eP303
ePoster Viewing
Basic science: biofilm pathophysiology

INVESTIGATION OF THE EFFECT OF DIFFERENT TEMPERATURE AND ACIDIC CONDITIONS ON BIOFILM STRUCTURE OF SALMONELLA ENTERICA SEROTYPE VIRCHOW BY FTIR SPECTROSCOPY

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Objectives: We aimed to determine the effect of environmental stress conditions (temperature and pH) on biofilm structure of *Salmonella enterica* serotype Virchow originated from Turkey.

Methods: In this study, the influence of environmental factors including temperature (20 °C, 25 °C, 30 °C) and pH (5.2, 5.9, 6.6) were determined on biofilm formation by *S. enterica* serotype Virchow which are isolated from food samples. To observe the pellicle formation of the isolates, LB broth without salt (Wo/NaCl) was used and the strains were incubated at different temperatures for 10 days. Afterwards, pellicles were analyzed visually and by Fourier transform infrared (FTIR) spectroscopy using attenuated total reflectance (ATR) method. For FTIR analyses, after 10-day incubation, pellicles were transferred to micro-centrifuge tubes under sterile conditions and then all pellicles were lyophilized by freeze-drying. FTIR spectra were recorded in the mid-IR region using Tensor 27 FTIR (Bruker) and spectral analyses were performed using OPUS 5.5 software (Bruker). Statistical significance was tested by ANOVA. Additionally, we used hierarchical cluster analysis as a multivariate analysis.

Results: After incubation at 20, 25 and 30^{o}C temperature, biofilm formation in standing LB broth was visualized after 10 days as a floating pellicle at the air–broth interface that totally blocked the surface of the culture. Pellicles which were prepared at 20 and 25^{o}C were not dispersed by shaking while they easily disrupted at 30°C. We observed 16 peaks corresponding different biomolecules such as protein, lipid, carbohydrate and nucleic acid in the FTIR spectra of samples (Figure 1). According to our results, the amount of carbohydrate, protein, lipid and nucleic acid was significantly decreased with increasing temperature (especially between 20 and 30 ^{o}C). We did not observe considerable alterations related to pH changes within temperature groups. Our hierarchical cluster analysis supported these results and separated groups according to the temperature.

Conclusions: In our samples, we assigned the carbohydrate peak as cellulose, which is one of the main components of biofilm structure. Taking into consideration our morphological and molecular results, we concluded that cellulose has an important role in biofilm formation and its stability. Additionally, decrease in protein level may be associated with curli fimbria protein. Alterations in lipids may reflect changes in membrane structure of bacterial cells during biofilm formation. Also our univariate and multivariate analyses showed that temperature has a more significant effect than pH on biofilm formation and stability.

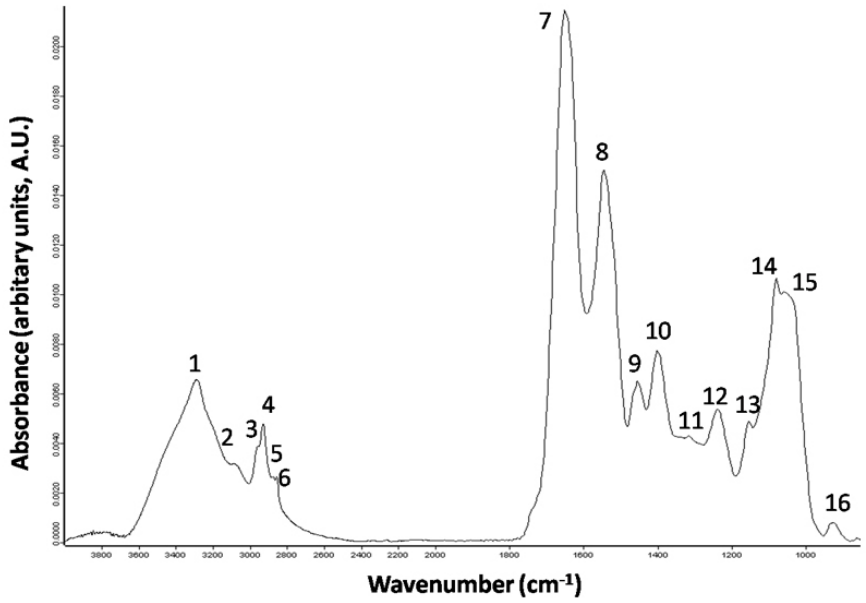

Figure. Representative FTIR absorbance spectrum of biofilm sample between 4500–850 cm^{-1} obtained by averaging all the spectra used in the analyses.