2}$$

Grasso et al 1978

Bacteria are diluted at same rate of antibiotic – growth rate!

In vitro model of IE. PUMP, peristaltic pumps used to introduce fresh or remove exhausted medium.

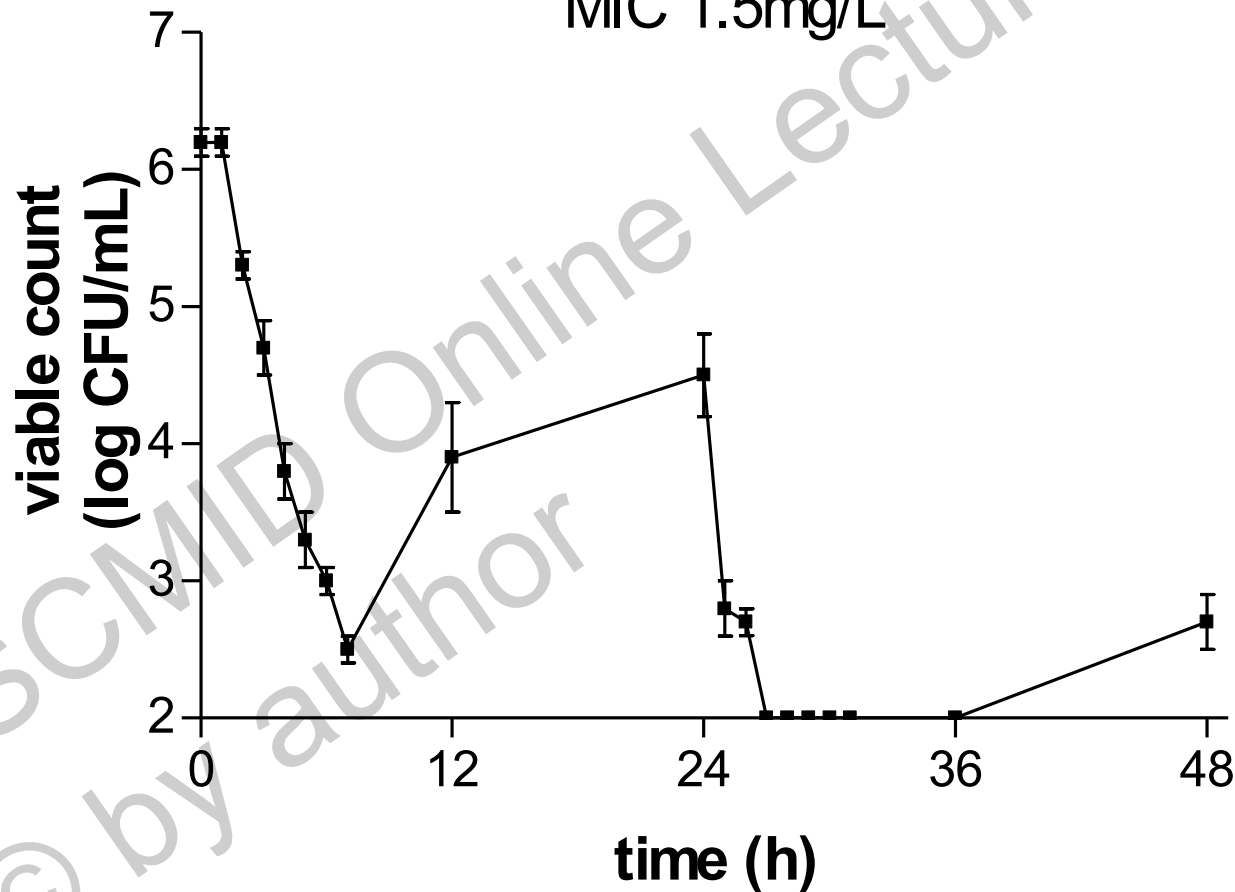


Mercier R et al. Antimicrob. Agents Chemother.  
2004;48:2551-2557

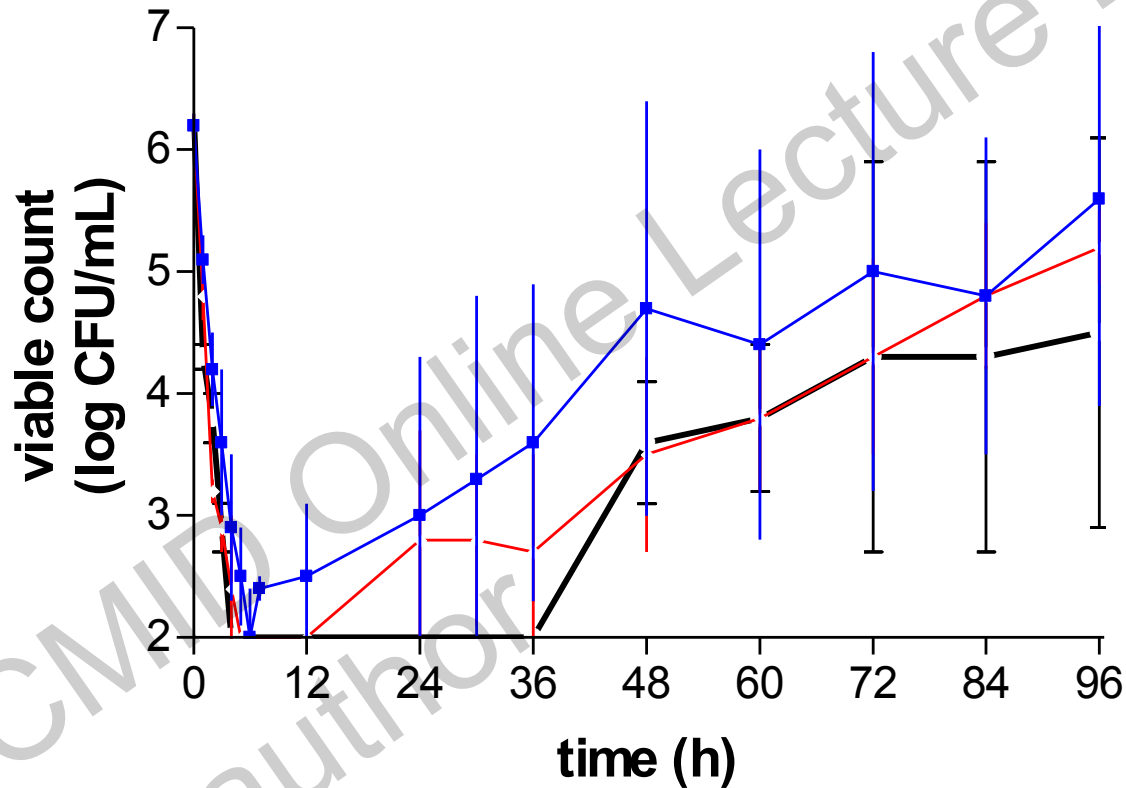
Antimicrobial Agents and Chemotherapy

# Descriptive studies - Ceftaroline

Ceftaroline 600mg 12hrly vs MRSA strain 33815  
MIC 1.5mg/L



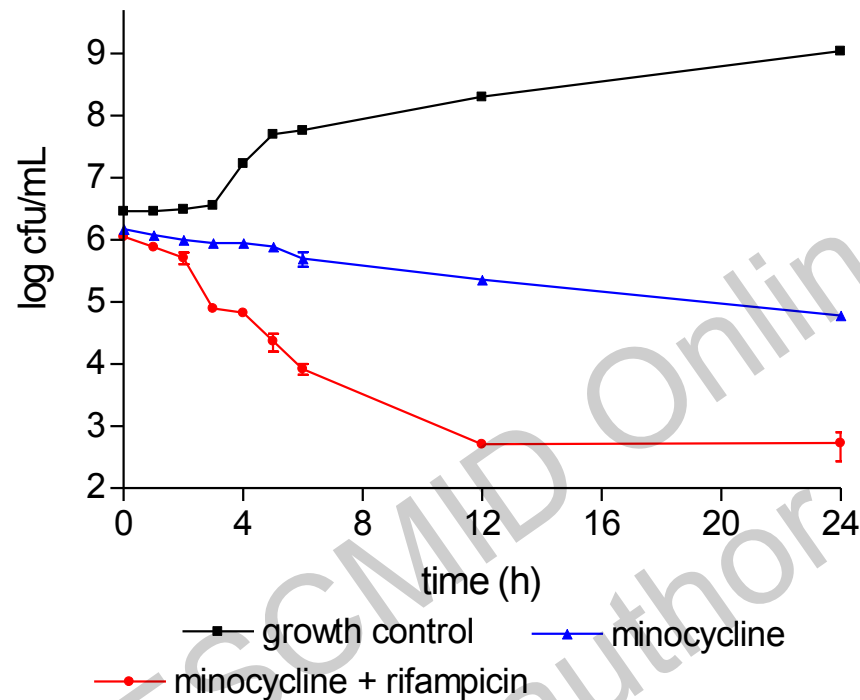
# Descriptive studies - Doripenem



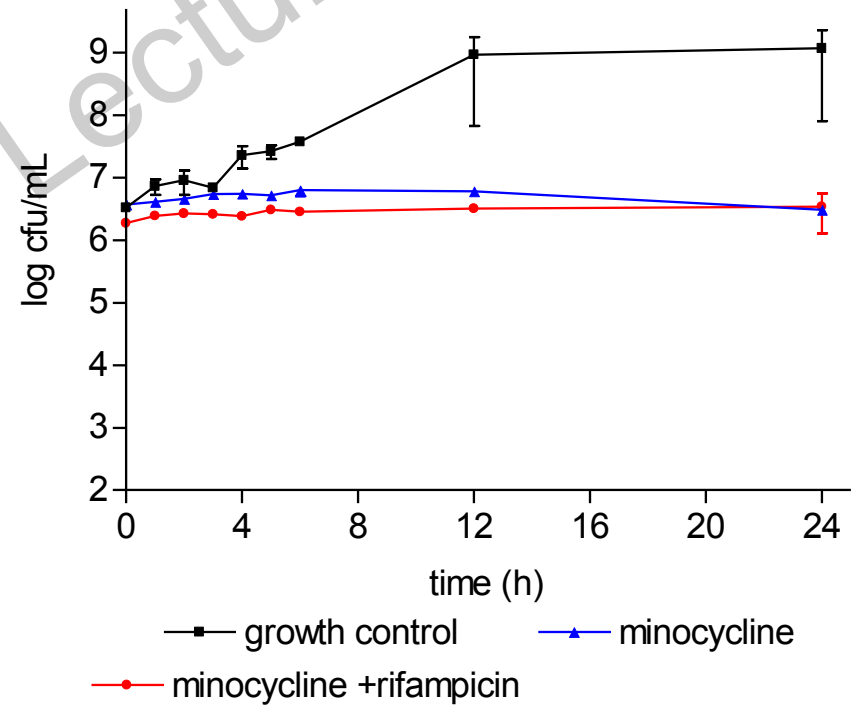
- doripenem 500mg 8hrly
- doripenem 1000mg 8hrly
- doripenem 2000mg 8hrly

# Descriptive studies - treatment failure

c) J1 pre treatment

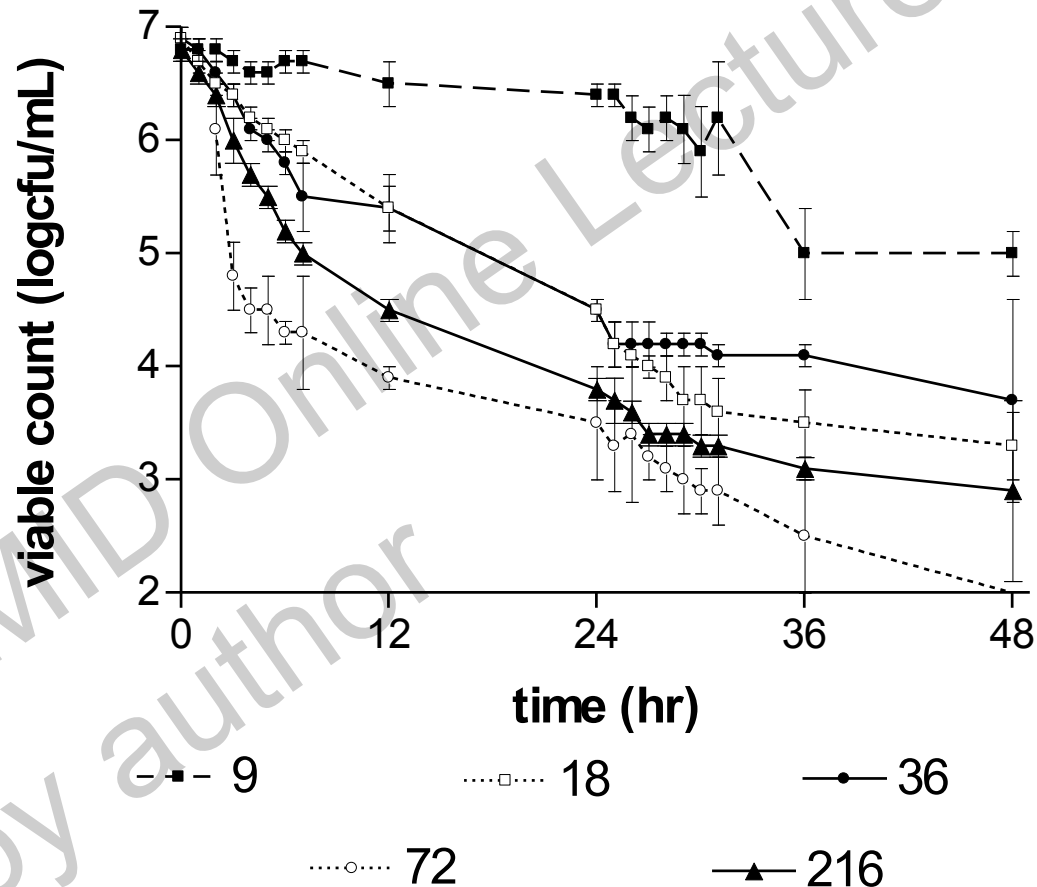


d) J2 post treatment



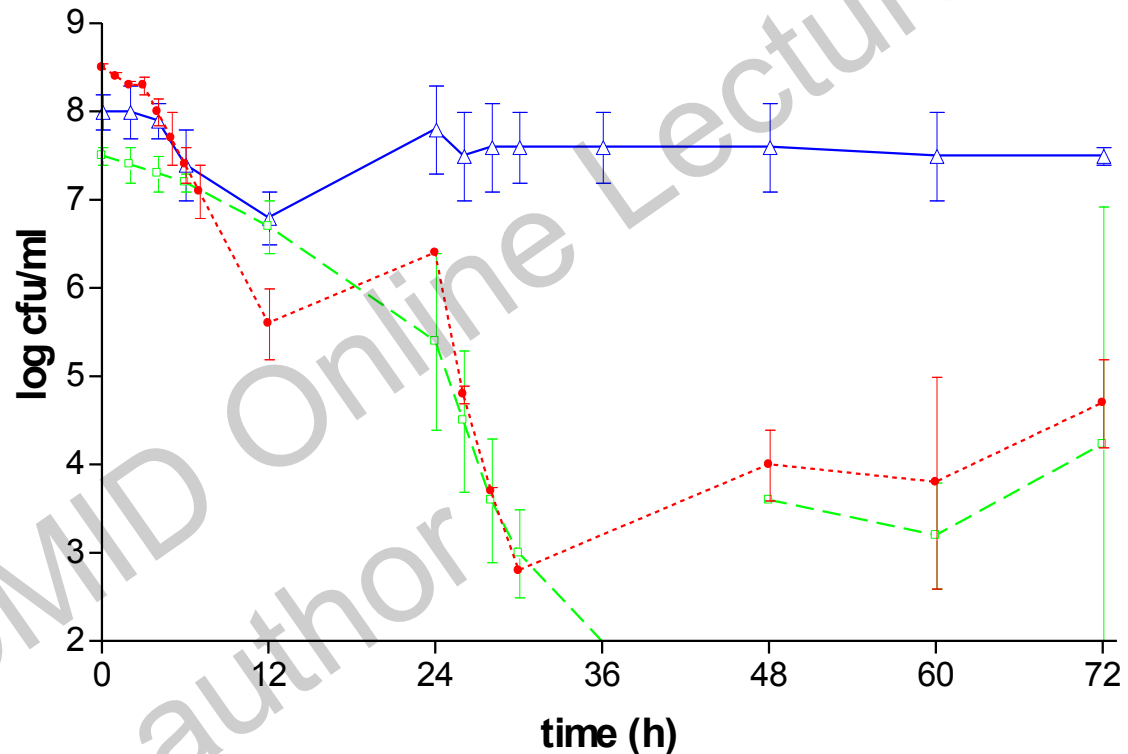
# Descriptive studies - Anaerobes

Figure 1 Antibacterial effect of moxifloxacin at AUC/MIC ratios in the range 9-216 against *B fragilis*



# Activity against defined resistance mechanisms - MIC does not drive outcome (gemifloxacin)

(gemifloxacin)



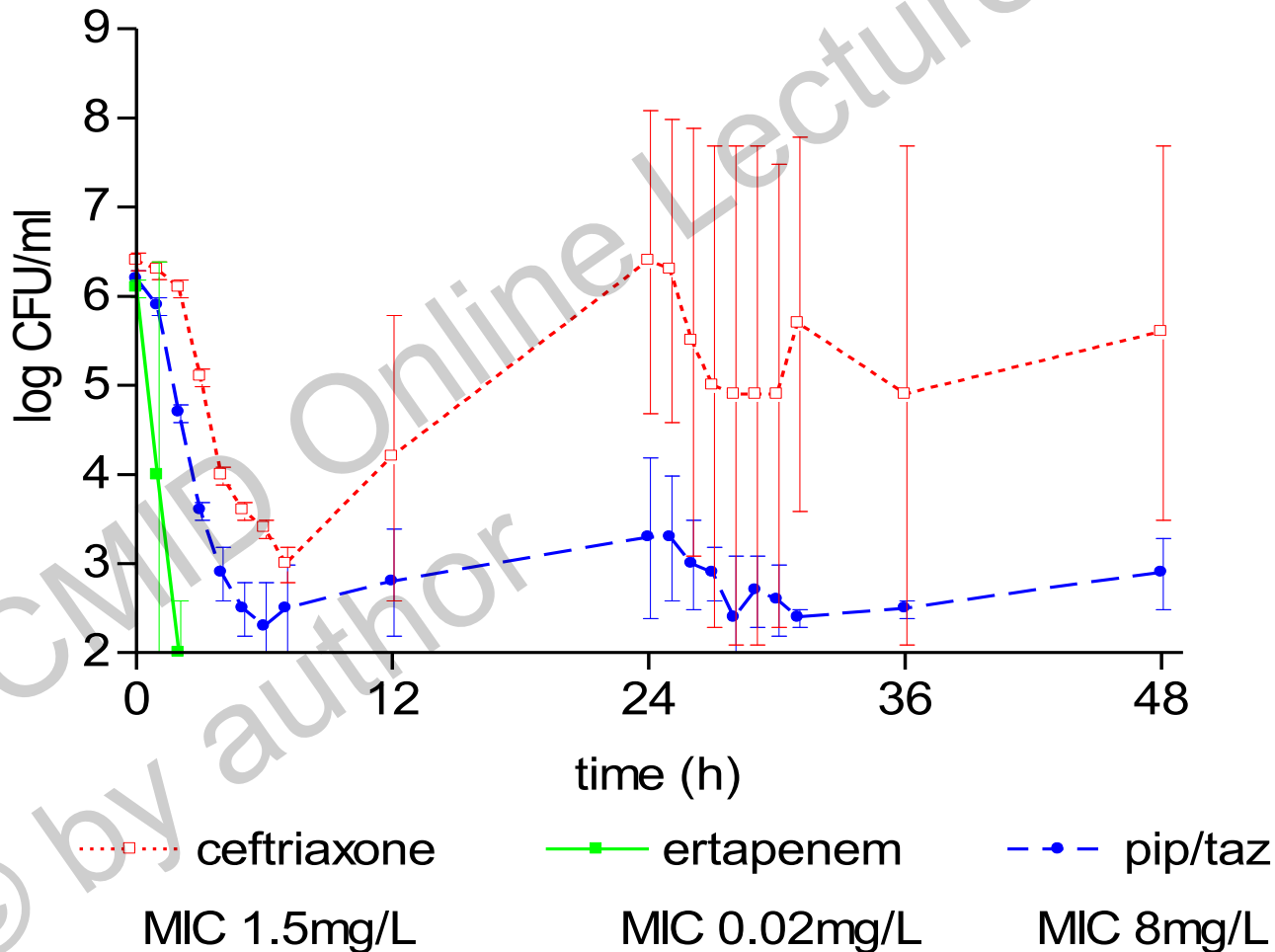
—□— MIC 0.06mg/L WT (SMH21810)

—△— MIC 0.06 mg/L 1st step parC mutation (SMH21813)

—●— MIC 0.06mg/L efflux (SMH21850)

# Activity against defined resistances - MIC drives outcome (ESBL)

d) E.coli 33212

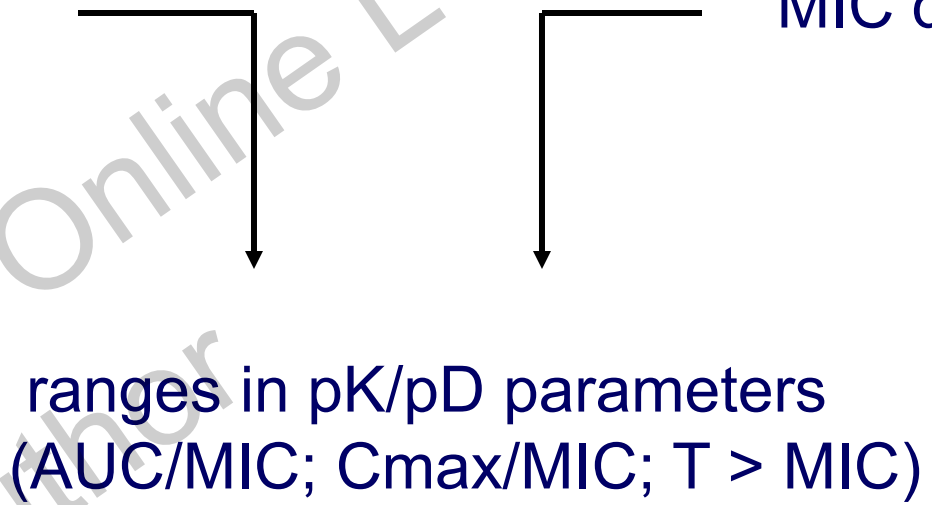




# Producing Variability in pK/pD Parameters

dose escalation  
dose fractionation

MIC differences

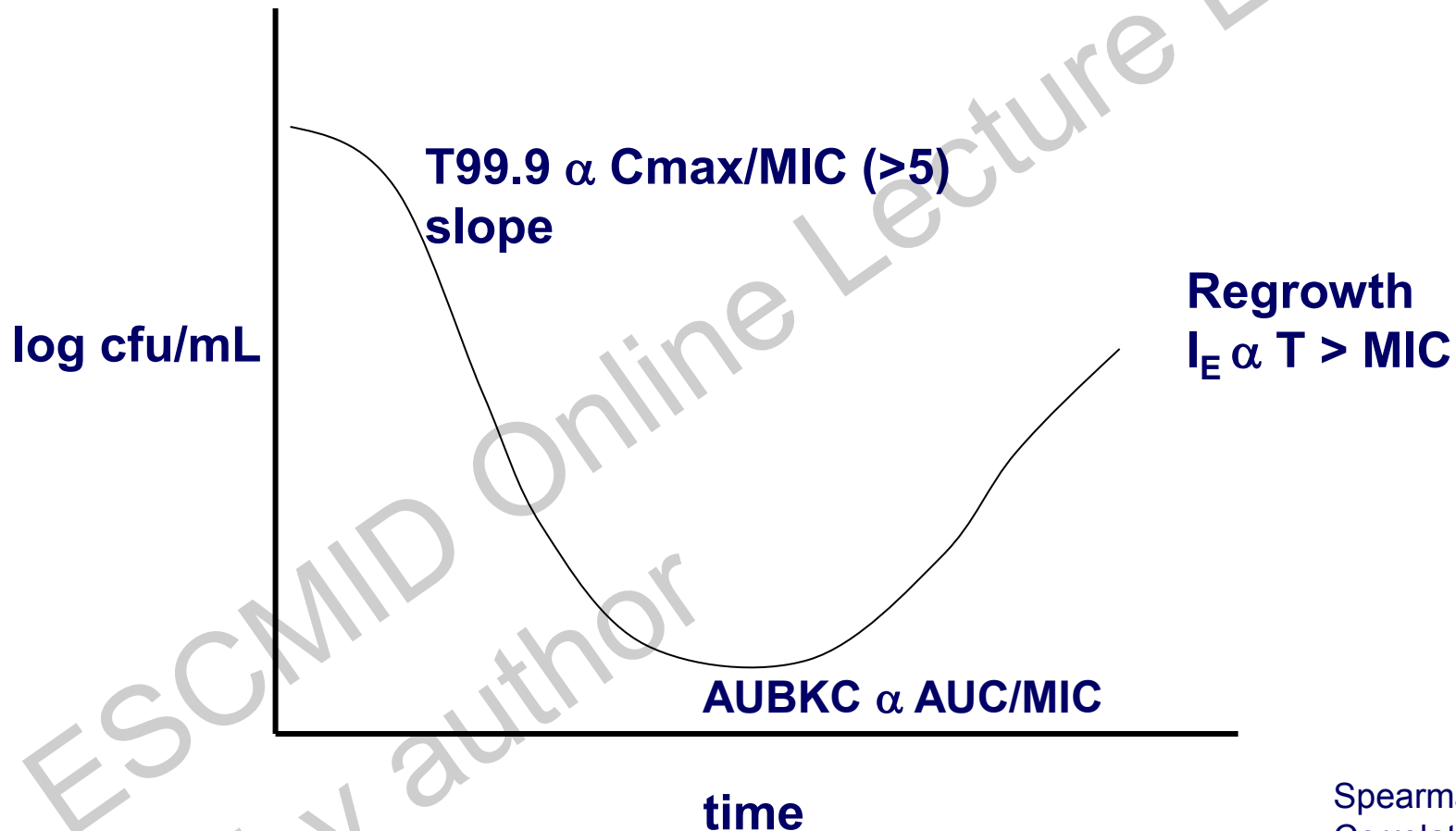


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# Dose fractionation studies – pK/pD targets



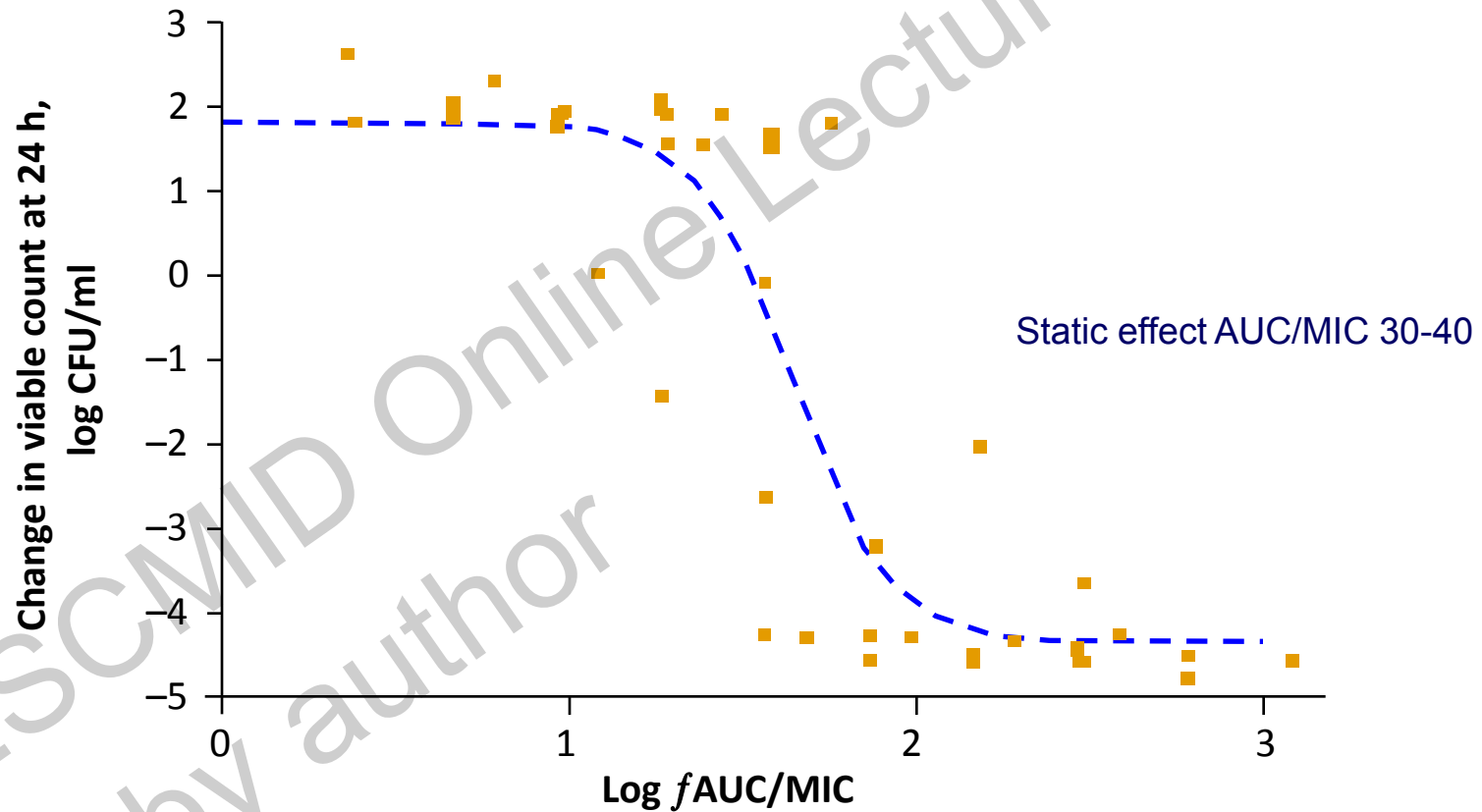
# Effect of antibacterial effect measure on dominant pK/pD parameter e.g. fluoroquinolones



	Spearman rank Correlation (95% CI)
AUC/MIC v Cmax/MIC	0.77 (0.42 - 0.92)
AUC/MIC v T>MIC	0.87 (0.60 - 0.96)
Cmax/MIC v T>MIC	0.42 (0.14 - 0.77)

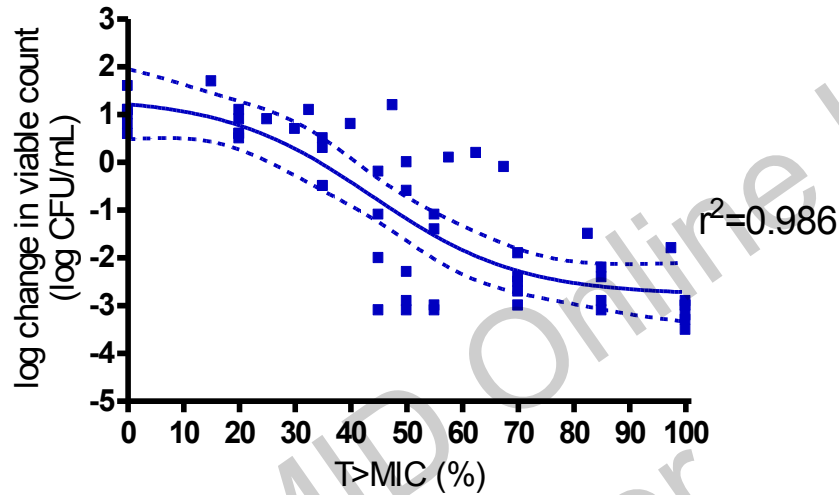
# Exposure – response relationship

Daptomycin  $fAUC/MIC$  ratio and ABE for *S. aureus*



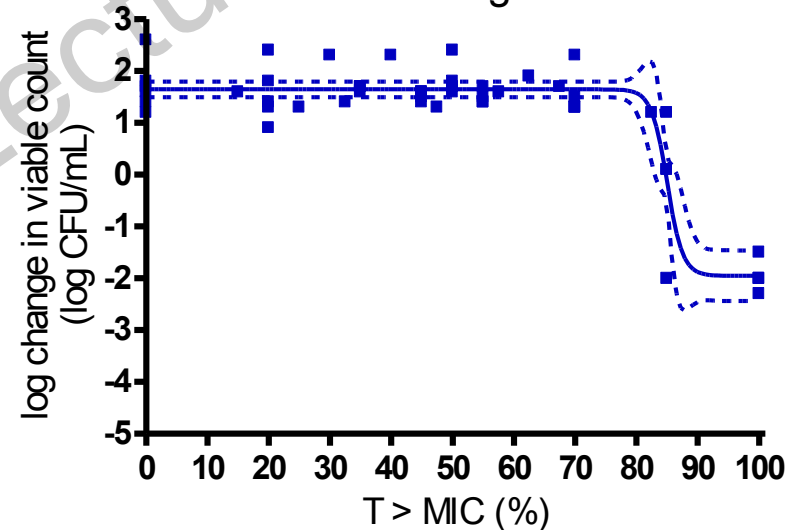
# Effect of exposure

Piperacillin-tazobactam fT>MIC relationship to antibacterial effect at 24 h for *P. aeruginosa*



Static effect, T > MIC 40%

fT > MIC relationship to antibacterial effect at 72h for *P. aeruginosa*



Static effect, T > MIC > 80%

# Effect of bacterial species: Ceftriaxone

Species	n	static effect	T>MIC % for 24hr		
			-1 log drop	-2 log drop	-3 log drop
<i>S.aureus</i>					
MSSA	4	27 ± 10	31±12	28 ± 6	33 ± 9
MRSA	4	22 ± 9	25 ± 7	27 ± 6	32 ± 8
<i>E.coli</i>	4	35 ± 6	37 ± 7	38 ± 8	40 ± 10
<i>K.pneumoniae</i>	5	36 ± 8	44 ± 9	52 ± 13	85 ± 15
<i>Citrobacter sp</i>	2	47	49	52	54
<i>Serratia sp</i>	1	65	66	70	>100
<i>P.mirabilis</i> *	3	39 ± 15	40 ± 26	41 ± 25	61 ± 15

\*5 - >75

Noel et al, 2012, Bowker et al 2013

# Inoculum - Vancomycin

UK MRSA 15 and 16

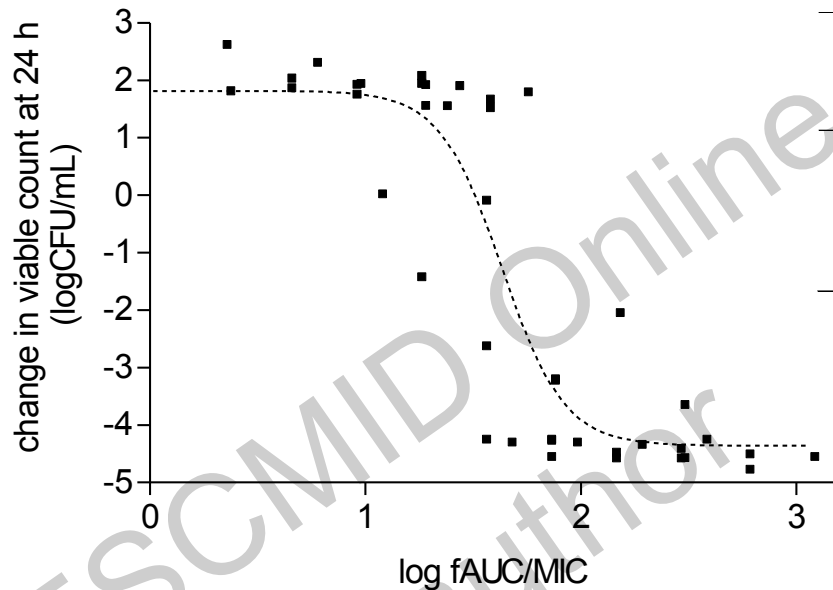
antibacterial effect log drop at 24h	initial inoculum			
	10 <sup>6</sup> CFU/ml		10 <sup>8</sup> CFU/ml	
	dose g/d	free drug AUC/MIC	dose g/d	free drug AUC/MIC
static	0.05	17	-	-
-1 log <sub>10</sub>	0.1	33	0.67	213
-2 log <sub>10</sub>	0.4	127	0.9	294
-3 log <sub>10</sub>	0.95	303	>4	>1274
-4 log <sub>10</sub>	>4	>1274	>4	>1274

MacGowan et al, 2008

# Effect of drug exposure

## Daptomycin and *S.aureus*

fAUC/MIC ratio relationship to change in viable count at 24hrs - *S aureus*



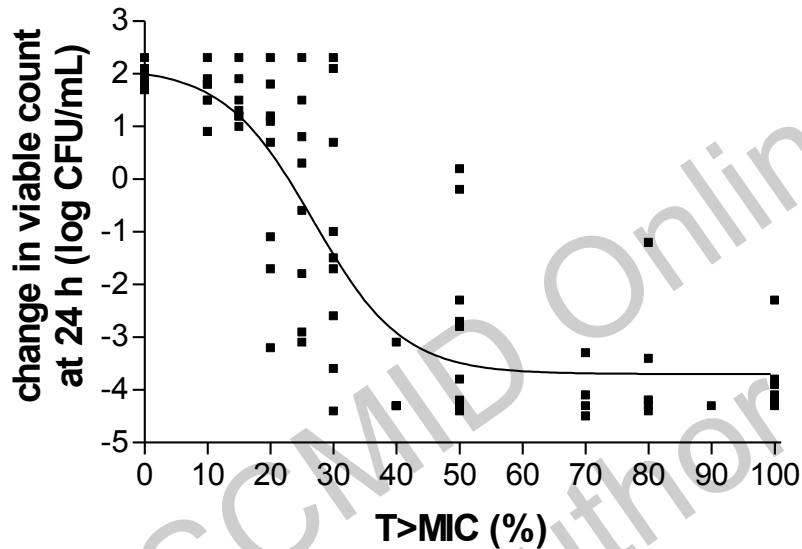
fAUC/MIC	number of experiments	% exps with growth on MICx4 plates	bacterial count on MICx4 plates
0.5-10	11	73(8)	4.3 ± 1.3
10-30	5	60(3)	3.9 ± 0.8
30-40	3	67(2)	3.6
>40	6	17(1)	4.7



# Effect of drug exposure

Ceftaroline and *S.aureus*

fT>MIC relationship to antibacterial effect and changes in population profile



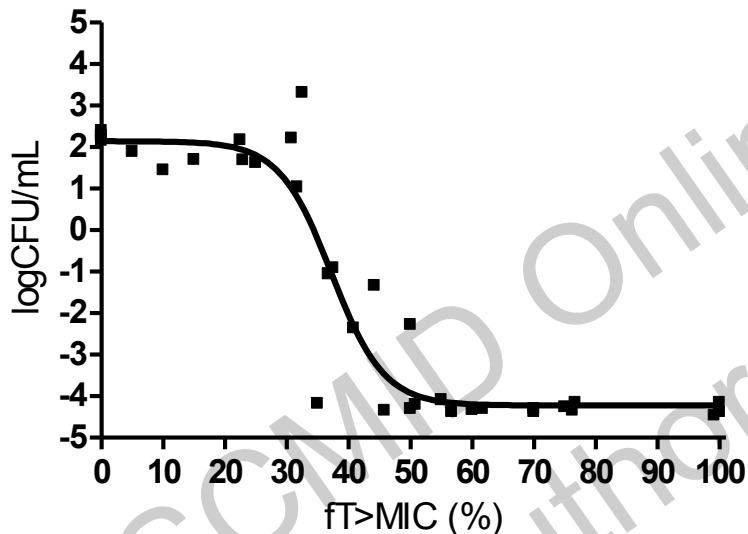
fT>MIC (%)	Number of experiments	% Exps with growth on MICx2 plates	Bacterial count on MICx2 plates
1-10	7	71(5)	6.4 ± 2.3
15-25	15	73(11)	5.9 ± 2.2
25-40	16	69(11)	6.3 ± 1.2
40-50	5	60 (3)	3.4 ± 1.6
≥50	29	7(2)	2.5

# Effect of drug exposure:

Ceftaroline and *E.coli*

fT>MIC relationship to antibacterial effect and changes in population profiles

Relationship between fT>MIC and antibacterial effect for *E.coli* at 24h



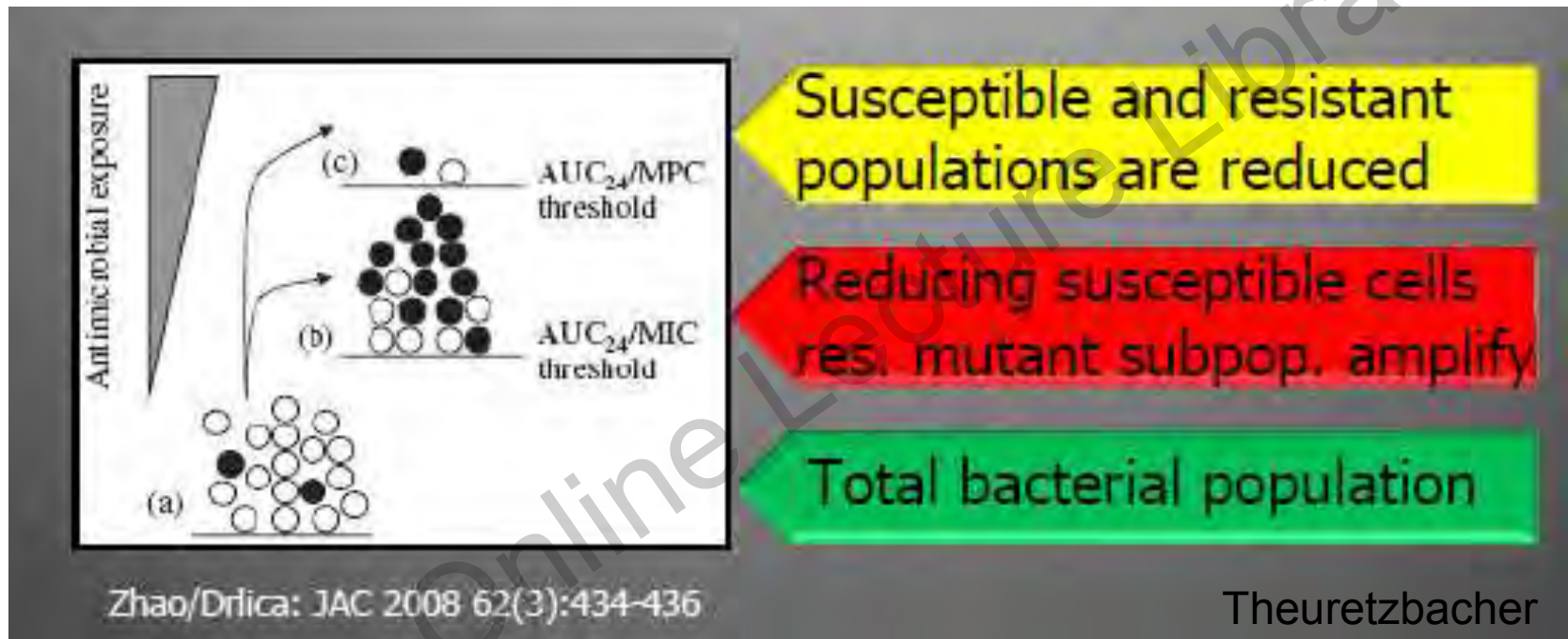
fT>MIC	Number of experiment	%Experiment with growth on MICx4 plate	Bacterial count on MICx4 plates
1-10	2	50% (1)	2.9
11-20	3	67% (2)	3.2
21-30	3	100% (3)	4.8 ± 0.9
31-40	5	66% (3)	4.5 ± 1.2
41-50	5	20% (1)	3.4
51-60	4	25% (1)	3.4
≥61	9	0	-

# Role of in vitro models in resistance

## **Critical factors for selection of subpopulations**

- Pathogen species/strain - heterogeneity
- Growth of organism
- Pre existing resistance mechanism (which?) - mutation ?spontaneous ?frequency
- Mixtures of S and R strains, same species
- Inoculum high inocula ( $10^{7-8}$  cfu/mL) + 360mL volumes  $\Rightarrow$  large bacterial exposure  $10^{10-12}$  cfu/mL
- Drug exposure, any drug class - how much? how often? how long?
- Flexible drug exposure;  $\Rightarrow$  free drug concs associated with standard drug doses  
 $\Rightarrow$  dose ranging and fractionation
- Combination therapy
- Clinical correlates - S.aureus & vancomycin

# Prevention of resistant sub populations

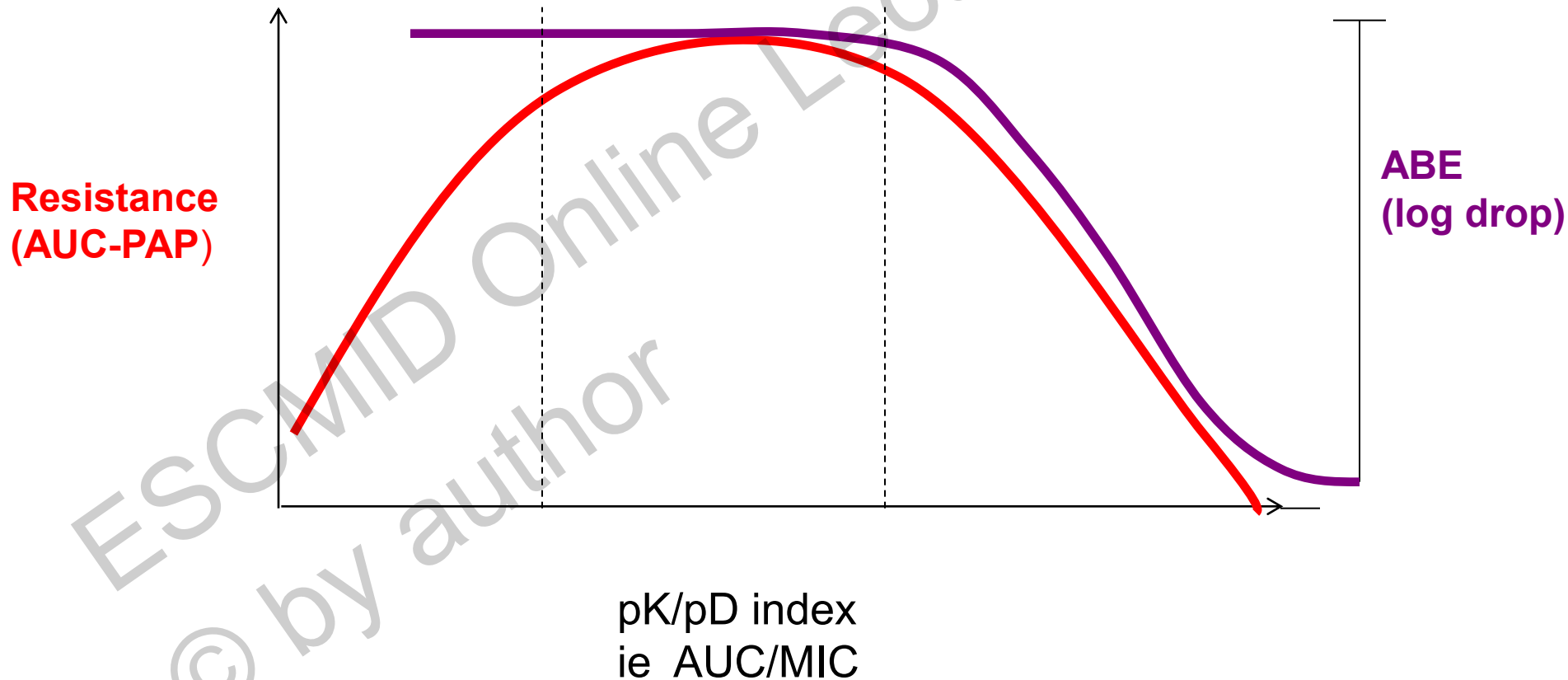


## End points to measure resistance

- increase in MIC (agar dilution, gradient strip, etc.)
- population analysis profiles (PAPs)
- absolute bacterial count on plates containing antibiotic (MIC multiple; absolute concentration)

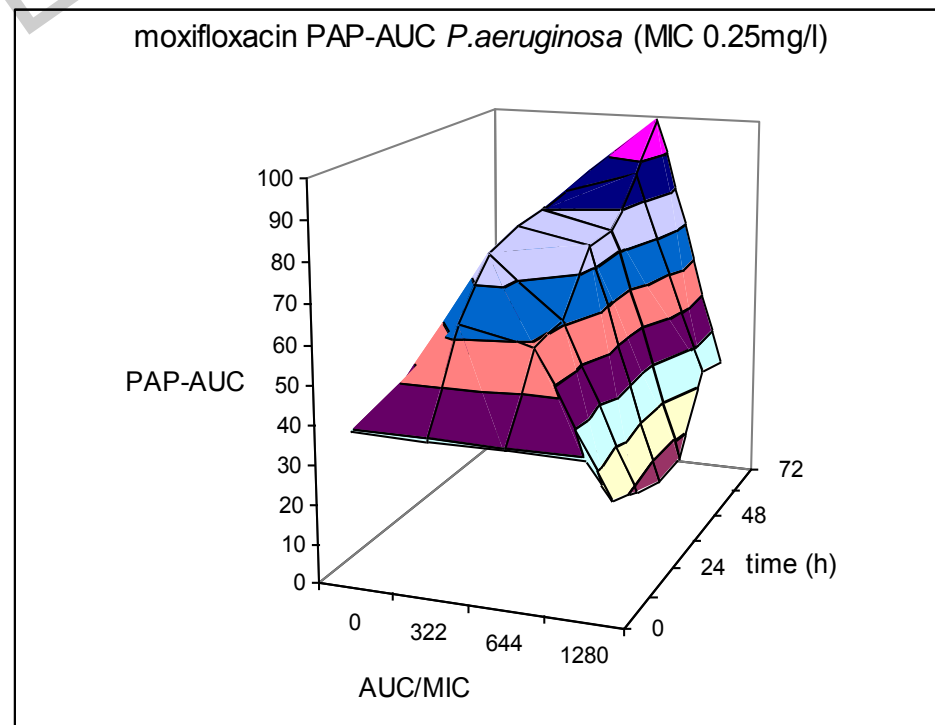
# Emergence of resistance

No clearance	No clearance	clearance
No EOR	EOR	No EOR



# Effect of drug exposure: emergence of resistance

dose (mg)/ MIC (mg/L)	<u>AUC</u> <u>MIC</u>	PAP-AUC (log cfu/ml.mg/L) at 72hr
0/0.25 & 1.0	0	34 ± 17 (pre exposure)
400 12hr/1.0 800 24h/1.0	320 320	95 88 ± 13
200 24h/0.25	322	84 ± 1
400 24h/0.25	664	99 ± 2
400 12h/0.25 800 12h/0.25	1280 1280	34 ± 29 28 ± 14



# Effect of inoculum and mechanism of resistance

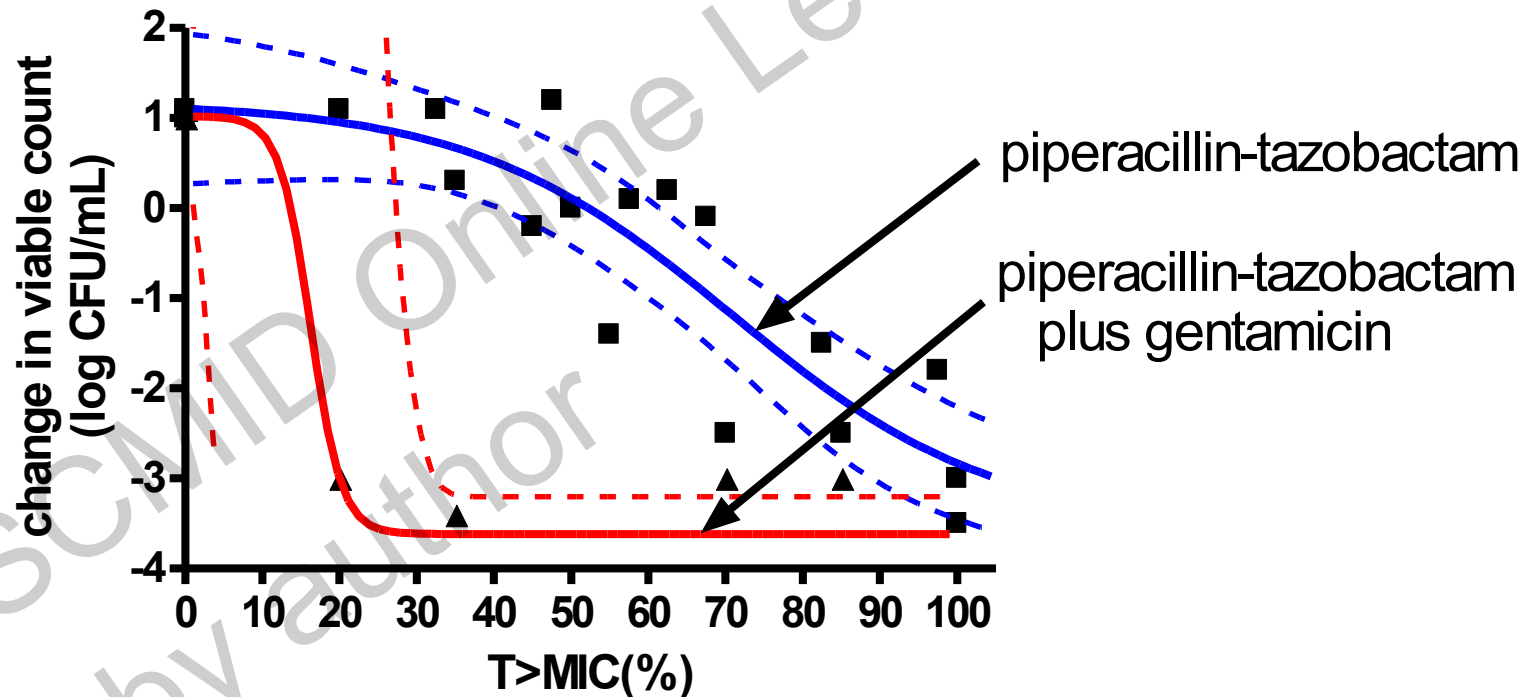
*S.pneumoniae* and fluoroquinolones: Emergence of resistance after 96h on MICx4 plates

Strains/dose simulations	fAUC/MIC	innoculum (CFU/ml)			
		10 <sup>6</sup> positive experiments	counts	10 <sup>8</sup> positive experiments	counts
<b>Strain 21843 wild type</b>					
levo 1g/day	56	0/3	-	0/3	-
levo 0.75g/day	42	0/3	-	3/3	2.8±3.9
moxi 0.4g/day	63	0/3	-	0/10	-
<b>Strain 21850 efflux</b>					
levo 1g/day	112	2/3	4.5	1/3	7.2
levo 0.75g/day	84	0/5	-	4/4	7.9±0.2
moxi 0.4g/day	63	0/5	-	0/4	
<b>Strain 21812 (par C)</b>					
levo 1g/day	56	3/3	7.0±0.1	3/3	3.5±0.4
levo 0.75g/day	42	2/2	4.1	1/4	2.8
moxi 0.4g/day	48	3/3	6.9±1.5	3/7	4.4±2.9

# Effect of combination antimicrobial chemotherapy

Piperacillin-tazobactam – dose ranging for  $fT > MIC$  0-100%, plus gentamicin 12mg/L.h

**P.aeruginosa 46042  $T > MIC$  for piperacillin-tazobactam alone and plus gentamicin (AUC 12mg/l.h)**





# Correlation with in vivo models

	in vitro $fT > MIC$			in vivo $fT > MIC$		
	static	-1log drop	-2log drop	static	-1log drop	-2log drop
<i>P.aeruginosa</i>						
doripenem	25 ± 11	30 ± 11	35 ± 11	29 ± 5	-	44.7 ± 7
ceftolozane	24.9	26.6	31.2	24.0 ± 3.3	31.5 ± 3.9	-
<i>E.coli</i>						
ceftolozane	27.8 ± 5.6	33 ± 5.6	39.4 ± 8.9	28	32.6	-
ceftaroline	35 ± 6.3	36.8 ± 7.1	38.3 ± 8.3	28 ± 9	41 ± 11	54.3 ± 21
<i>S.aureus</i>						
ceftaroline	24.5 ± 8.9	27.8 ± 9.5	27.7 ± 5.7	26 ± 8	33 ± 9	45 ± 13

# Conclusions

- Simple descriptive effects of simulated doses
- Allow identification of dominant pD index and size for effect
- pK/pD provides a rational framework for deciding drug doses, dosing regimens target, pathogens and clinical breakpoints
- pK/pD allows for prediction of therapeutic effect and provides a rationale to explain drug performance pK/pD may allow for performance of small therapeutic trials to establish efficacy (single RCT per indication)
- pK/pD can justify drug use in rare or difficult pathogens/indications (getting clinical cases very difficult)
- pK/pD of antibacterials can be applied to antivirals/antifungals
- a knowledge of a drugs pK/pD is essential in terms of drug licensing