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Genomics of *Tropheryma whippelii*, the agent of Whipple's disease

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Tropheryma whippelii

- 1907 First description of Whipple's disease (Metabolic trouble)
- 1991 Identification of the agent of Whipple's disease
A rare bacterium causing a rare chronic infection
- 2000 First establishment of a strain of *T. whippelii*
- 2003 2 full genome sequences of 2 different strains of *T. whippelii*
 1 from the index strain TWIST (Cardiac valve from a Canadian patient with endocarditis)
 1 from another strain TW08/27 (CSF from a German patient with a relapse of classic WD)

Raoult D, Ogata H, Audic S, et al. *T. whippelii*/twist: a human pathogenic actinobacteria with a reduced genome. *Genome Res* 2003;13:1800-9.
 Bentley SD, Malwald M, Murphy LD, et al. Sequencing and analysis of the genome of the Whipple's disease bacterium *T. whippelii*. *Lancet* 2003;361:637-44.

Main characteristics of *T. whippelii* genome

- *Tw* is an Actinobacteria and presents a small genome slightly less than 1 Mbp (a 46% GC content) whereas other actinomycete genomes are from approximately 2.1–10.1 Mbp with usually a rich GC content.
- Several pathways are lacking:
These features are those of reductive genome evolution.
- *Tw* seems to have evolved to a host-dependent organism, relying on the uptake of nutrients from outside sources.
- *Tw* carries only a few genes that regulate transcription.
- The cell surface seems relatively abundant.

Predictions suggest that approximately 15% of all proteins are present at the cell surface.
Tw possesses many WSPs, which are surface proteins belonging to a family characterized by large size ranges and architectures.

Genome analysis

Presence of repeated sequences

Development of the « repeat-PCR »

Fenollar F, Fournier PE, Robert C, Raoult D. Use of genome selected repeated sequences increases the sensitivity of PCR detection of *T. whippelii*. *J Clin Microbiol* 2004;42:401-3.

Several steps of the development of repeat-PCR

1. Specificity of *T. whippelii* repeat-DNA target *in silico* (Genbank)
2. Sensitivity of repeat-PCR *in vitro* versus classic PCR
3. Specificity of repeat-PCR *in vitro* (collection of bacteria)
4. Analysis of samples from patients with Whipple's disease and from a control group tested with classic PCR and repeat-PCR

Specimens from patients	Number	Positive using classic PCR	Positive using Repeat-PCR
Total	98	33 (33.6%)	54 (55%)

All the specimens from the control group were negative.

Repeat-PCR = Our current diagnostic tool
 2 PCR targeting 2 different repeated sequences should be positive to conclude to a positive assay

Genome = Information about the metabolism of the bacterium

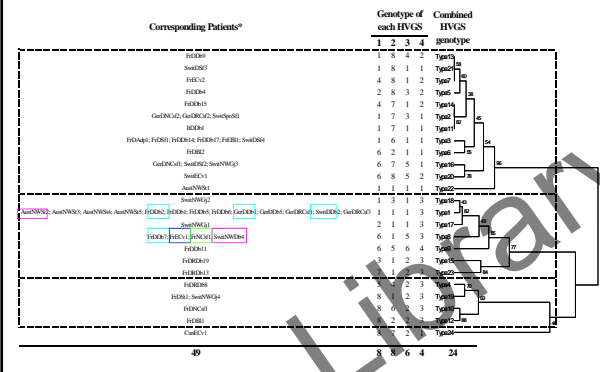
- Lack of numerous way for the synthesis of amino acids
- Development of a cell-free culture medium that provided all the missing amino acids
- Culture of *T. whippelii* on axenic medium

***T. whipplei* genotyping**

Li W, Fenollar F, Robin JM, Fournier PE, Fourie GE, Muller C, Moos V, Marth T, Altwegg M, Calligaris-Malbach RC, Schneider T, Biagi F, La Scala B, Raoult D. Genotyping reveals a wide heterogeneity of *T. whipplei*. *Microbiology* 2008;154:521-7.

- Selection of 4 highly variable sequences from the comparison of the 2 *T. whipplei* genomes available
- Study performed on 39 DNA extracts from 39 patients
 - 26 classic WD with digestive involvement
 - 6 relapses of classic WD (3 digestive and 3 neurologic)
 - 5 blood culture negative endocarditis
 - 1 encephalitis
 - 1 spondylodiscitis
- 10 DNA extracts from 10 asymptomatic carriers
- Origin of the strains: France, Italy, Germany, Austria, Switzerland, Canada
- Classification in 24 different genotypes
- High genetic variety of *T. whipplei*

Concatenation of the 4 highly variable sequences (UPGMA tree)



Concatenation of the 4 highly variable sequences (UPGMA tree)

High genetic variety of *T. whipplei*

In this tree: 61 genotypes from 198 specimens

- Carriers
- *T. whipplei* chronic infections (classic and localized)

Cases of gastroenteritis

Genotype* : Same genotype detected between the family members of a same family

Sene: Senegal



***T. whipplei* prevalence strongly supports human transmission in homeless shelters**

Alta A, Brauqui P, Badaga S, Benkouiten S, Romanov P, Raoult D, Fenollar F. *Trachyroma whipplei* prevalence strongly supports human transmission in homeless shelters. *Int J Infect Dis* 2013;17:667-8.



Year	3	85
2010	1	1*
2011	4	3*
Genotypes previously identified in France	33/125 (26.4%)	No

*Both sputa and saliva specimens from the same individual present the same genotype.

Year	3	85	84	27
2010	1	3	1	1
2011	4	3	1	1
Genotypes previously identified in France	33/125 (26.4%)	No	No	2/125 (1.6%)

- Suspicion of small outbreaks in this population (genotype 3 in Shelter 1 and 85 in Shelter 2 in 2011).

Predominance of *T. whipplei* genotypes 1 and 3 in central Europe

Wetzstein N, Fenollar F, Buffet S, Moos V, Schneider T, Raoult D. *EID* 2013; 19:341-2.

- 191 samples from central Europe. Genetic diversity by finding 72 different *T. whipplei* genotypes
- 28.8% of all samples showed a unique genotype

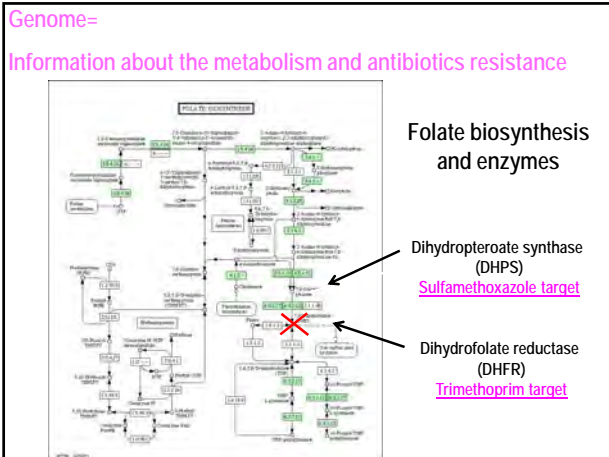


- Genotype 3: The most common in Europe (19.9%) and could be considered epidemic and specific to France, Switzerland and Italy
- Genotype 1: The second-most frequent genotype (15.2%) found all over central Europe (Germany: 46.2% and Austria: 80%)

Genome = Information about genes coding for antibiotics resistance Fluoroquinolones and DNA gyrase

	Susceptibility to fluoroquinolones	70	80	83	90	100
GyrA <i>E. coli</i> K12	S	ARVVDVIGKYHPHGD	AAVYDTLVVMAQ	PFSRLRYPLVDQ		
ParC <i>E. coli</i> K12	S	ARTVGDVIGKYHPHGD	SAC	AEAMVLMQ	PFSRYPLVDQ	
GyrA <i>T. whipplei</i>	R	ARVVDVVMGQPHPHGD	AA	ADALVRLVQ	PWAMRPLAQDQ	
ParC <i>T. whipplei</i>	R	ARVTEVMGKLPHPHGD	AA	IYDTLVRMSQ	DFTMR1PLIDGH	
GyrA <i>M. avium</i>	R	ARSVAETMGNYPHGD	AA	SIYDTLVVMAQ	PWLSRYPPLVDQ	
GyrA <i>M. leprae</i>	R	ARSVAETMGNYPHGD	AA	SIYDTLVVMAQ	PWLSRYPPLVDQ	
GyrA <i>M. smegmatis</i>	R	ARSVAETMGNYPHGD	AA	SIYDTLVVMAQ	PWLSRYPPLVDQ	
GyrA <i>M. intracellulare</i>	R	ARSVAETMGNYPHGD	AA	SIYDTLVVMAQ	PWLSRYPPLVDQ	
GyrA <i>M. tuberculosis</i>	R	ARSVAETMGNYPHGD	AA	SIYDTLVVMAQ	PWLSRYPPLVDQ	
GyrA <i>M. fortuitum</i>	S	ARSVAETMGNYPHGD	BS	SIYDTLVVMAQ	PWLSRYPPLVDQ	

- Confirmation with *In vitro* susceptibilities: Ofloxacin and ciprofloxacin are not effective *in vitro*



- Confirmation with *In vitro* susceptibilities:

Antibiotics	MICs (µg/ml)		
	Twist	Endo 5	Slow
Sulfamethoxazole/Trimethoprim	2/0.5	4/1	4/1
Sulfamethoxazole	0.5	1	1
Trimethoprim	64	100	64
Sulfadiazine	0.5		

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