

O267

Oral Session

PCR and other molecular tests directly on blood: what is new?

FIELD EVALUATION OF THE NEW QUICKFISH™ ASSAYS FOR THE RAPID DIAGNOSIS OF SEPSIS

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Objectives. The rapid identification of pathogens responsible for sepsis is a major goal to administer a targeted therapy and to reduce patient mortality rates. In our laboratory about 90 cases of sepsis per month were detected, mainly caused by staphylococci and Gram negative bacteria.

Peptidenucleic acid fluorescence *in situ* hybridization (PNA FISH) is a molecular diagnostic tool for the rapid identification (about 2 hours) of pathogens directly from positive blood cultures (BC), currently used in our laboratory and, on the basis of the available panel used, able to distinguish between *Staphylococcus aureus* (SA) and other coagulase-negative staphylococci (CNS), between *Enterococcus faecalis* (EF) and other enterococci (OE), between *Escherichia coli* (EC), *Klebsiella pneumoniae* (KP) and *Pseudomonas aeruginosa* (PA), between *Candida albicans*/C. *parapsilosis*, *C. tropicalis* and *C. glabrata*/C. *krusei*.

A recent evolution of PNA FISH, the QuickFISH™ assay (AdvanDx) uses a faster (about 20 minutes) and simpler protocol at present limited to bacteria. The aim of this study was to evaluate the SA/CNS and the EC/KP/PA QuickFISH™ assays on positive blood culture detected in our laboratory in the first month of use, as compared to conventional cultures and subsequent MALDI-TOF MS identification methods.

Methods. In a period of 1 month 109 cases of sepsis were detected, on a total of 860 analysed samples belonging to 373 patients. On the basis of the Gram stain microscopy result, from 60 positive blood culture bottles (Bactec FX, BD) (1 bottle per patient) the QuickFISH™ assays were applied selecting the appropriate panel. These samples were then cultured using standard techniques and the identification of grown microorganisms were performed by MALDI-TOF MS and biochemical identifications by traditional assays when needed for discordant results.

Results. On all 60 positive BC, QuickFISH™ showed a 100% agreement with the identification by other methods (6 SA, 30 CNS, 2 SA+CNS, 12 EC, 4 KP, 2 PA and 4 negative). The QuickFISH™ assay provided species identification in average 1 day earlier in the case of monomicrobial infections and 2 days for polymicrobial infections than the other methods.

Conclusion. In the first month of use QuickFISH™ demonstrated to be an excellent tool for the rapid identification of main pathogens directly on blood culture, improving the PNA FISH method in terms of rapidity and ease of handling and reducing significantly the reporting time as compared to other methods (MALDI-TOF MS performed on conventional cultures) with a positive impact for the appropriate management of the patient and the administration of a targeted therapy.